

Development Support Document Proposed, August 2007

## 1,3-Butadiene

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TEXAS COMMISSION ON ENVIRONMENTAL QUALITY

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#### **Chapter 1 Summary Tables and Figure** 1

2 Table 1 provides a summary of health- and welfare-based values based on an acute and chronic evaluation

3 of 1,3-butadiene (BD). Chapters 3 and 4 of the Development Support Document (DSD) provide

4 information on the development of the acute and chronic values, respectively. Table 2 provides summary

5 information on BD's physical/chemical data.

6

Table 1. Health- and Welfare	-Based Values	
Short-Term ESL for Air Permit	Reviews: 240 μg/m <sup>3</sup> (110 p	opb)
Short-Term Values	Values	Notes
Acute ReV <sup>acute</sup> ESL	800 μg/m <sup>3</sup> (360 ppb) 240 μg/m <sup>3</sup> (110 ppb)	<b>Critical Effect:</b> Maternal toxicity; reduction in weight gain in pregnant CD-1 mice
[1 h] <sup>acute</sup> ESL <sub>odor</sub> [1 h]	510 μg/m <sup>3</sup> (230 ppb) <sup>1</sup>	50% detection threshold, mild aromatic odor
acuteESL <sub>veg</sub>		Concentrations producing vegetative effects were significantly above other ESLs
Long-Term ESL for Air Permit	<b>Reviews: 10 μg/m<sup>3</sup> (4.5 ppb</b>	))
Long-Term Values	Values	Notes
Chronic ReV	$33 \ \mu g/m^3 (15 \ ppb)^1$	<b>Critical Effect:</b> Reproductive toxicity: ovarian atrophy
<sup>chronic</sup> ESL <sub>nonlinear(nc)</sub>	10 µg/m <sup>3</sup> (4.5 ppb)	in B6C3F1 mice
chronic ESLlinear(c)	$62 \ \mu g/m^3 (28 \ ppb)^2$	Cancer Endpoint:
		Leukemia in occupational exposure study of styrene-butadiene synthetic rubber production workers
<sup>chronic</sup> ESL <sub>veg</sub>		No data found

7

<sup>1</sup> Screening value for air monitoring data.

<sup>2</sup> Based on unit risk factor (URF) =  $0.00016/\text{mg/m}^3(0.00036/\text{ppm})$ 

8 9

10

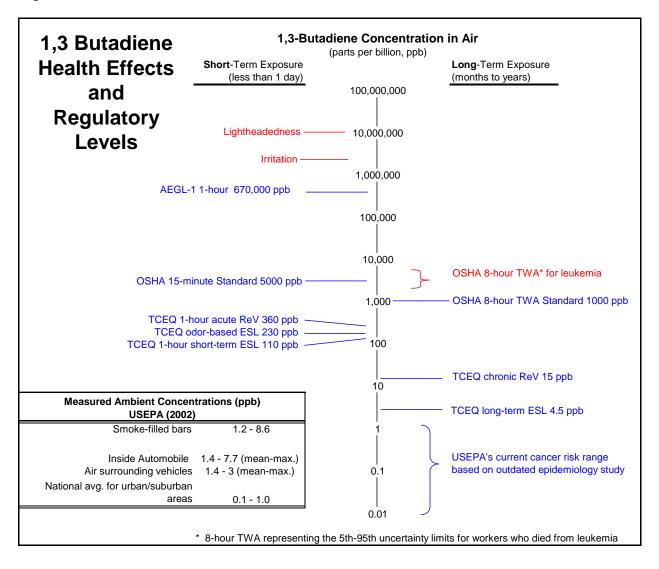
11

Abbreviations used: ppb, parts per billion;  $\mu g/m^3$ , micrograms per cubic meter; h, hour; ESL, Effects Screening Levels; ReV, Reference Value; <sup>acute</sup>ESL, acute health-based ESL; <sup>acute</sup>ESL<sub>odor</sub>, acute odor-based ESL; <sup>acute</sup>ESL<sub>veg</sub>, acute vegetation-based ESL; <sup>chronic</sup>ESL<sub>linear(c)</sub>, chronic health-based ESL for linear dose-response cancer effect; <sup>chronic</sup>ESL<sub>nonlinear(nc)</sub>, chronic health-based ESL for nonlinear dose-response noncancer effects; and <sup>chronic</sup>ESL<sub>veg</sub>, chronic vegetation-based ESL 12

13

1

Parameter	Value	Reference	
Iolecular Formula	C <sub>4</sub> H <sub>6</sub> or H <sub>2</sub> C:CHHC:CH <sub>2</sub>	Lewis 1993	
nemical Structure		ChemIDplus Lite	
	H <sub>2</sub> C		
Iolecular Weight	54.1	TRRP 2006	
ysical State	gas/organic	TRRP 2006	
olor	colorless	Lewis 1993	
lor	mild aromatic odor	ACGIH 2001	
S Registry Number	106-99-0	TRRP 2006	
nonyms	vinylethylene; erythrene; bivinyl; divinyl; biethylene; pyrrolylene; a,g- butadiene	Lewis 1993 NTP 1993	
blubility in water	735 mg/L	TRRP 2006	
g K <sub>ow</sub>	2.03	TRRP 2006	
oor Pressure	2,100 mm Hg at 20 <sup>0</sup> C	TRRP 2006	
por Density (air $= 1$ )	1.87	Lewis 1992	
nsity (water = 1)	0.6211 (liquid at 20 <sup>0</sup> C)	Lewis 1993	
elting Point	-113°C	Lewis 1992	
ling Point	<sup>-</sup> 4.41 ° C	Lewis 1993	
onversion Factors	1 $\mu$ g/m <sup>3</sup> = 0.45 ppb @ 25°C 1 ppb = 2.21 $\mu$ g/m <sup>3</sup>	NTP 1993	



8

Abbreviations used: BD, 1,3-butadiene; TCEQ, Texas Commission on Environmental Quality; TWA,
 Time-Weighted Average; ESL, Effects Screening Level; ReV, Reference Value; OSHA, Occupational
 Safety and Health Administration; USEPA, United State Environmental Protection Agency; and AEGL-

Figure 1. BD Health Effects and Regulatory Levels. This figure compares BD's acute toxicity

values (acute ReV, odor-based ESL, and health-based, short-term ESL) and chronic toxicity values

based on an outdated epidemiology study, OSHA's occupational values, and the AEGL-1 value.

(chronic ReV and long-term ESL) found in Table 1 to USEPA's current acceptable cancer risk range

12 1, Acute Exposure Guideline Levels-1.

13

14

## 1 Chapter 2 Major Sources or Uses

2 BD is used as an intermediate in the production of polymers, elastomers, and other chemicals. Its major

- 3 uses are in the manufacture of styrene-butadiene rubber (SBR) (synthetic rubber) and thermoplastic
- 4 resins. Elastomers of BD are used in the manufacture of tires, footwear, sponges, hoses and piping,
- 5 luggage, packaging, and a variety of other molded products. In addition, BD is used as an intermediate to
- produce a variety of industrial chemicals, including the fungicides captan and captfol. The primary way
   that BD is released into the environment is via emissions from gasoline- and diesel-powered vehicles and
- 8 equipment. Lesser releases occur from the combustion of other fossil fuels and biomass. Minor releases
- 9 occur in production processes, tobacco smoke, gasoline vapors, and vapors from the burning of plastics as
- 10 well as rubber (Miller 1978; USEPA 2002). United States Environmental Protection Agency's (USEPA)
- 11 (2001) National-Scale Air Toxics Assessment of emissions from the 1996 National Toxics Inventory
- 12 indicates that statewide BD emissions from mobile sources (onroad and nonroad) accounted for
- 13 approximately 54% of the National Toxics Inventory BD emissions in Texas, with major facility sources
- 14 and area/other sources (e.g., smaller facilities) comprising the remainder of 46%.

## 15 Chapter 3 Acute Evaluation

16 3.1 Health-Based Acute ReV and <sup>acute</sup>ESL

## 17 3.1.1 Physical/Chemical Properties and Key Studies

## 18 3.1.1.1 Physical/Chemical Properties

- BD is a highly volatile, colorless gas with a mildly aromatic odor. The main chemical and physical
- 20 properties of BD are summarized in Table 2. It is soluble in ethanol, diethyl ether, and organic solvents,
- 21 and only slightly soluble in water.

## 22 3.1.1.2 Key Studies

- 23 This section is based on USEPA (2002) and AEGL (2005). Both of these sources state "The acute toxicity
- of BD is of low order." (USEPA 2002; AEGL 2005). A review of the scientific literature since 2002
- 25 indicates that a subchronic inhalation study in rats conducted by the American Chemistry Council (ACC
- 26 2003) is a new study that was not considered by USEPA (2002). Therefore, this study is discussed in
- 27 Section 3.1.1.2.2. Animal data show BD is a potential reproductive/developmental hazard to humans.
- 28 Since the reproductive/developmental effects of BD in rats and mice are among the effects observed at the
- 29 lowest exposure levels following acute inhalation exposure, the following sections focus on these health 20  $\frac{1}{2}$  affects Charter 5 of Harleh Agazan (LISERA 2002)
- 30 effects. Chapter 5 of *Health Assessment of 1,3-Butadiene* (USEPA 2002) provides a detailed discussion 31 on potential reproductive/developmental affects in humans and animals and AECL (2005) discusses
- 31 on potential reproductive/developmental effects in humans and animals, and AEGL (2005) discusses
- 32 other types of acute toxicity data.

## **33 3.1.1.2.1 Human Studies**

- 34 Mice are more susceptible to BD-induced reproductive/developmental effects than rats, whereas
- 35 reproductive/developmental effects in humans after exposure to BD have never been observed (Albertini
- 36 et al. 2007; USEPA 2002).
- 37

- 1 The health effects observed in humans occur at high concentrations and include the following: odor
- 2 perception (ACGIH 2001; Ruth 1986; and Nagata 2003); slight smarting of the eyes and difficulty in
- 3 focusing (Carpenter et al. 1944); and tingling sensation and dryness of the nose and throat (Larionov et al.
- 4 1934) (Table 3). A poorly reported study conducted by Ripp (1967) in human volunteers reported effects
- of olfactory perception at 4.0 milligram per cubic meter  $(mg/m^3)$  (1.8 parts per million (ppm)) and
- 6 sensitivity of the eye to light at  $3.9 \text{ mg/m}^3$  (1.7 ppm). There were no effects on the occurrence of an
- 7 electrocortical conditioned reflex at  $3 \text{ mg/m}^3$  (1.4 ppm).
- 8 9

Table 3. Acute Effects	of BD in Humans		
Study	Concentration	Subjective Symptoms	Differences Observed
	(Exposure Duration)		
Carpenter et al. 1944	2,000 ppm <sup>1</sup>	Slight smarting of the	
2 males	(7 hour (h))	eyes; difficulty in	
1-h lunch break		focusing	Results of tapping test
Nominal Concentrations	4,000 ppm	Slight smarting of the	and steadiness test – no
	(6 h)	eyes; difficulty in	differences
		focusing	
	8,000 ppm	No subjective	
	(8 h)	complaints <sup>2</sup>	
		r r	
Larionov et al. (1934)	1% (10,000 ppm)	Tingling sensation and	Slight increase in pulse
No details on number of	5 minute (min)	dryness of the nose and	rate. No effects on
subjects and gender	. ,	throat.	blood pressure or
			respiration

- <sup>1</sup> Difficulty in focusing was the basis of the AEGL-1 value. The 1-h AEGL-1 value of 670 ppm = 2,000
- 11 ppm divided by an intraspecies uncertainty factor of 3.
- 12
- $^{2}$  No subjective complaints because of slight anxiety of subjects concerning the possibility of an
- 14 explosion.

## 15 **3.1.1.2.2 Animal Studies**

### 16 3.1.1.2.2.1 Reproductive/Developmental Toxicity in Rats

17 In 1982, Hackett et al. (International Institute of Synthetic Rubber Producers (IISRP) 1982) conducted a

- reproductive/developmental study that included exposure of pregnant rats at 0, 200, 1,000, and 8,000 ppm
- 19 6 hours/day (h/day) on gestation day (GD) 6-15 and then sacrifice on GD 20. The most sensitive
- 20 endpoints were a significant decrease in maternal body weight gain on GD 6-9 and extragestational
- 21 weight gain (lowest observed adverse effect level (LOAEL) of 1,000 ppm and no observed adverse effect
- 22 level (NOAEL) of 200 ppm for both endpoints). Minor skeletal defects were found to be significantly
- elevated at the lowest concentration, and the percentage of fetuses with major skeletal defects was
- significantly elevated at 1,000 ppm and above. The incidence of marked-to-severe wavy ribs and the total
- number of abnormal ossifications and irregular ossification of the ribs were elevated at 8,000 ppm.
- 26
- 27 In 1987, Hackett et al. (1987a) repeated the IISRP (1982) study at slightly lower concentrations to
- 28 confirm the 1982 findings in rats and to compare the effects of similar BD exposures in mice (Hackett et

1 al. 1987b). The results of the Hackett et al. (1987b) study in mice are discussed in the next section. Pregnant rats (Hackett et al. 1987a) were exposed for 10-days via inhalation to 0, 40, 200, and 1,000 ppm 2 on GD 6-15 for 6 h/day (Hackett et al. 1987a). For rats, the most sensitive short-term endpoints were 3 4 decreases in maternal body weight gain on GD 6-11 and decreases in extragestational weight gain 5 (LOAEL of 1,000 ppm and NOAEL of 200 ppm for both endpoints). Effects from BD exposure for fetal 6 measures were not observed (i.e., no developmental toxicity was observed). 7 8 In 2003, a subchronic reproductive/developmental study in rats sponsored by the American Chemistry 9 Council was conducted by WIL Research Laboratories, Inc (ACC 2003). Since this study was not 10 available for USEPA's BD assessment (USEPA 2002), the major findings of the study are discussed below. The study was conducted using the following guidelines: 11 12 13 USEPA TSCA Good Laboratory Practice Standards; 14 The protocol met or exceeded applicable regulations of the Organisation for Economic • Cooperation and Development (OECD) Guideline for Testing of Chemicals, Guideline 421, 15 16 Reproduction/Development Toxicity Screening Test (July 27, 1995); and 17 Office of Prevention, Pesticides & Toxic Substances (USEPA) 870.3550 (July 2000) • 18 requirements. 19 20 This study was conducted to provide information on the potential adverse effects of BD on male and 21 female reproduction within the scope of a screening study. Assessments of gonadal function, mating 22 behavior, conception, gestation, parturition, lactation of the  $F_0$  generation, and the development of  $F_1$ 23 offspring from conception through weaning and post-weaning exposure were included. Three groups of 24  $F_0$  animals, each consisting of 12 male and 12 female Crl:CD®(Sprague-Dawley) IGS BR rats, were 25 exposed to 300, 1,500, and 6,000 ppm BD via whole-body inhalation exposure 6 h/day for 14 days prior 26 to the breeding period and continuing throughout the gestation and lactation periods. A control group was 27 exposed to clean, filtered air on a comparable regimen. For  $F_0$  dams, the daily inhalation exposures were 28 suspended on GD 21 through lactation day 4, to avoid any confounding effects of exposure on nesting or 29 nursing behavior. Exposures were resumed for these dams on lactation day 5. The  $F_1$  generation pups 30 were potentially exposed to the BD *in utero*, and through nursing during lactation until weaning. 31 Beginning on postnatal day (PND) 21, one male and one female from each litter were exposed for seven 32 consecutive days to the same concentration of the BD concentration as its dam. Beginning on PND 28, 33 one previously unexposed male and one previously unexposed female per litter were exposed for seven 34 consecutive days to the same BD concentration as its dam. 35 36 Under the conditions of the current study, there were no adverse BD-related effects on any parameter 37 measured in either the  $F_0$  or  $F_1$  animals at the exposure level of 300 ppm. Adverse BD-related effects were 38 noted at 1,500 and 6,000 ppm and consisted of persistent reductions in body weight parameters in  $F_0$  and 39 F<sub>1</sub> males and females and transient reductions in food consumption (week 0-1) for F<sub>0</sub> males and females.

40

41 Adverse BD-related effects noted exclusively at 6,000 ppm consisted of clinical observations indicative of

42 chromodacryorrhea, chromorhinorrhea, and salivation in F<sub>0</sub> males and females as well as infrequent

43 occurrences of dried red material in the perioral and perinasal regions of four exposed  $F_1$  pups (three 44 males and one female).

45

46 Based on the results of this study, an exposure level of 300 ppm was considered to be the NOAEL in rats

for  $F_0$  parental systemic toxicity and for systemic toxicity for  $F_1$  animals following post-weaning 6-h daily 47

1 exposures (PND 21-27 or PND 28-34). The NOAEL for effects on gonadal function, mating behavior,

- 2 conception, gestation, parturition, lactation of the  $F_0$  generation, and the development of  $F_1$  offspring from 3 conception through weaning was considered to be 6,000 ppm.
- 4
- 5 The findings of this subchronic reproductive/developmental study showed effects of reduction in body
- 6 weight parameters as the most sensitive endpoint in male and female rats with a NOAEL of 300 ppm.
- 7 Developmental effects were not observed. This study is included in the acute toxicity section because it is
- 8 a well-conducted, high-quality study with a NOAEL of 300 ppm, which is slightly higher than the
- 9 NOAEL of 200 ppm determined in previous rat studies (IISRP 1982; Hackett et al. 1987a).

### 10 3.1.1.2.2.2 Reproductive/Developmental Toxicity in Mice

- 11 Hackett et al. (1987b) exposed pregnant mice for 10 days via inhalation at 0, 40, 200, and 1,000 ppm on
- 12 GD 6-15 for 6 h/day. Maternal toxicity manifested as reduced body weight gain (GD 11-16) and
- extragestational weight gain was observed at 200 and 1,000 ppm. Total body weight at GD 18 was
- decreased at 1,000 ppm. Therefore, the NOAEL for maternal toxicity was 40 ppm. Hackett et al. (1987b)
- 15 reported the most sensitive short-term developmental endpoint was decreased fetal weight in male mice at
- 16 40 ppm. BD caused reduced fetal body weight and increased frequency of skeletal variations at 200 and
- 17 1,000 ppm which are concentrations corresponding to maternal toxicity expressed as reduced body
- 18 weight. Major malformations in the mouse fetus were not detected although the potential for altered
- development was indicated by a dose-related increase in supernumerary ribs and reduced ossifications,
   particularly of the sternebrae.
- 20 21
- Hackett et al. (1987b) reported that statistical differences were observed at the lowest exposure
- concentration of 40 ppm for male fetal body weight. Therefore, a NOAEL was not identified for this
- 24 effect. However, Hackett et al. (1987b) conducted analyses of variance (ANOVA) on the average pup
- 25 weight followed-up by Student's t-tests comparing the average pup weight for different treatment groups.
- 26 Their pairwise comparisons using Student's t-test did not adjust significance levels for the number of
- 27 multiple tests. In addition, their analyses did not adjust for well-known important covariate effects such as
- 28 litter size. Christian (1996) noted that the apparent significant decrease in male fetal weight in the 40 ppm 29 group was the result of the statistical analysis used, which was considered to be inappropriate.
- 29 30
- Data reported by Hackett et al. (1987b) were reanalyzed by Green (2003). The Green (2003) reanalysis
- was based on analysis of covariance (ANCOVA) on the average pup weight adjusted for covariates and
- 32 was based on analysis of covariance (11(COVII) on the average pup weight adjusted for covariates and 33 used the Dunnett-Hsu test to compare the mean weights for each of the exposed groups to the mean
- weight for the control group. Application of the statistical analysis indicates that the 40 ppm exposure
- concentration is a NOAEL in this study. Other previously analyzed endpoints were also analyzed by more
- appropriate methodology (Green 2003). In each instance, the NOAEL was at least as high as previously
- reported. For a few endpoints, a higher NOAEL was found. The overall NOAEL for this study is 40 ppm,
- 38 based on the fetal weights.
- 39
- 40 In order to assess the Green (2003) reanalysis, Sielken et al. (Appendix 1) conducted a review of the
- 41 Hackett et al. (1987b) study and the Green (2003) reanalysis, concentrating on male fetal body weight.
- 42 The Sielken et al. review (Appendix 1) indicates that Green's (2003) conclusions are reasonable and
- 43 based on standard statistical analyses practices that were overlooked by Hackett et al. (1987b). Green
- 44 used the Dunnett-Hsu test to compare the mean weights for each of the exposed groups to the mean
- 45 weight for the control group after both were adjusted for the effects of the covariates. This is the specific

1 situation for which the Dunnett-Hsu test was designed. In addition to reviewing the statistical

2 methodology used in the Hackett et al. (1987b) and Green (2003) studies, Sielken et al. (Appendix 1) re-

3 analyzed the fetal weight data to confirm the numerical results obtained by Green (2003) and performed a

- 4 sensitivity analysis with respect to the effects of covariates, and determined the outcome of the more
- 5 powerful statistical analyses where the individual pup weights were analyzed and the dams were treated
- as random effects. These analyses support the finding that the NOAEL based on fetal weight for this
- 7 study is 40 ppm (Sielken et al. (Appendix 1)).
- 8
- 9 Table 4 is similar to Table 5-6 in USEPA (2002) but only contains parameters that were significantly
- 10 different from controls. There were no statistical differences in number of pregnant dams, litters with live
- 11 fetuses, implantations per dam, resorptions per litter, dead fetuses per litter, fetuses per number of litters
- 12 examined, or sex ratio (% males) between treated mice and control mice (data not shown). The
- 13 highlighted cells in Table 4 have been corrected based on the Hackett et al. (1987b) study reanalyses by
- 14 Green (2003) and Sielken et al. (Appendix 1). The appropriate LOAEL for early resorptions is 1,000 ppm
- 15 (not 200 ppm as reported by Hackett et al. (1987b)) and the LOAEL for decreases in male fetal body
- 16 weight is 200 ppm (not 40 ppm). Decreases in male fetal body weight occur at the same concentrations as
- 17 decreases in maternal weight gain (Table 6).
- 18

19 Table 5 is similar to Table 5-7 in USEPA (2002) but only contains parameters that were significantly

- 20 different from controls. There were no results contrary to those of the Hackett et al. (1987b) after the
- 21 reanalysis by Green (2003). The only fetal effects noted were slight, significant increases in minor
- skeletal abnormalities at 200 and/or 1,000 ppm, indicative of growth retardation. These effects occurred at
- the same concentrations as decreases in maternal weight gain (Table 6).
- 24
- 25
- 26

Parameters	<b>Concentration (ppm)</b>			
	0	40	200	1,000
Early resorptions	$1.00 \pm 0.23$	$0.58 \pm 0.21$	$0.43 \pm 0.13^{\text{ c, g}}$	$0.75 \pm 0.16$
Fetal body weight (g)	$1.34 \pm 0.03$ <sup>b</sup>	$1.28 \pm 0.01$	$1.13 \pm 0.02$ °	$1.04 \pm 0.03^{\circ}$
Females	$1.30 \pm 0.03$ <sup>b</sup>	$1.25 \pm 0.01$	$1.10 \pm 0.02^{\circ}$	$1.06 \pm 0.02^{\mathrm{c,f}}$
Males	$1.38 \pm 0.03^{b}$	$1.31 \pm 0.02^{\text{ c, d}}$	$1.13 \pm 0.02^{\circ}$	$1.06 \pm 0.02^{\circ}$
Placental weight (mg)	$86.8 \pm 2.99^{b}$	85.4 ± 2.29	$78.6 \pm 3.24^{\circ}$	$72.6 \pm 1.88^{\circ}$
Females	$83.1 \pm 3.03^{b}$	$80.9 \pm 2.46$	$74.7 \pm 3.52^{\circ}$	$70.1 \pm 2.33^{\circ}$
Males	$89.3 \pm 3.03^{b, e}$	$89.5 \pm 2.27$	$80.1 \pm 2.35^{\circ}$	$74.5 \pm 1.81^{\circ}$

27 <sup>a</sup> All values mean  $\pm$  standard error from USEPA (2002)

28 <sup>b</sup>  $p \le 0.05$ , significant linear trend

29  $c p \le 0.05$ , pairwise comparison with corresponding control parameter based on Hackett et al. (1987b)

 $^{d}$  p > 0.05 based on Green (2003) and Sielken et al. reanalyses(Appendix 1)

- 31  $e^{8}89.3 \pm 3.05$  (Hackett et al. 1987b)
- 32  $f 1.02 \pm 0.02$  (Hackett et al. 1987b)
- 33  $^{g} p \ge 0.05$  based on Green (2003)
- 34 Source: USEPA (2002)
- 35
- 36

Table 5. Variations in CD-1 Mice	Exposed to B	D by Inhalation		
Parameters		Concentra	tion (ppm)	
	0	40	200	1,000
Variations: Abnormal sternebrae <sup>a, b</sup>	$0.6 \pm 0.9$	$0.4 \pm 0.7$	$0.4 \pm 0.8$	$0.8 \pm 1.3^{\circ}$
Variations: Supernumerary ribs <sup>a, b</sup>	$1.7 \pm 2.3$	$1.6 \pm 2.1$	$6.0 \pm 3.6^{\circ}$	$9.9 \pm 3.0^{\circ}$
Reduced ossification (all sites) <sup>a</sup>	$1.7 \pm 1.7$	$1.2 \pm 1.5$	$2.7 \pm 2.7$	$3.9 \pm 2.6^{\circ}$
Sternebrae	31/13	20/9	57/16 <sup>d</sup>	76/19 <sup>d</sup>

<sup>a</sup> Mean percentage per litter (mean  $\pm$  SD)

<sup>b</sup>  $p \le 0.05$ , significant linear trend, orthogonal contrast test

<sup>c</sup>  $p \le 0.05$ , Tukey's test <sup>d</sup>  $p \le 0.05$ , Fisher exact test (fetal incidence)

Source: USEPA (2002)

1

Table 6. Maternal Toxicity in	Pregnant CD-	1 Mice Exposed	to BD by Inhala	tion <sup>a</sup>
Parameters		Concent	ration (ppm)	
	0	40	200	1,000
Whole-body weight (g)				
Day 0	$28.4\pm0.25$	$28.3\pm0.32$	$28.3\pm0.32$	$28.4\pm0.32$
Day 18	$54.9 \pm 1.21^{\text{b}}$	$55.4 \pm 1.09$	$52.5 \pm 1.01$	$50.8 \pm 0.86$ <sup>c, f</sup>
Body weight gain (g)				
Days 0-6	$2.7\pm0.3$	$3.0 \pm 0.3$	$2.5 \pm 0.2$	$2.3 \pm 0.2$
Days 6-11	$5.5 \pm 0.4$	$5.8 \pm 0.3$	$5.6 \pm 0.3$	$4.8 \pm 0.3$
Days 11-16	$13.3 \pm 0.6^{b}$	$12.7 \pm 0.4$	$11.4 \pm 0.5^{\circ}$	$10.6 \pm 0.4$ <sup>c</sup>
Days 16-18	$5.5 \pm 0.3^{b}$	$5.7 \pm 0.3$	$4.7 \pm 0.4$	$4.8 \pm 0.3$
Gravid uterine weight (g)	$19.3 \pm 1.00^{b}$	$20.3\pm0.80$	$18.0\pm0.87$	$16.8 \pm 0.67$ <sup>c, g</sup>
Extragestational weight (g) <sup>d</sup>	$35.5 \pm 0.48$ <sup>b</sup>	$35.1 \pm 0.44$	$34.5 \pm 0.46$	$34.1 \pm 0.36$ °
Extragestational weight gain (g) <sup>e</sup>	$7.60 \pm 0.48$ <sup>b</sup>	$6.99\pm0.38$	$6.20 \pm 0.38$ <sup>c</sup>	$5.91 \pm 0.28$ <sup>c</sup>

<sup>a</sup> All values mean  $\pm$  standard error from USEPA (2002) 10

<sup>b</sup>  $p \le 0.05$ , significant linear trend 11

 $^{c}$  p  $\leq$  0.05, pairwise comparison with corresponding control parameter  $^{d}$  Body weight on GD 18 minus gravid uterine weight 12

13

<sup>e</sup> Extragestational weight minus body weight on GD 0 14

15  $f 50.8 \pm 0.87$  (Hackett et al. 1987b)

 $^{g}$  16.7  $\pm$  0.67 (Hackett et al. 1987b) 16

Source: USEPA (2002) 17

1

2 Table 6 is similar to Table 5-5 in USEPA (2002), but only lists data on maternal weight loss measures 3 which are the main parameters that were significantly different from controls. There were no results 4 contrary to those of Hackett et al. (1987b) based on the reanalysis of Green (2003). Table 6 indicates that 5 there was a statistical reduction in extragestational weight gain (i.e., maternal weight minus gravid uterine 6 weight) and weight gain (GD 11-16) at 200 ppm. A statistical decrease in gravid uterine weight did not 7 occur at 200 ppm but did occur at 1,000 ppm. These results suggest that BD produces maternal toxicity 8 but little or no intrauterine effects at 200 ppm. For mice and rats, body weight changes and changes in 9 body weight gain in pregnant dams with no change in gravid uterine weight usually indicate maternal 10 toxicity as discussed by Pohl et al. (1998):

11

"Changes in maternal body weight corrected for gravid uterine weight at sacrifice may indicate
 whether the effect is primarily maternal or fetal. For example, there may be a significant
 reduction in weight gain and in gravid uterine weight throughout gestation but no change in
 corrected maternal weight gain, which would generally indicate an intrauterine effect.
 Conversely, a change in corrected weight gain and no change in gravid uterine weight generally
 suggest maternal toxicity and little or no intrauterine effect."

Therefore, if a point of departure (POD) for maternal toxicity is determined using the endpoints of "extragestational weight gain" and "weight gain at GD 11-16," then potential effects on the developing fetus (i.e., reduction in fetal weight, minor skeletal abnormalities) would be prevented (Tables 5 and 6). Reduction in maternal body weight gain was an effect that was consistently observed in studies in rats, although at much higher concentrations (IISRP 1982; Hackett et al. 1987a; and ACC 2003).

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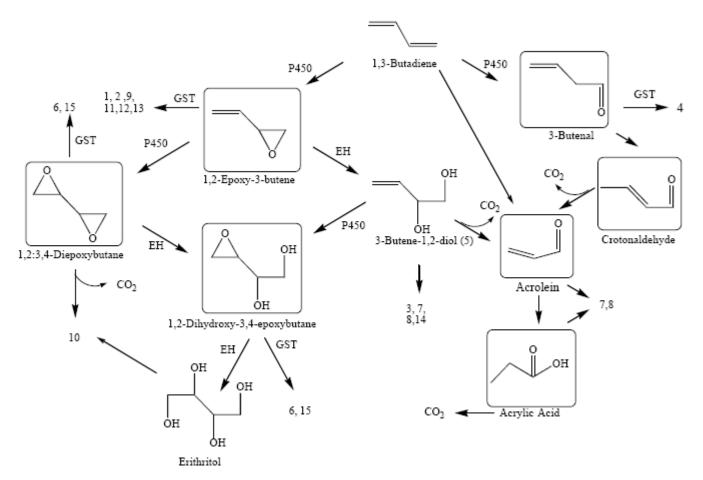
42 43

## 26 **3.1.2 Mode-of-Action (MOA) Analysis**

It is generally agreed that BD produces toxicity when it is metabolized to its reactive metabolites after animals are exposed to BD. However, there is a difference in the metabolism amongst species. The basis of the species differences between rats and mice may be related to the greater production of toxic intermediates and a lower capacity for detoxification of these intermediates, although uptake of BD via inhalation exposure appears to be faster in mice compared to rats and humans (USEPA 2002).

The following chemical terminology, similar to the terminology in USEPA (2002), is used in the DSD.
Figure 2 (below) is Figure 5 from USEPA (2002):

- 1,2-Epoxy-3-butene (EB). EB is also used for epoxybutene, 1,3-butadiene monoepoxide, 1,3-butadiene monoxide, 1,2-epoxybutene-3, vinyl oxirane, and 3,4-epoxy-1-butene;
- 1,2:3,4-Diepoxybutane (DEB). DEB is also used for diepoxybutane, butadiene diepoxide, and butadiene bisoxide;
- 3-Butene-1,2-diol (butene-diol). Butene-diol is also used for 1,2-dihydroxybut-3-ene; and
- 44
   1,2-Dihydroxy-3,4-epoxybutane (EBD). EBD is also used for epoxybutanediol, 3,4-epoxybutanediol, 3,4-epoxybutane-1,2-diol, and 3,4-epoxy-1,2-butanediol.





P450 stands for cytochrome P450, EH stands for epoxide hydrolase, GST stands for glutathione transferase, and GSH stands for glutathione. The reactive metabolites are shown inside boxes. The urinary metabolites are numbered and listed in Table 3-1 of USEPA (2002).

1 The general metabolic scheme of BD, which has been reviewed by Himmelstein et al. (1997), is shown in 2 Figure 2. BD is first metabolized to 1,2-epoxy-3-butene (EB), a process that is primarily associated with CYP 2E1, but can also be accomplished by additional isoforms including CYP 2A6 and 4B1. This 3 4 electrophilic metabolite can be detoxified by conjugation with glutathione and subsequent excretion in the 5 urine as M2. It can also undergo hydrolysis by epoxide hydrolase (EH) to form 3-butene-1,2-diol (butene-6 diol). Butene-diol can also be conjugated with glutathione and subsequently excreted in the urine as M1. 7 It can be further oxidized by cytochrome P450 to the 1,2-dihydroxy-3,4-epoxybutane (EBD). An 8 alternative pathway for the metabolism of EB is oxidation to the 1,2:3,4-diepoxybutane (DEB) which can 9 be further hydrolyzed to EBD or conjugated by glutathione and excreted as M3. This series of 10 epoxidation and detoxication steps generates three electrophilic metabolites: EB, DEB, and EBD. 11 12 Cochrane and Skopek (1994) have shown that DEB is 100 times more mutagenic than EB and 200 times 13 more mutagenic than EBD. Kligerman and Yu (2007) used an in vitro system of lymphocytes treated with 14 EB or DEB and measured sister chromatid exchange and chromosome aberrations. DEB-induced damage 15 for both sister chromatid exchange and chromosome aberrations were persistent in  $G_0$  cells and DEB was much more genotoxic than EB. EB did not induce sister chromatid exchange in lymphocytes unless 16 17 actively cycling cells were treated. The extent to which DEB is produced and reaches target tissues will 18 play a role in the toxicity. The ability of EB to reach actively dividing or repair deficient cells will also 19 somewhat contribute to toxicity (Kligerman and Yu 2007). Mice form more DEB than rats or humans 20 whereas EBD is more readily formed in humans than in rats (Slikker et al. 2004; Swenberg et al. 2007). Human genetic polymorphisms are likely to affect individual susceptibility to BD and its metabolites.

21

22

23 Activation rates in humans exhibit a high degree of variability and appear to span the range of activation

24 rates between mice and rats, so humans may be as sensitive as mice. Several genes appear to be important

25 in the BD metabolic pathway. Inherent susceptibilities have been shown for both EB and DEB (Weincke

26 and Kelsey 1993), which may be due to glutathione S-transferase theta (GSTT1) status. Also, glutathione

27 S-transferase GSTM1 appears to be an important detoxifying factor for EB, so that GSTM1 null

28 individuals would be expected to have greater effects following formation of EB. Unfortunately, no data 29

have been published on the effects of GST polymorphisms of EBD. Genetic polymorphisms have also 30 been identified for EH and CYP 2E1 that would be expected to affect susceptibility to BD and its

31 metabolites. The role of these proteins in the toxicokinetics of numerous chemicals is reasonably well

32 known. Three in vitro studies (Csanády et al. 1992; Seaton et al. 1995; and Duescher and Elfarra 1994)

33 using rodent and human tissue samples have demonstrated that CYP 2E1 plays a role in the oxidation of

34 both BD and EB. Polymorphisms that reduce EH activity may increase susceptibility to BD-induced

35 effects. Likewise, rapid CYP 2E1 metabolizers may potentially be at greater risk.

36

37 The specific mode-of-action (MOA) for the reproductive/developmental effects produced by BD is

38 unknown. Uptake of BD in mice is faster than rats and may account for the increased susceptibility of

39 mice compared to rats. However, the basis of the species differences between rats and mice may be

40 related to the greater production of toxic intermediates and a lower capacity for detoxification of these

41 intermediates (USEPA 2002). Conjugation with GSH is an important detoxification route. Himmelstein et 42 al. (1997) points out that GSH depletion occurs at longer exposure duration or at higher concentrations

43 leading to higher body burdens of EB and DEB (Himmelstein et al. 1997). Based on the above

44 information and consistent with USEPA (2002), the reproductive/developmental effects in mice are

considered to have a threshold (i.e., a nonlinear MOA) and to be concentration and duration dependent. 45

#### 1 **3.1.3 Dose Metric**

2 In the reproductive/developmental studies selected as key studies, data on the exposure concentration of 3 the parent chemical are available. Since the MOA of the toxic response is not fully understood and data

4 on other more appropriate dose metrics are not available (e.g. blood concentration of parent chemical,

5 area under blood concentration curve of parent chemical, or putative metabolite concentrations in blood

- 6 or target tissue), the exposure concentration of the parent chemical was used as the default dose metric.
- 7

## 8 **3.1.4 Points of Departure (PODs) for Key Studies**

9 The LOAEL for maternal toxicity in rats (1,500 ppm) reported from a subchronic study conducted by the

10 American Chemistry Council (ACC 2003) is more than seven times the LOAEL for maternal toxicity

11 observed in mice (200 ppm), so maternal toxicity in rats will not be considered. Decreases in

12 extragestational weight gain and body weight gain (GD 11-16) from the Hackett et al. (1987b) study were

13 modeled because they had the lowest NOAEL and LOAEL (40 and 200 ppm, respectively) (Table 7).

14

Parameters		Concentrati	on (ppm) <sup>a</sup>	
	0	40	200	1,000
No. litters	18	19	21	20
Body weight gain (g)	$13.3 \pm 0.6$ <sup>b</sup>	$12.7 \pm 0.4$	$11.4 \pm 0.5^{\circ}$	$10.6 \pm 0.4$ <sup>c</sup>
Days 11-16	(2.55 SD)	(1.74 SD)	(2.29 SD)	(1.79 SD)
% decrease from controls		4.5%	14.3	20.3%
Extragestational weight gain (g)	$7.60 \pm 0.48$ <sup>b</sup>	$6.99 \pm 0.38$	$6.20 \pm 0.38$ <sup>c</sup>	$5.91 \pm 0.28$ <sup>c</sup>
	(2.04 SD)	(1.66 SD)	(1.74 SD)	(1.25 SD)
% decrease from controls		8.0%	18.4%	22.2%

 $^{a}$  All values mean  $\pm$  standard error from USEPA (2002). Standard deviation (SD) equals the standard error times the square root of n (no. litters)

17 <sup>b</sup>  $p \le 0.05$ , significant linear trend

18  $c^{2} p \leq 0.05$ , pairwise comparison with corresponding control parameter

19

20 Decreases in maternal weight gain on GD 11-16 and decreases in extragestational weight gain were

21 modeled with Benchmark Dose Modeling (BMDS) Software (Version 1.4.1) using both the continuous

22 polynomial model (using three doses and four doses) and the Hill model. Both models provided an

adequate statistical fit to the data (Appendix 2). The polynomial model was fit with unrestricted

24 parameters and is not monotone. Adequacy of fit to the Hill model and the three-dose polynomial model

were determined by visual inspection of the data plot. The three-dose continuous polynomial model and

the Hill model provided a better fit of the data than the four-dose continuous polynomial model based on visual inspection and residual evaluation. Therefore, only the results from these models were considered

visual inspection and residual evaluation. Therefore, only the results from these models were consideredfor selection of the POD. The three- and four-dose linear model (data not shown), and the three- and four-

dose power model (data not shown) provided an adequate statistical fit to both maternal weight gain (GD

30 11-16) and decreases in extragestational weight gain, but did not fit the data as well based on visual

31 inspection and residual evaluation.

32

The 95% lower confidence limit on the concentration corresponding to the benchmark response of a 5%

reduction in weight gain (BMCL<sub>05</sub>) was considered a NOAEL (Table 8). Calculated values at a BMR of 100% and 1 SD are provided in Table 8 as well as modeling results for the polynomial model using four

35 10% and 1 SD are provided in Table 8 as well as modeling results for the polynomial model using four

doses for comparison purposes. Appendix 2 provides modeling output files for modeling results of a 5%
 reduction in extragestational weight gain and maternal weight gain (GD 11-16) for all models in Table 8.

3

4 Reduction in extragestational weight gain was a more sensitive endpoint than reduction in maternal

5 weight gain (GD 11-16) (Table 8). For reduction in extragestational weight gain, the three-dose

6 polynomial  $BMCL_{05}$  was 8.909 ppm and the Hill  $BMCL_{05}$  was 2.811 ppm, which is within a factor of

7 three. The three-dose polynomial model and the Hill model had similar Akaike's information criterion

8 (AIC) values (132 and 165, respectively). According to guidance in USEPA (2000), the average BMCL<sub>05</sub>

9 of the modeling results should be used ((8.909 ppm + 2.811 ppm)/2 = 5.860 ppm). Therefore, 5.860 ppm 10 was chosen as the POD for reduction in extragestational weight gain.

10 11

For reduction in maternal weight gain (GD 11-16), the three-dose polynomial BMCL $_{05}$  was 13.95 ppm

13 and the Hill  $BMCL_{05}$  was 10.31 ppm, which is within a factor of three. The three-dose polynomial model

14 and the Hill model had similar AIC values (155 and 201, respectively). According to guidance in USEPA

15 (2000), the average BMCL<sub>05</sub> of the modeling results should be used ((13.95 ppm + 10.31 ppm)/2 = 12.13

16 ppm). Therefore, the average  $BMCL_{05}$  of 12.13 ppm was chosen as the POD for reduction in maternal 17 weight gain (GD 11-16).

18

## 19 **3.1.5 Dosimetric Adjustments**

20 The USEPA closely examined the physiologically-based toxicokinetic (PBTK) models for BD to

21 determine if additional modeling could reduce uncertainties in the interspecies scaling between mice and

humans for ovarian atrophy and other endpoints (USEPA, 2002, Chapter 9). USEPA stated that despite

advances in the models over the past decade, the current models are inadequate for this purpose. For example, the PBTK models do not yet accurately describe the distribution of the major metabolites in

24 example, the PBTK models do not yet accurately describe the distribution of the major metabolites in 25 various compartments, they do not yet include the reportedly important epoxydiol metabolites, and they

- have not been adequately validated. Recently, Filser et al. (2007) measured and evaluated the BD-
- dependent blood burden of the following metabolites: EB, DEB, EBD and butene-diol (refer to Figure 2)

in rats and mice. Smith et al. (2001) investigated genetic and dietary factors affecting human metabolism

- of BD. Human volunteers were exposed to 2 ppm BD for a 20-min exposure with a 40-min washout
- 30 period. Smith et al. (2001) fitted a three-compartment PBTK model to investigate BD uptake and estimate

31 model parameters. Brochot et al. (2007) conducted a global sensitivity analysis for a proposed PBTK

32 model. However, relevant parameters and a validated PBTK model for extrapolation from animals to

humans is still lacking. Therefore, default duration exposure and dosimetric adjustments from animal-to

- human exposure were used.
- 35

2

Response	Model	Cutoff	BMC (ppm)	BMCL (ppm)
	Polynomial AIC = 199	5% reduction	64.01	41.00
	4 Doses Poor fit	10% reduction	135.8	85.19
		1 SD	230.0	133.5
Reduction in maternal	Polynomial AIC = 155	5% reduction	44.83	13.95
weight gain (GD 11-16)	3 Doses	10% reduction	103.9	29.82
		1 SD		
	Hill AIC = 201	5% reduction	44.49	10.31
		10% reduction	103.8	28.09
		1 SD	245.9	
	Polynomial AIC = 164	5% reduction	50.31	31.23
	doses Poor fit	10% reduction	105.46	64.34
		1 SD	272.5	146.0
Decrease in	Polynomial AIC = 132	5% reduction	23.61	8.909
extragestational weight gain	3 doses	10% reduction	51.93	18.57
0 0		1 SD		
	Hill AIC = 165	5% reduction	23.69	2.811
		10% reduction	53.35	8.174
		1 SD	599.4	

#### 1 3.1.5.1 Default Exposure Duration Adjustments

- 2 The POD of 5.860 ppm based on 6 h/day for 10 days (Hackett et al. 1987b) for reduction in
- 3 extragestational weight gain and 12.13 ppm for reduction in maternal weight gain (GD 11-16) were used.
- 4 The 6-h exposure duration ( $C_1$ ) was adjusted to a POD<sub>ADJ</sub> of 1-h exposure duration ( $C_2$ ) using Haber's
- 5 Rule with n = 3 where both concentration and duration play a role in toxicity: 6
- 7 Reduction in extragestational weight gain 8 POD<sub>ADI</sub> =  $C_2$  =  $[(C_1)^3 x (T_2)^3 x (T_2)^3$
- 9 10

 $POD_{ADJ} = C_2 = [(C_1)^3 x (T_1 / T_2)]^{1/3}$  $= [(5.860 \text{ ppm})^3 x (6 \text{ h/1 h})]^{1/3}$ = 10.65 ppm

1112 Reduction in maternal weight gain (GD 11-16)

13  $POD_{ADJ} = C_2 = [(C_1)^3 x (T_1 / T_2)]^{1/3}$ 14  $= [(12.13 \text{ ppm})^3 x (6 \text{ h/1 h})]^{1/3}$ 15 = 22.04 ppm

#### 16 3.1.5.2 Default Dosimetry Adjustments from Animal-to-Human Exposure

17 BD is only slightly soluble in water and is moderately soluble in blood (USEPA 2002). It is readily 18 absorbed from the air into the blood through the lungs. The health effects it produces at lower 19 concentrations are mainly remote effects, so dosimetric adjustments were performed as a Category 3 gas 20 which is consistent with USEPA (2002) and based on guidance in USEPA (1994). For Category 3 gases, 21 the default dosimetric adjustment from animal-to-human exposure is conducted using the following 22 equation: 23  $POD_{HEC} = POD_{ADJ} \times \left[ (H_{b/g})_A / (H_{b/g})_H \right]$ 24 25 26 where: 27 28 ratio of the blood:gas partition coefficient H<sub>b/g</sub> 29 А = animal 30 Η = human 31 32 For BD, the blood:gas partition coefficients for mice range from 1.2 to 3.0 with a mean of 1.67 (Appendix 33 3 of USEPA 2005a) and for humans  $1.22 \pm 0.30$  (mean  $\pm$  SD) (Brochot et al. 2007). When  $(H_{b/g})_A$  /  $(H_{b/g})_{H}$ , > 1, a default value of 1 is used for  $(H_{b/g})_{A} / (H_{b/g})_{H}$ , the regional gas dose ratio (RGDR) (USEPA) 34 35 1994). 36 37 Reduction in extragestational weight gain 38  $POD_{HEC} = POD_{ADJ} \times RGDR = 10.65 \text{ ppm x } 1 = 10.65 \text{ ppm}$ 39 40 Reduction in maternal weight gain (GD 11-16) 41  $POD_{HEC} = POD_{ADJ} \times RGDR = 22.04 \text{ ppm x } 1 = 22.04 \text{ ppm}$ 

## **3.1.6 Adjustments of the POD<sub>HEC</sub> and Critical Effect**

#### 2 3.1.6.1 Uncertainty Factors (UFs)

The MOA by which BD produces maternal toxicity is not understood (Section 3.1.2), so the default for noncarcinogenic effects is to determine a POD and apply UFs to derive a ReV (i.e., assume a nonlinear MOA). The following UFs were applied to the POD<sub>HEC</sub> of 10.65 ppm and 22.04 ppm: 10 for intraspecies variability (UF<sub>H</sub>), 3 for extrapolation from animals to humans (UF<sub>A</sub>), 1 for extrapolation from a LOAELto-NOAEL (UF<sub>L</sub>), and 1 for database uncertainly (UF<sub>D</sub>), a total UF = 30:

9 Reduction in extragestational weight gain

10 acute ReV = POD<sub>HEC</sub> / (UF<sub>H</sub> x UF<sub>A</sub> x UF<sub>L</sub> x UF<sub>D</sub>) = 10.65 ppm / (10 x 3 x 1 x 1) = 0.3550 ppm

1112 Reduction in maternal weight gain (GD 11-16)

13 acute ReV = POD<sub>HEC</sub> / (UF<sub>H</sub> x UF<sub>A</sub> x UF<sub>L</sub> x UF<sub>D</sub>) = 22.04 ppm / (10 x 3 x 1 x 1) = 0.7347 ppm

14

8

15 A full  $UF_H$  of 10 was used to account for intraspecies variability because there is experimental evidence

16 that indicates that BD-sensitive human subpopulations may exist due to metabolic genetic polymorphisms

17 (USEPA 2002). A  $UF_A$  of 3 was used for extrapolation from animals to humans because default

18 dosimetric adjustments from animal-to-human exposure were conducted which accounts for toxicokinetic

19 differences but not toxicodynamic differences. This approach is conservative, since existing studies

20 indicate that mice are relatively sensitive laboratory animals in regards to the reproductive effects of BD

21 (e.g., relatively high respiratory rates, greater production of toxic intermediates, and a lower capacity for 22 detoxification of these intermediates (USEPA 2002)). A UF<sub>L</sub> of 1 was used because BMC modeling was

performed to determine a POD based on the BMCL<sub>05</sub> and the BMR was set at a response level considered

to be a NOAEL (i.e., 5% decrease in weight gain). A database  $UF_D$  of 1 was used because the overall

acute toxicological database for BD meets the minimum database requirements used to derive an acute

26 ReV (TCEQ 2006) (i.e., acute inhalation studies in humans, two inhalation bioassays in different species

27 investigating a wide range of endpoints and two prenatal developmental toxicity studies in different

28 species (USEPA 2002; AEGL 2005)). Both the quality of the studies and the confidence in the acute

29 database is high.

## 30 3.1.6.2 Critical Effect

The critical effect is a 5% reduction in extragestational weight gain for pregnant CD-1 mice exposed to

BD (Hackett et al. 1987b) because it has the lowest  $POD_{HEC}$  and the lowest acute ReV of 0.3550 ppm. In

33 addition, reduction in body weight was the health effect observed at the lowest BD concentrations in

34 studies conducted in both mice and rats.

## 35 **3.1.7 Health-Based Acute ReV and** <sup>acute</sup>ESL

36 The acute ReV value in Section 3.1.6.2 for extragestational weight gain was rounded to the least number

of significant figures for a measured value at the end of all calculations. Rounding to two significant

figures, the 1-h acute ReV is 360 ppb (800  $\mu$ g/m<sup>3</sup>). The rounded acute ReV was then used to calculate the

 $^{acute}ESL$ . At the target hazard quotient of 0.3, the  $^{acute}ESL$  is 110 ppb (240  $\mu$ g/m<sup>3</sup>) (Table 9). This acute

40 ReV and <sup>acute</sup>ESL are conservative since pregnant mice exposed to BD develop maternal toxicity much

41 easier than similarly exposed rats do, available scientific information suggests mice are more sensitive

42 than humans, and reproductive/developmental effects have never been observed in humans.

1 2

Table 9. Derivation of the Acute ReV and <sup>acute</sup>	ESL
Study	Hackett et al. 1987b
Study population	CD-1 mice (18-21 pregnant mice per dose group)
Study quality	high
Exposure Methods	10-day exposures via inhalation at 0, 40, 200, and 1,000 ppm on gestation days (GD) 6-15 for 6 h/day
Critical Effects	Reduction in extragestational weight gain; maternal toxicity
POD	5.86 ppm (average BMCL <sub>05</sub> )
Exposure Duration	6 h
Extrapolation to 1 h	Haber's Law with $n = 3$
$POD_{ADJ}(1 h)$	10.65 ppm
POD <sub>HEC</sub>	10.65 ppm (gas with systemic effects, based on default RGDR = $1.0$ )
Total uncertainty factors (UFs)	30
Interspecies UF	3
Intraspecies UF	10
LOAEL UF	1
Incomplete Database UF	1
Database Quality	high
acute ReV [1 hr] (HQ = 1)	800 µg/m <sup>3</sup> (360 ppb)
$^{acute}ESL [1 h] (HQ = 0.3)$	240 µg/m <sup>3</sup> (110 ppb)

3

### 4 3.2. Welfare-Based Acute ESLs

### 5 3.2.1 Odor Perception

ACGIH (2001) reports BD has a mildly aromatic odor with recognition occurring at 1 to 1.6 ppm. Ruth
(1986) states the 50% odor detection threshold is 352 μg/m<sup>3</sup> (160 ppb) and the 100% recognition
threshold is 2,860 μg/m<sup>3</sup> (1,300 ppb). The 50% odor detection threshold for BD determined by the
triangular odor bag method was 230 ppb (Nagata 2003). Both Ruth (1986) and Nagata (2003) are listed as
sources of information for odor thresholds in Appendix B of the ESL Guidelines (TCEQ 2006). However,
only the Nagata (2003) study meets the criteria for acceptable odor threshold measurement techniques
developed by the American Industrial Hygiene Association (TCEO 2006). Therefore, the <sup>acute</sup>ESL<sub>odor</sub> is

developed by the American Industrial Hygiene Association (TCEQ 2006). Therefore, the  $^{acute}ESL_{odor}$  is 230 ppb (510 µg/m<sup>3</sup>). Since odor is a concentration-dependent effect, the same 1-h  $^{acute}ESL_{odor}$  is assigned

14 to all averaging times.

### 15 **3.2.2 Vegetation Effects**

16 BD concentrations that produce vegetative effects, such as abscission and inhibition of growth, are orders

17 of magnitude higher than concentrations of ethylene, propylene, and acetylene that produce similar effects

1 (USDHEW 1970). Since concentrations producing vegetative effects (approximately > 10,000 ppm) are 2 significantly above other health- and odor-based concentrations, an <sup>acute</sup>ESL<sub>veg</sub> was not developed for BD.

## 3 3.3. Short-Term ESL and Values for Air Monitoring Evaluation

4 The acute evaluation resulted in the derivation of the following values:

- Acute ReV =  $800 \ \mu g/m^3$  (360 ppb)
- $^{acute}ESL = 240 \ \mu g/m^3 \ (110 \ ppb)$
- $^{\text{acute}}\text{ESL}_{\text{odor}} = 510 \ \mu\text{g/m}^3 \ (230 \text{ ppb})$

8 9 The short-term ESL for air permit evaluations is the health-based <sup>acute</sup>ESL of 240  $\mu$ g/m<sup>3</sup> (110 ppb) as it is 10 lower than the <sup>acute</sup>ESL<sub>odor</sub> (Table 1). For the evaluation of ambient air monitoring data, the <sup>acute</sup>ESL<sub>odor</sub> of 11 510  $\mu$ g/m<sup>3</sup> (230 ppb) is lower than the acute ReV of 800  $\mu$ g/m<sup>3</sup> (360 ppb) (Table 1), although both values 12 will be used for the combustion of air data (Table 1)

12 will be used for the evaluation of air data (Table 1).

# *3.4 Derivation of Acute ReV versus USEPA's Acute Reference Concentration* (*RfC*)

USEPA (2002) derived an acute RfC of 3.2  $\mu$ g/m<sup>3</sup> (7 ppb) representative for a 24-h exposure duration

16 based on developmental toxicity (decreased male fetal body weight at 40 ppm was inappropriately

assumed to be the LOAEL). A value of 2.9 ppm for a 24-h POD<sub>HEC</sub> is reported in Table 10-25 of USEPA.

18 USEPA applied UFs of 3 for interspecies variability, 10 for intraspecies variability, 4 for effect level

19 extrapolation factor (to decrease risk to below the benchmark response level; analogous conceptually to

20 the LOAEL-to-NOAEL UF) and 3 for incomplete database because a neurodevelopmental toxicity study

has not been completed (total UF = 400) (Table 10).

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23 The acute ReV for a 1-h exposure duration is based on maternal toxicity (5% reduction in extragestational

24 weight gain). The adjusted 1-hr POD<sub>HEC</sub> is 10.65 ppm. A UF of 3 was applied for interspecies

extrapolation and 10 for intraspecies variability (total UF = 30). The Toxicology Section (TS) did not

apply an effect level extrapolation factor analogous to a LOAEL-to-NOAEL UF since numerous

27 investigators have demonstrated or recommended that the BMCL<sub>05</sub> is analogous to the NOAEL and 28 should be treated as such (Derros et al. 1995; Fauda et al. 1996; Filingeon et al. 2002). An assist database

should be treated as such (Barnes et al. 1995; Fowles et al. 1999; Filipsson et al. 2003). An acute database
 UF was not applied because the acute database for BD is adequate (i.e., meets the minimum database used

29 UF was not applied because the acute database for BD is adequate (i.e., meets the minimum database used 30 to derive an acute Rev (TCEQ (2006)). Table 10 compares the derivation of the 1-h acute ReV and

 $a^{\text{cute}}$ ESL to USEPA's 24-h acute RfC (USEPA 2002).

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Table 10. Acute	Table 10. Acute ReV Compared to USEPA's Acute RfC										
POD <sub>HEC</sub>	Inter- species	Intra- species	Effect Level Extrapolation Factor	Incomplete Database	Total UF	Acute Reference Value					
TCEQ 10.65 ppm [1 h] <sup>1</sup> Maternal toxicity (5% reduction in extragestational weight gain)	3	10			30	acute ReV [1 h] 360 ppb <sup>acute</sup> ESL [1 h] 110 ppb					
USEPA 2.9 ppm [24 h] <sup>2</sup> Fetal toxicity (5% reduction in fetal body weight)	3	10	4	3	400	acute RfC [24 h] 7 ppb					

1

<sup>1</sup> The unadjusted 6-h BMCL<sub>05</sub> for a 5% reduction in maternal extragestational weight gain was 5.860 ppm

 $2^{2}$  The unadjusted 6-h BMCL<sub>05</sub> for a 5% reduction in decreased fetal body weight was 11.6 ppm

## 3 Chapter 4 Chronic Evaluation

## 4 4.1 Noncarcinogenic Potential

## 5 4.1.1 Physical/Chemical Properties and Key Studies

- 6 Refer to Section 3.1.1.1 for a discussion of physical/chemical properties.
- 7

8 This section is based on USEPA (2002). Chapter 5 of USEPA (2002) discusses the chronic

9 reproductive/developmental effects of BD. Animal data indicate that BD is a potential reproductive

10 hazard because reproductive effects are observed at the lowest concentrations tested in animals. Chapter 6

11 of USEPA (2002) discusses other subchronic and chronic health effects observed in animals exposed to

12 BD. Few adverse noncarcinogencic effects have been observed other than reproductive and

13 developmental effects, except for hematological effects in mice exposed to higher concentrations and

14 increases in organ weights in rats (USEPA 2002, Chapter 6). A review of the scientific literature since

15 2002 did not reveal any other chronic inhalation studies that could be used instead of the 2-year chronic

16 bioassays conducted by the National Toxicology Program (NTP 1993) which are summarized in the

17 following sections but discussed in detail in USEPA (2002).

### 18 4.1.1.1 Human Studies

- 19 Albertini et al. (2007) conducted a molecular epidemiological study of BD-exposed Czech workers to
- 20 compare female to male responses. The focus of the study was to collect data on urine concentrations of
- 21 BD metabolites and blood concentrations of BD-metabolite hemoglobin adducts. However, questionnaire
- 22 responses for female-specific adverse health questions in control and exposed females were obtained.
- 23 There were 26 female control workers and 23 female BD-exposed workers. The years spent in the

- 1 company were  $17.6 \pm 9.3$  years for control and  $19.4 \pm 9.9$  years for exposed females (mean  $\pm$  S.D.).
- 2 Multiple external exposure measurements were obtained (10 full 8-h shift measures by personal
- 3 monitoring per worker) over a 4-month period before biological samples were collected. Mean 8-h time-
- 4 weighted average (TWA) exposure levels were  $0.008 \text{ mg/m}^3$  (0.0035 ppm) for controls and 0.397 mg/m<sup>3</sup>
- 5 (0.180 ppm) for exposed but with individual single 8-h TWA values up to  $9.793 \text{ mg/m}^3$  (4.45 ppm) in the
- 6 exposed group. Analysis of questionnaire responses for female-specific adverse health questions showed
- 7 no significant differences between controls and exposed for miscarriages, still births, ectopic pregnancies, 8 malar programming law hitth weight (< 2.500 g) babies, or pro-term hitths, head on information collected
- 8 molar pregnancies, low birth weight (<2,500 g) babies, or pre-term births, based on information collected 9 on all pregnancies.
- 10

### 11 4.1.1.2 Animal Studies

The most sensitive reproductive effects observed in 2-year chronic exposure studies were ovarian atrophy in female mice and testicular atrophy in male mice (NTP 1993). Testicular atrophy was primarily a high-

- exposure effect so this section focuses on ovarian atrophy. In this bioassay, groups of 70 female B6C3F1
- mice were exposed by inhalation 6 h/day, 5 days/week to 0, 6.25, 20, 62.5, or 200 ppm BD for up to 103
- 16 weeks and groups of 90 female mice were exposed to 625 ppm. An interim evaluation of ovarian atrophy
- 17 was conducted at 9 months on ten mice per group and also at 15 months. Significant concentration-related
- decreases in survival were seen in female mice exposed to concentrations  $\geq 20$  ppm, primarily due to the
- 19 development of malignant neoplasms. Statistically significant increases in the incidence of ovarian
- 20 atrophy were observed in all exposure groups following lifetime exposures. The LOAEL for ovarian
- atrophy was observed at the lowest exposure level (6.25 ppm, 6 h/day, 5 days/week, for 2 years). Uterine
- atrophy was also observed in the highest exposure groups; however, this is likely to be a secondary effect
- 23 of ovarian atrophy. Similarly exposed rats did not develop adverse reproductive effects, thus providing
- 24 further evidence that rats are less sensitive to the effects of BD than mice.

## 25 4.1.2 MOA Analysis

- 26 Refer to Section 3.1.2 for a discussion of BD metabolism. There is strong evidence that ovarian atrophy is
- 27 mediated by the diepoxide metabolite, DEB, the most reactive of BD metabolites (Doerr et al. 1995,
- 28 1996; USEPA 2002). There are marked species differences in effects seen between rats which do not
- 29 exhibit BD-induced ovarian atrophy, and mice which do exhibit BD-induced ovarian atrophy Doerr et al.
- 30 (1995, 1996) evaluated the ovarian effects of the metabolites of BD in mice and rats and also examined 4-
- 31 vinylcyclohexene, a structurally similar compound. Doerr et al. (1995, 1996) showed that the diepoxide of 32 BD or 4-vinylcyclohexene is required for ovarian toxicity to occur in the rat. EB was ovotoxic to mice but
- BD or 4-vinylcyclohexene is required for ovarian toxicity to occur in the rat. EB was ovotoxic to mice but not rats. Thus, the resistance of the rat to ovarian toxicity of BD is likely due to the decreased ability of
- the rat to produce DEB. Filser et al. (2007) was unable to detect DEB in venous blood of male Sprague-
- 250 J and 250 J was unable to detect DED in vehiclds blood of male Sprague-35 Dawley rats (detection limit 0.01  $\mu$ mol/l) when they were exposed to 1,200 ppm for 6-8 hr, whereas DEB
- 36 was detected in B6C3F1 mice at 3.2 µmol/l at 1,280 ppm BD. Humans are similar to rats in that they do
- 37 not readily produce the diepoxide metabolite.
- 38
- 39 Swenberg et al. (2007) compared results in Czech Republic occupationally-exposed workers to results in
- 40 mice and rats for a N,N-(2,3-dihydroxy-1,4-butadiyl) valine (pry-Val) hemoglobin adduct specific for
- 41 DEB at similar BD concentrations (Table 11). The pry-Val adduct was not detected in human females or
- 42 males, while female mice were 78 times more likely than human females to produce DEB as evaluated
- 43 with pry-Val adducts (Table 11). Pry-Val adducts for human females were based on the limit of
- 44 quantitation (LOQ) because pry-Val adducts were not detected (Swenberg et al. 2007). At the 2007

1 Society of Toxicology meeting, Georgieva et al. (2007) presented results using a more sensitive analytical

2 method to measure pry-Val adducts. Pry-Val adducts were detected at low concentrations in Czech

3 Republic workers, although there was not a clear dose-response relationship between pry-Val adducts and

4 BD concentrations which may indicate pry-Val adducts are formed from other unknown sources.

5 6

# Table 11. DEB-Specific pyr-Val Hb Adduct in Mouse, Rat, and Human from Swenberg et al. (2007)

	1	DD	1	DD	M 0.10 C	M 0.27 C		
Concentration	1 ppm BD		1 ppm BD		Mean 0.18 ppm for	Mean 0.37 ppm for		
	6 h/day	4 weeks	6 h/day 4 weeks		4 months	4 months		
	(4.0 ppm-weeks)		(4.0 ppm-weeks)		(4.0 ppm-wee		(3.1 ppm-weeks)	(6.3 ppm-weeks)
Species	Female mice	Male	Female	Male	Female	Male		
		mice	rat	rat	human	human		
Pyr-VAL	$23.5 \pm 3.1$	$30.8\pm4.6$	$0.7 \pm$	$0.9 \pm$	< 0.3 (LOQ)	< 0.3 (LOQ)		
Hb adducts			0.1	0.03				
	female mice	male mice have						
(pmol/g in	have 78 times	103 times more						
50 mg globin)	more pyr-Val	pyr-Val adducts						
	adducts than	than male						
	female humans	humans						

7

## 8 4.1.3 Dose Metric

9 For ovarian atrophy, data on the exposure concentration of the parent chemical are available whereas data

10 on more appropriate dose metrics, such as the monoepoxide or diepoxide metabolites in blood or target

11 tissue, are not available. As discussed previously in Section 3.1.5, a validated PBTK model for

12 extrapolation from animals to humans is still lacking. Therefore, the exposure concentration of the parent

13 chemical was used as the default dose metric.

## 14 **4.1.4 PODs for Key Studies and Critical Effect**

15 Using benchmark concentration dose modeling and a Weibull time-to-response model, USEPA (2002)

16 calculated a BMC<sub>10</sub> of 1.05 ppm and BMCL<sub>10</sub> of 0.88 ppm based on the 1993 NTP 2-year inhalation

17 bioassay, including interim sacrifice data. In calculating the  $BMC_{10}$  and  $BMCL_{10}$ , lesion severity was not

taken into account, and the 625 ppm group was excluded because of high early mortality. In addition,

19 ovarian atrophy was modeled to reflect extra risks only until age 50, because BD-induced ovarian atrophy

20 is believed to result from follicular failure, and after menopause, follicles would no longer be available.

- 22 The PODs for all prenatal deaths (dominant lethal effect) (BMCL $_{05}$  = 10 ppm) and for testicular atrophy
- 23 (BMCL<sub>10</sub> = 16 ppm) were also determined by USEPA (2002) and were significantly higher than the
- 24 BMCL<sub>10</sub> of 0.88 ppm. Therefore, ovarian atrophy was selected as the critical effect (USEPA 2002).
- 25
- 26 Sielken et al. (Appendix 3) repeated the BMC modeling performed by USEPA using the same procedures
- described above and calculated the  $BMC_{05}$  and  $BMCL_{05}$  as well as the  $BMC_{10}$  and  $BMCL_{10}$  (Appendix 3).
- 28 The BMCL<sub>05</sub> has generally been considered a NOAEL (Barnes et al. 1995; Fowles et al. 1999; Filipsson
- et al. 2003) whereas the BMCL<sub>10</sub> may be analogous to a NOAEL or LOAEL. The BMC<sub>10</sub> and BMCL<sub>10</sub>

1 calculated by Sielken et al. (Appendix 3) were 1.15 ppm and 0.881 ppm, respectively, which agreed with 2 the  $BMC_{10}$  of 1.05 ppm and  $BMCL_{10}$  of 0.88 ppm calculated by USEPA (2002).

3

4 USEPA (2002) analyzed ovarian atrophy data excluding the highest dose group and also including all the 5 data. Traditionally, EPA drops the highest dose group when the model does not fit the data well due to 6 some biological phenomenon or when quantal data are fit with a quantal model and there is high mortality 7 in the highest dose group. The ovarian atrophy data, however, were modeled with a time-to-response 8 model (i.e., a model that accounts for the time of death) as opposed to a quantal model which do not 9 account for time of death. Furthermore, the model fit to the data that excluded the highest dose group was 10 not better than the model fit to the data that included the highest dose group as shown by Sielken et al. (Appendix 3). However, USEPA (2002) excluded the highest dose group because of early mortality. The 11 12  $BMC_{05}$  and  $BMCL_{05}$  were 0.560 ppm and 0.429 ppm, respectively, excluding the highest dose and 0.607 13 ppm and 0.462 ppm, respectively, including the highest dose. Since a time-to-response model was used, 14 the TS used the BMCL<sub>05</sub> modeling result of 0.462 ppm that uses all the data as the POD. 15 16 Because the Weibull time-to-response model in these analyses is linear in dose, the BMC<sub>05</sub> and BMCL<sub>05</sub> 17 values are approximately half the corresponding BMC<sub>10</sub> and BMCL<sub>10</sub> values. The values of BMC<sub>05</sub> and 18  $BMCL_{05}$  can be used if the dose-response relationship below the lowest experimental dose is believed to 19 be the linear Weibull time-to-response model fit to the data. The assumption of linearity below the lowest 20 experimental dose is usually conservative and, therefore, health protective. However, the motivation

behind the benchmark dose methodology is to identify the POD ( $BMC_{05}$  and  $BMCL_{05}$ ) to be within the

range of the experimental data (the range of the non-zero doses in the experimental data) and to be a dose

whose risk can be reasonably reliably estimated without undue sensitivity to the dose-response model selected or the model estimation. Here, the  $BMC_{05}$  and  $BMCL_{05}$  are below the range of the experimental

24 selected of the model estimation. Here, the BMCL<sub>05</sub> and BMCL<sub>05</sub> are below the range of the experimental 25 data and, hence, introduce an additional element of uncertainty into the POD. However, the BMCL<sub>05</sub> for

ovarian atrophy was used as the POD because the TS preferentially uses a benchmark response level of

27 5% for more severe effects such as ovarian atrophy, and the BMCL<sub>05</sub> is considered to be a NOAEL

- 28 (TCEQ 2006).
- 29

## 30 4.1.5 Dosimetric Adjustments

Based on the summary of information in Section 3.1.5 and the detailed discussion in USEPA (2002,

Chapter 9), default duration exposure and dosimetric adjustments from animal-to-human exposure wereused.

## 34 4.1.5.1 Default Exposure Duration Adjustments

The BMCL<sub>05</sub> = 0.462 ppm for ovarian atrophy (Appendix 3) represents exposure concentrations that were already adjusted from discontinuous to continuous exposures.

## 37 4.1.5.2 Default Dosimetry Adjustments from Animal-to-Human Exposure

BD is only slightly soluble in water and is moderately soluble in blood (USEPA 2002). It is readily

- absorbed from the air into the blood through the lungs. The health effects it produces at lower
- 40 concentrations are mainly remote effects, so dosimetric adjustments were performed as a Category 3 gas
- 41 which is consistent with USEPA (2002) and based on guidance in USEPA (1994). For Category 3 gases,

the default dosimetric adjustment from animal-to-human exposure is conducted using the following
 formula:

3				
4		$POD_{H}$	$_{\rm EC} = P$	$OD_{ADJ} \ge [(H_{b/g})_A / (H_{b/g})_H]$
5				
6		where	:	
7				
8		$H_{b/g}$	=	ratio of the blood:gas partition coefficient
9		Α	=	animal
10		Н	=	human
11				
10	T.	DD 1 11		

For BD, the blood:gas partition coefficients for mice range from 1.2 to 3.0 with a mean of 1.67 (Appendix
3 of USEPA 2005a) and for humans 1.22 ± 0.30 (mean ± SD) (Brochot et al. 2007). When (H<sub>b/g</sub>)<sub>A</sub> /
(H<sub>b/g</sub>)<sub>H</sub> > 1, a default value of 1 is used for (H<sub>b/g</sub>)<sub>A</sub> / (H<sub>b/g</sub>)<sub>H</sub>, the regional gas dose ratio (RGDR) (USEPA
1994).
POD<sub>HEC</sub> = POD<sub>ADJ</sub> x RGDR = 0.462 ppm x 1 = 0.462 ppm

## 17 4.1.6 Adjustments of the POD<sub>HEC</sub>

The MOA by which BD produces ovarian atrophy is metabolism of the parent compound to DEB (Section 4.1.2), which is considered a threshold, nonlinear MOA. Therefore, a POD was determined and UFs applied to derive a ReV. The following UFs were applied to the POD<sub>HEC</sub> of 0.462 ppm: 10 for intraspecies variability (UF<sub>H</sub>), 1 for extrapolation from animals to humans (UF<sub>A</sub>), 1 for extrapolation from a LOAEL-to-NOAEL (UF<sub>L</sub>), 1 for extrapolation from a subchronic to chronic study (UF<sub>Sub</sub>), and 3 for database uncertainly (UF<sub>D</sub>), a total UF = 30:

25	Chronic ReV	=	$POD_{HEC} / (UF_H \times UF_A \times UF_L \times UF_{Sub} \times UF_D)$
26		=	0.462 ppm / (10 x 1 x 1x 1 x 3)
27		=	0.0154 ppm
28			

A full  $UF_H$  of 10 was used to account for intraspecies variability because there is experimental evidence to indicate that BD-sensitive human subpopulations may exist due to metabolic genetic polymorphisms (USEPA 2002).

32

33 The UF<sub>A</sub> is composed of a toxicokinetic and toxicodynamic component. A toxicokinetic UF<sub>A</sub> of 1 was 34 used for extrapolation from animal to human because default dosimetric adjustments from animal-tohuman exposure were conducted (Section 4.1.5.2). A toxicodynamic UF<sub>A</sub> of 1 was used because humans 35 36 produce much lower levels of DEB than mice as demonstrated by experimental data on DEB-specific pyr-37 Val adducts (Section 4.1.2). DEB is the BD metabolite responsible for ovarian atrophy (Section 4.1.2; 38 USEPA 2002). Although these experimental data are not sufficient to develop a chemical-specific 39 adjustment factor (CSAF) for BD, it would support an interspecies UF substantially less than 1. At the 40 present time, procedures for developing a CSAF based on Hb adducts or for decreasing the UF to a value 41 less than one based on other considerations are not available. 42 43 A UF<sub>L</sub> of 1 was used because BMC modeling was performed to determine a POD based on the BMCL<sub>05</sub>

- 44 (TCEQ 2006). A UF<sub>sub</sub> of 1 was used because the study was a chronic study. The toxicological database
- 45 for BD is extensive. However, a UF<sub>D</sub> of 3 was applied because of the absence of multigenerational

1 reproductive studies (USEPA 2002). Both the quality of the studies and the confidence in the chronic

2 database is high.

3

#### 4.1.7 Health-Based Chronic ReV and <sup>chronic</sup>ESL<sub>nonlinear(nc)</sub> 4

5 The chronic ReV value based on ovarian atrophy was rounded to the least number of significant figures

6 for a measured value at the end of all calculations. Rounding to two significant figures, the chronic ReV is

15 ppb (33  $\mu$ g/m<sup>3</sup>). The rounded chronic ReV was then used to calculate the <sup>chronic</sup>ESL <sub>nonlinear(nc)</sub>. At the target hazard quotient of 0.3, the <sup>chronic</sup>ESL <sub>nonlinear(nc)</sub> is 4.5 ppb (10  $\mu$ g/m<sup>3</sup>) (Table 12).

7 8 9

Table 12. Derivation of the Chronic l	Table 12. Derivation of the Chronic ReV and <sup>chronic</sup> ESL <sub>nonlinear(nc)</sub>					
Study	2-year bioassays (NTP 1993)					
Study Population	70 female B6C3F1 mice; 90 female mice					
Study Quality	high					
Exposure Method	103 week exposures via inhalation at 0, 6.25, 20, 62.5, or 200 ppm; 90 female mice exposed to 625 ppm					
Critical Effects	ovarian atrophy in female mice					
POD (original animal study)	Not available. BMD modeling was conducted on data already adjusted from discontinuous to continuous exposure					
Exposure Duration	6 hs/day, 5 days/week, for 2 years					
Extrapolation to continuous exposure $(POD_{ADJ})$	0.462 ppm (BMCL <sub>05</sub> )					
POD <sub>HEC</sub>	0.462 ppm (gas with systemic effects, based on default RGDR = 1.0)					
Total UFs	30					
Interspecies UF	1					
Intraspecies UF	10					
LOAEL UF	1					
Subchronic to chronic UF	1					
Incomplete Database UF	3					
Database Quality	high					
Chronic ReV (HQ = 1)	33 μg/m <sup>3</sup> (15 ppb)					
$^{chronic}ESL_{nonlinear(nc)} (HQ = 0.3)$	10 μg/m <sup>3</sup> (4.5 ppb)					

## 1 4.1.8 Derivation of Chronic ReV versus USEPA's Chronic RfC

2 Table 13 provides a comparison of the derivation of the chronic ReV of 33  $\mu$ g/m<sup>3</sup> (15 ppb) versus the

3 chronic RfC of 2  $\mu$ g/m<sup>3</sup> (0.9 ppb) (USEPA 2002). USEPA (2002) applied a toxicodynamic UF<sub>A</sub> of 3 but

4 the TS applied a toxicodynamic  $UF_A$  of 1 for reasons explained in Sections 4.1.2 and 4.1.6. Based on

5 experimental data and the MOA of BD, a UF<sub>A</sub> of 1 is conservative and a UF<sub>A</sub> < 1 may be justified.

6 USEPA applied an effect level extrapolation factor to the BMCL<sub>10</sub> because USEPA considered it a

- significant response level. In contrast, the TS used a BMCL<sub>05</sub>, which is considered to be a NOAEL, so a  $UF_{L}$  was not applied.
- 9

Table 13. Compa	Fable 13. Comparison of Chronic ReV and Chronic RfC									
Chronic Toxicity Value	POD <sub>HEC</sub>	UF <sub>H</sub>	UFA	$UF_L$ or Effect Level Extrapolation Factor	UF <sub>Sub</sub>	UF <sub>D</sub>	Total UFs	Chronic Toxicity Value		
ReV based on ovarian atrophy	462 ppb BMCL <sub>05</sub> including highest dose	10	1	1 UF <sub>L</sub>	1	3	30	15 ppb		
RfC based on ovarian atrophy	880 ppb BMCL <sub>10</sub> excluding highest dose	10	3	10 Effect Level extrapolation Factor	1	3	1,000	0.88 ppb		

## 10 4.2 Carcinogenic Potential

## 11 4.2.1 Carcinogenic Weight-of-Evidence and MOA

12 USEPA has classified BD as known to be carcinogenic to humans by inhalation (DHHS 2000; USEPA

13 2002) based on the following findings:

- Increased lymphohematopoietic cancers in workers occupationally exposed via inhalation to BD based on epidemiologic studies (leukemias in polymer workers and non-Hodgkin's lymphoma in monomer workers);
- BD causes a variety of tumors in mice and rats by inhalation in various studies;
- Demonstration that BD is metabolized into genotoxic metabolites by experimental animals and humans.

20 Table 14 provides information on the carcinogenic weight-of-evidence provided by other organizations.

21 Although the MOA by which BD produces tumors is unknown, scientific evidence suggests that

22 carcinogenic effects are mediated by genotoxic metabolites of BD (i.e., EB, DEB, and EBD, Section 3.1.2

and Figure 2). A detailed review of the weight-of-evidence, carcinogenic hazard assessment, and MOA

analysis for lifetime exposure potential is included in USEPA (2002). Preston (2007) recently reviewed

- 1 the evidence that BD works through a mutagenic MOA and concluded: "For butadiene, the MoA is DNA-
- 2 reactivity and subsequent mutagenicity and so following the EPA's cancer guidelines, a linear
- extrapolation is used from the POD, unless additional data support a non-linear extrapolation." Therefore, 3
- an inhalation unit risk factor (URF) and <sup>chronic</sup>ESL<sub>linear(c)</sub> (i.e., air concentration at 1 in 100,000 excess 4
- 5 cancer risk) was developed for BD.
- 6

Table 14. Carcinogenic W	Table 14. Carcinogenic Weight-of-Evidence						
International Agency for Research on Cancer 2007	Group 1, Carcinogenic to humans						
National Institute for Occupational Safety and Health 1997	Potential occupational carcinogen						
Occupational Safety and Health Administration 1996	"Potential occupational carcinogen" There is strong evidence that workplace exposure to BD poses an increased risk of death from cancers of the lymphohematopoietic system.						
ACGIH 2001	A2, Suspected Human Carcinogen						
USEPA 2002; DHHS 2000	Carcinogenic to humans by inhalation						

7

#### 4.2.2 Epidemiological Studies and Exposure Estimates 8

9 Chapter 7 of USEPA (2002) discusses the epidemiologic studies of carcinogenicity for BD and Chapter

10 10 discusses the dose-response assessment of the preferred occupational epidemiological study conducted

by researchers at the University of Alabama at Birmingham (UAB) (Delzell et.al. 1995; 1996). Numerous 11

12 epidemiology studies were reviewed, but USEPA (2002) concluded the UAB exposure estimates

provided the best published set of data to evaluate human cancer risk from BD exposure. USEPA 13

14 published an inhalation URF of 3.0 x  $10^{-5}/(\mu g/m^3)$  or 0.08/ppm based on leukemia mortality data from the UAB occupational epidemiological study (Delzell et.al. 1995; 1996).

- 15
- 16

17 Delzell et al. (1995, 1996) investigated a cohort of synthetic rubber production workers exposed to BD in 18 a retrospective cohort mortality study. The investigators developed a job exposure matrix (JEM) for BD,

19 styrene, and benzene based on industrial hygiene data, which contained estimates of the average daily

20 exposure (in ppm based on the 8-h TWA) and the number of annual peaks (defined as > 100 ppm for BD

21 and >50 ppm for styrene) for each area and job code for each study year. The investigators were then able

22 to estimate cumulative exposures (ppm-years) and number of peak exposures (peak years) for each

- 23 individual worker by linking the JEM with the study subjects' work histories.
- 24

25 Recently, the UAB epidemiology study of leukemia was updated (Sathiakumar et al. 2005; Graff et al.

26 2005; HEI 2006; Cheng et al. 2007) as well as the BD occupational exposure estimates for the

27 occupational cohort (Macaluso et al. 2004). In addition, Sathiakumar et al. (2007) assessed the validity of

28 the BD exposure estimates. These new, updated studies were used by the TS to update the USEPA (2002)

29 assessment. A review of the scientific literature indicated there were no other epidemiology studies (e.g.,

30 Alder et al. 2006) that would be appropriate to evaluate human cancer risk from BD exposure.

1

2 Subjects included in the updated study were 16,579 men classified as having worked (for at least one year

3 before 1 January 1992) at any of six synthetic rubber plants located in Texas (two plants), Louisiana (two

4 plants), Kentucky (one plant) and Canada (one plant). Of the 16,579 subjects in the updated study, 488

5 subjects were excluded because they dropped out of follow-up at ages younger than the youngest

6 leukemia decedent (age 33 years) (Cheng et al. 2007). Thus, results of leukemia analyses were based on

16,091 subjects and 485,732 person-years of observation. The updated study provided seven more years
 of follow-up (through 1998), a larger number of decedents, and a total of 81 deaths with leukemia as the

9 primary or contributing cause. The association of BD exposure to lymphoid and myeloid neoplasms was

investigated. BD-exposure estimates were also updated, and quantitative estimates of each subject's

exposure to butadiene, styrene and dimethyldithiocarbamate (DMDTC) were calculated. DMDTC is an

12 immune system depressant (Irons and Pyatt 1998; Irons et al. 2001).

### 13 **4.2.3 Dose-Response Assessment**

#### 14 4.2.3.1 Beta coefficient (β) and Standard Error Based on Observed Data

15 Cheng et al. (2007) investigated the dose-response relationship between BD and leukemia rate ratios

16 using an exponential Cox regression analysis. Cumulative exposure to BD in ppm-years, cumulative

17 number of exposures to >100 ppm BD (referred to as peaks), and average intensity (ppm) of exposure to

BD were considered as BD exposure dose metrics. The term "peak" is used by the UAB group to refer to

19 the cumulative number of exposures to >100 ppm BD. These exposures were frequently of short duration

20 (several seconds to several minutes). However, the term "peak" or "peak exposures" is misleading and

21 will not be used in this assessment. Instead, the more descriptive term "number of high-intensity tasks"

22 (i.e., number of HITs) was used.

23

24 Whereas Cheng et al. (2007) used the exponential Cox regression analysis, Sielken et al. (2007) used a 25 linear Poisson regression analysis to investigate the relationship between BD and leukemia rate ratios. Cheng et al. (2007) and Sielken et al. (2007) calculated betas ( $\beta$ ) (maximum likelihood estimates (MLEs)) 26 27 and standard errors (SE) from the updated UAB epidemiological study and updated exposure estimates 28 (Table 15). The TS used these values to calculate 95% upper confidence limit (UCL) values, URFs and 29 corresponding air concentrations at 1 in 100,000 excess cancer risk (Table 16). The data needed to 30 conduct a detailed MOA analysis were not available, so the use of a biologically-based model was not possible. Rather, standard models such as the linear Poisson regression, a conservative linear default 31 32 model, and exponential Cox regression hazard proportional models were selected. The dose metric was 33 cumulative BD parts per million years (ppm-years), a dose metric commonly used for epidemiological 34 studies. Three other BD exposure indices were evaluated by Cheng et al. (2007): (1) continuous, time-

dependent BD exposure indices ppm-years); (2) the total number of exposures to BD concentrations >100

ppm (number of HITs) and (3) average intensity of BD. All three BD exposure indices were positively

37 associated with leukemia.

1

Table 15. Values of Maximum Likelihood Estimate (MLE) of Beta (β), Standard Error (SE), and 95% Upper Confidence Limit (UCL) on β <sup>a</sup>								
Covariates	Model	Source	β (MLE)	Standard Error (SE)	β (95% UCL) <sup>b</sup>			
Age	Cox regression ppm-years continuous <sup>c</sup>	Cheng et al. (2007)	2.9E-04	1.0E-04	4.545E-04			
0	Cox regression ppm-years mean-scored deciles <sup>d</sup>	Cheng et al. (2007)	7.5E-04	2.2E-04	1.112E-03			
	Poisson regression ppm-years mean-scored deciles <sup>d</sup>	Sielken et al. (2007)	1.68E-03	8.21E-04	3.031E-03			
Age & Number of HITs > 100	Cox regression ppm-years continuous <sup>c</sup> , # of HITs continuous <sup>e</sup>	Cheng et al. (2007)	2.5E-04	1.2E-04 <sup>g</sup>	4.474E-04			
ppm	Cox regression ppm-years continuous <sup>c</sup> , # of HITs categorical <sup>f</sup>	Sielken et al. (Appendix 4)	2.0E-04	1.3E-04	4.138E-04			
	Cox regression ppm-years mean-scored deciles <sup>d</sup> , # of HITs categorical <sup>f</sup>	Sielken et al. (Appendix 4)	2.8E-04	2.4E-04	6.748E-04			
	Poisson regression ppm-years mean-scored deciles <sup>d</sup> , # of HITs categorical <sup>f</sup>	Sielken et al. (2007)	1.89E-04	3.6E-04	7.812E-04			
Age &	Cox regression ppm-years continuous <sup>c</sup>	Cheng et al. (2007)	3.0E-04	1.4E-04	5.303E-04			
Other Covariates <sup>h</sup>	Cox regression ppm-years mean-scored deciles <sup>d</sup>	Cheng et al. (2007)	5.8E-04	2.7E-04	1.024E-03			

<sup>a</sup> units are in ppm-years and based on occupational exposure concentrations

<sup>b</sup>  $\beta$  (95% UCL) =  $\beta$ (MLE) + (1.645 x SE)

<sup>°</sup> ppm-years is included as a continuous variable (untransformed) in a parametric model of the effect of ppm-years

<sup>d</sup> ppm-years is included as a categorical variable (based on mean-scored deciles, untransformed) in a parametric model of the effect of ppm-years

<sup>e</sup> number of HITs > 100 ppm is included as a continuous variable (untransformed) in a parametric model of the effect of the number of HITS > 100 ppm

8

9 <sup>f</sup> number of HITs > 100 ppm is included as a categorical variable (based on quintiles) in a nonparametric model of

10 the effect of the number of HITs > 100 ppm

11 <sup>g</sup> back calculated from the corresponding p-value in Cheng et al. (2007)

12 <sup>h</sup> other covariates are year of birth, race, DMDTC, years since hire and plant

13

14

1 Table 15 shows results from the BD dose-response relationship conducted by Cheng et al. (2007) using 2 exponential Cox regression procedures adjusted either for age as a covariate, age + number of HITs > 100ppm, or adjusted for other covariates (age, year of birth, race, plant, years since hire and DMDTC). Cox 3 4 regression procedures permit estimation of the dose-response relationship throughout the exposure range 5 and also potentially provide optimal control of confounding by age (in these Cox regressions age is the 6 index variable and implicitly a covariate). Cheng et al. (2007) results support the presence of a 7 relationship between high cumulative exposure and leukemia and high intensity of exposure and 8 leukemia. Dose-response modeling was conducted with continuous, untransformed data and mean-scored 9 deciles of BD exposure. The impact of exposure estimate errors/misclassification may be somewhat 10 alleviated by the use of mean-scored deciles of BD exposure (Cheng et al. 2007). Sielken et al. (Appendix 11 4) provided  $\beta$  (MLE) and SE values not included in the Cheng et al. (2007) analyses: (1) Cox regression, 12 mean-scored deciles, when evaluating age + number of HITs as covariates and (2) Cox regression, ppm-13 years continuous, number of HITs categorical, when evaluating age + number of HITs as covariates. 14 15 The UAB group recommended the estimate of the dose-response relationship that is based on the continuous, untransformed form of BD ppm-years, age as the covariate, and the full range of exposure 16 17 data (2.9E-04 ( $\beta$ ), 1.0E-04 (S.E.)). However, due to the high potential for distortion of the dose-response 18 relationship as a result of exposure misclassification, Cheng et al. (2007) also recommended that an 19 uncertainty analyses be incorporated into any risk assessment that uses these data. 20 21 Beta estimates were also calculated by Cheng et al. (2007) for unlagged and lagged BD exposure but these  $\beta$  estimates were not used by the TS because lagging BD exposure had little impact on the dose-22 23 response relationship between leukemia and BD ppm-years. The association of BD exposure with 24 leukemia, lymphoid neoplasms and myeloid neoplasms was examined. Lymphoid neoplasms were

associated with ppm-years and myeloid neoplasms were associated with number of HITs in models that
controlled only for age but not after adjusting for multiple covariates (age, year of birth, race, plant, years
since hire and DMDTC). These potency estimates were not used by the TS because evidence of an
association between BD and all lymphoid neoplasm or all myeloid neoplasms was not persuasive (Cheng
et al. 2007).

29 30

31 Sielken et al. (2007) used a linear Poisson regression model to examine the dose-response relationships

32 adjusted either for age as a categorical covariate and age + number of HITs as covariates (Table 15).

33 Sielken et al. (2007) found that if the exposure dosimetric is cumulative ppm-years, the performance of

34 the predictor for leukemia rate ratio is statistically significantly improved if the categorical covariates age

+ number of HITs are included in the Poisson regression model. If covariates other than age + number of

36 HITs are included, the model fit using cumulative ppm-years was not significantly improved except for

37 styrene. However, if styrene was included as a covariate, the slope was negative, so styrene was not

included as a covariate. Although Sielken et al. (2007) performed this statistical analysis for covariates

using the Poisson regression model, his findings are generally applicable for the Cox regression model.

40

#### 41 4.2.3.2 Dosimetric Adjustments

42 Occupational concentrations were converted into environmental concentrations for the general population

- 43 using the following equation:
- 44

1 2	Concentration <sub>HEC</sub> = Concentration <sub>OC</sub> x ( $VE_{ho}/VE_{h}$ ) x (days per year <sub>oc</sub> /days per year <sub>res</sub> )
3	where: $VE_{ho}$ = occupational ventilation rate for an 8-h day (10 m <sup>3</sup> /day)
4	$VE_h$ = non-occupational ventilation rate for a 24-h day (20 m <sup>3</sup> /day)
5	days per year <sub>oc</sub> = occupational exposure frequency (240 days)
6	days per year <sub>res</sub> = residential exposure frequency (365 days)
7	
8	RG-442 ESL Guidelines (TCEQ 2006) recommends using the ratio "5 days per week/7 days per week" to
9	adjust occupational exposure concentrations to concentrations relevant to the general population, but it is
10	standard practice in USEPA epidemiological cancer risk assessments to use the ratio "240 days/365
11	days," which is slightly more conservative. The value of 240 days per year = $52$ weeks x 5 days per week
12	= 260 days per year minus approximately 10 holidays (Christmas, New Year's, Independence Day, etc.)
13	and minus approximately 2 weeks vacation or sick days.
14	1 7 2 2 Entranglation to Lower Engaging
14	4.2.3.3 Extrapolation to Lower Exposures
15	4.2.3.3.1 URFs and Air Concentrations at 1 in 100,000 Excess Cancer Risk
16	Table 16 shows estimates of air concentrations at 1 in 100,000 excess cancer risk (10 <sup>-5</sup> -risk air
17	concentrations) based on $\beta$ s (column three) and 95% UCLs (column five) using the exponential Cox
18	Regression and linear Poisson regression models. Air concentrations were solved iteratively with life-
19	table analyses using the BEIR IV approach (NRC 1988). Air concentrations based on extra risk were
20	calculated as opposed to added risk. The following mortality and survival rates were used to calculate air
21	concentrations based on a lifetime exposure of 70 years, the default used by TCEQ for exposure analysis:
22	
23	• US mortality rates for 2000-2003 for all leukemia (Surveillance, Epidemiology, and End Results
24	database (SEER 2006)) (Appendix 5)
25	• US survival rates for 2000 (Arias 2002) (Appendix 5).
26	
27	Columns four and six provide URFs calculated using the linear extrapolation default approach (USEPA
28	2005a; TCEQ 2006). Risk estimates are obtained by first calculating a $POD_{HEC}$ at the low end of the range
29	of observations using appropriate models, and then extrapolating to zero by means of a straight line
30	(linear extrapolation default). The air concentration at 0.1% excess risk level (i.e., 1 in 1,000 excess
31 32	cancer risk) is chosen for determining the POD because it is within the observable response range of leukemia deaths. The URFs in units of $ppm^{-1}$ at the POD <sub>HEC</sub> (when the POD <sub>HEC</sub> was set to either the
32	effective concentration (EC) or the 95% UCL lowest effective concentration (LEC)) was calculated as
33 34	follows:
35	10110WS.
36	$URF = 0.001/EC_{001}$
37	$URF = 0.001/LEC_{001}$
38	
39	Columns four and six also provide 10 <sup>-5</sup> -risk air concentrations based on the corresponding URFs. Air
40	concentrations calculated using the corresponding URFs are more conservative than air concentrations
41	calculated based on the Cox regression model, because this model is an exponential model. As a health-
40	$10^{-5}$ right on a construction of the time of the

protective policy decision, 10<sup>-5</sup>-risk air concentrations calculated with a URF based on the default linear approach was adopted and all subsequent discussions will refer to the URF (MLE) or URF (95%UCL) and their corresponding 10<sup>-5</sup>-risk air concentration values. 42

- 43
- 44

Covariates	URFs and Air Concer Model	β (MLE)	EC <sub>001</sub>	β (95% UCL)	LEC <sub>001</sub>
Covariates	type of data	р (MILE)	$\frac{\text{LC}_{001}}{\text{URF (MLE)}^{a}}$	p (9376 UCL)	URF (95% UCL)
		Air Concentration 1 in 100,000 excess cancer risk using model	Air Concentration 1 in 100,000 excess cancer risk using URF	Air Concentration 1 in 100,000 excess cancer risk using model	Air Concentration 1 in 100,000 excess cancer risk using URF
Age	Cox regression <sup>h</sup>	80.42 ppb	1.490E-04/ppm	51.31 ppb	2.334E-04/ppm
	ppm-years continuous <sup>c</sup> Cox regression <sup>h</sup> ppm-years mean-scored deciles <sup>d</sup>	31.09 ppb	67.14 ppb 3.852E-04/ppm 25.96 ppb	20.97 ppb	42.84 ppb 5.712E-04/ppm 17.51 ppb
	Poisson regression <sup>i</sup> ppm-years mean-scored deciles <sup>d</sup>	13.91 ppb	7.184E-04/ppm 13.92 ppb	7.713 ppb	1.296E-03/ppm 7.715 ppb
Age & Number of HITs > 100	Cox regression <sup>h</sup> ppm-years continuous <sup>c</sup> , # of HITs continuous <sup>e</sup>	93.28 ppb	1.284E-04/ppm 77.88 ppb	52.13 ppb	2.298E-04/ppm 43.52 ppb
ррт	Cox regression <sup>j</sup> ppm-years continuous <sup>c</sup> , # of HITs categorical <sup>f</sup>	116.6 ppb	1.027E-04/ppm 97.35 ppb	56.36 ppb	2.125E-04/ppm 47.05 ppb
	Cox regression <sup>j</sup> ppm-years mean-scored deciles <sup>d</sup> , # of HITs categorical <sup>f</sup>	83.29 ppb	1.438E-04/ppm 69.53 ppb	34.56 ppb	3.466E-04/ppm 28.85 ppb
	Poisson regression <sup>i</sup> ppm-years mean-scored deciles <sup>d</sup> , # of HITs categorical <sup>f</sup>	123.7 ppb	8.083E-05/ppm 123.7 ppb	29.92 ppb	3.341E-04/ppm 29.93 ppb
Age & Other	Cox regression <sup>h</sup> ppm-years continuous <sup>c</sup>	77.74 ppb	1.541E-04/ppm 64.90 ppb	43.98 ppb	2.724E-04/ppm 36.71 ppb
Covariates g	Cox regression <sup>h</sup> ppm-years mean-scored deciles <sup>d</sup>	40.21 ppb	2.979E-04/ppm 33.57 ppb	22.78 ppb	5.260E-04/ppm 19.01 ppb

<sup>a</sup> URF = 0.001/EC<sub>001</sub> <sup>b</sup> URF = 0.001/LEC<sub>001</sub> <sup>c</sup> ppm-years is included as a continuous variable (untransformed) in a parametric model of the effect of ppm-years <sup>d</sup> ppm-years is included as a categorical variable (based on mean-scored deciles, untransformed) in a parametric model of the

effect of ppm-years <sup>e</sup> number of HITs > 100 ppm is included as a continuous variable (untransformed) in a parametric model of the effect of the

number of HITs > 100 ppm is included as a continuous variable (unralistorined) in a parametric model of the effect of the number of HITs > 100 ppm is included as a categorical variable (based on quintiles) in a nonparametric model of the effect of the number of HITs > 100 ppm

<sup>g</sup> Other covariates are year of birth, race, DMDTC, years since hire and plant

<sup>h</sup> Cheng et al. (2007)

<sup>i</sup> Sielken et al. (2007) <sup>j</sup> Sielken et al. (Appendix 4)

#### 1 **4.2.3.3.2** Age as a Covariate

2 Models that only include age as a non-exposure covariate have the advantage of model parsimony (i.e., 3 the model includes as few variables as necessary to explain the relationship when there is not sufficient

4 biological knowledge to justify the inclusion or exclusion of a variable). When age is included as a

5 covariate (Table 17), the  $10^{-5}$ -risk air concentrations using the Poisson regression model were the most

6 conservative: 13.92 ppb (MLE) and 7.715 ppb (95% UCL). However, as stated previously, Cox

7 regression models provide optimal control of confounding by age, so estimates from the Cox regression

8 model are preferred over estimates using the Poisson regression model. Using the Cox regression model,

9 the  $10^{-5}$ -risk air concentrations for mean-scored deciles (25.96 ppb MLE and 17.51 ppb 95% UCL) were 10 more conservative than continuous, untransformed data (67.14 ppb MLE and 42.84 ppb 95% UCL). The

more conservative than continuous, untransformed data (67.14 ppb MLE and 42.84 ppb 95% UCL). The  $10^{-5}$ -risk air concentration estimates based on mean-scored deciles are preferred because the impact of

- exposure estimate errors/misclassification may be somewhat alleviated by the use of categorical deciles of
- 13 BD exposure (Cheng et al. 2007).

14

Table 17. Age as a Covariate		
	EC <sub>001</sub>	LEC <sub>001</sub>
Model	URF (MLE) <sup>a</sup>	URF (95% UCL) <sup>b</sup>
	Air Concentration	Air Concentration
type of data	1 in 100,000 excess	1 in 100,000 excess cancer
	cancer risk using URF	risk using URF
Cox regression	1.490E-04/ppm	2.334E-04/ppm
Cheng et al. (2007)	67.14 ppb	42.84 ppb
ppm-years continuous <sup>c</sup>		
Cox regression	3.852E-04/ppm	5.712E-04/ppm
Cheng et al. (2007)		
ppm-years mean-scored deciles <sup>d</sup>	25.96 ppb	17.51 ppb
Poisson regression	7.184E-04/ppm	1.296E-03/ppm
Sielken et al. (2007)		
ppm-years mean-scored deciles <sup>d</sup>	13.92 ppb	7.715 ppb

15 <sup>a</sup> URF =  $0.001/EC_{001}$ 

16 <sup>b</sup> URF =  $0.001/LEC_{001}$ 

<sup>c</sup> ppm-years is included as a continuous variable (untransformed) in a parametric model of the effect of
 ppm-years

<sup>19</sup><sup>d</sup> ppm-years is included as a categorical variable (based on mean-scored deciles, untransformed) in a

20 parametric model of the effect of ppm-years

21

#### 22 **4.2.3.3.3 Age + Number of HITs as Covariates**

23 The USEPA Science Advisory Board (USEPA 1998) recommended that consideration of peak exposures

to BD (i.e., number of BD HITs > 100 ppm) be evaluated during its review of the draft health risk

assessment of BD (USEPA 1998b). Sielken et al. (2007) demonstrated that when the categorical

covariates of age + number of HITs are included in the Poisson regression model, the model's ability to

27 predict the leukemia rate ratio was statistically improved. The cumulative number of HITs may better

28 explain the increased leukemia mortality observed in the BD worker cohort.

1

2 If age + number of HITs are included as covariates, the  $10^{-5}$ -risk air concentrations using URFs (MLE)

- range from 69.53 to 123.7 ppb (Table 18), less than a factor of two. The most conservative  $10^{-5}$ -risk air
- 4 concentration that is obtained using the URF (MLE) is calculated with the Cox regression model, mean-
- 5 scored deciles. The  $10^{-5}$ -risk air concentrations using URFs (95% UCL) range from 28.85 to 47.05 ppb,
- 6 less than a factor of two. The most conservative  $10^{-5}$ -risk air concentration of 28.85 ppb is obtained using 7 the URF (95% UCL) calculated from the Cox regression model, mean-scored deciles, although the  $10^{-5}$ -
- the URF (95% UCL) calculated from the Cox regression model, mean-scored deciles, although the  $10^{-5}$ risk air concentration of 29.93 ppb from the Poisson regression, mean-scored deciles, is essentially
- identical. The use of mean-scored deciles for both the Cox regression and the Poisson regression tend to
- minimize the effects of exposure misclassification and address concerns about exposure estimates and are
- preferred. For predicting risk relevant to the general population, the estimates that account for age +
- number of BD HITs > 100 ppm as covariates are the most relevant because it is highly unlikely that the

general population will ever be exposed to peak exposures of BD HITs > 100 ppm.

13 14

> Table 18. Age + Number of BD HITs > 100 ppm as Covariates
>  Model EC<sub>001</sub> LEC<sub>001</sub> URF (MLE)<sup>a</sup> URF (95% UCL) b type of data **Air Concentration Air Concentration** 1 in 100,000 excess cancer 1 in 100.000 excess cancer risk risk using URF using URF 1.284E-04/ppm 2.298E-04/ppm Cox regression Cheng et al. (2007) ppm-years continuous <sup>c</sup> 77.88 ppb 43.52 ppb # of HITs continuous <sup>e</sup> Cox regression 1.027E-04/ppm 2.125E-04/ppm Sielken et al. (Appendix 4) ppm-years continuous <sup>c</sup> 97.35 ppb 47.05 ppb # of HITs categorical <sup>f</sup> Cox regression 1.438E-04/ppm 3.466E-04/ppm Sielken et al. (Appendix 4) ppm-years mean-scored deciles <sup>d</sup> 69.53 ppb 28.85 ppb # of HITs categorical <sup>f</sup> Poisson regression 8.083E-05/ppm 3.341E-04/ppm Sielken et al. (2007) ppm-years mean-scored deciles <sup>d</sup> 123.7 ppb 29.93 ppb # of HITs categorical<sup>f</sup>

15 <sup>a</sup> URF =  $0.001/EC_{001}$ 

 $^{10}$   $^{10}$  URF = 0.001/LEC<sub>001</sub>

- <sup>c</sup> ppm-years is included as a continuous variable (untransformed) in a parametric model of the effect of
   ppm-years
- <sup>19</sup> <sup>d</sup> ppm-years is included as a categorical variable (based on mean-scored deciles, untransformed) in a
- 20 parametric model of the effect of ppm-years
- 21 <sup>e</sup> number of HITs > 100 ppm is included as a continuous variable (untransformed) in a parametric model
- 22 of the effect of the number of HITs > 100 ppm
- <sup>1</sup> number of HITs > 100 ppm is included as a categorical variable (based on quintiles) in a nonparametric
- 24 model of the effect of the number of HITs > 100 ppm

#### 1 4.2.3.3.4 Other Covariates

2 Cheng et al. (2007) fit models that adjusted for age, year of birth, race, DMDTC, years since hire and

- 3 plant. Except for the exposure covariate DMDTC, an immune system depressant (Irons and Pyatt 1998;
- 4 Irons et al. 2001), these covariates are the ones typically evaluated in epidemiology dose-response
- 5 models. Sielken et al. (2007) included statistically-based covariates and determined that if covariates
- 6 other than age + number of HITs were included, the model fit using cumulative ppm-years was not
- significantly improved except for the covariate, styrene. Therefore, URF (MLE) and URF (95% UCL)
   from models that adjusted for age, year of birth, race, DMDTC, years since hire and plant were not
- roll models that adjusted for age, year of birth, race, DMDTC, years since fine and plant were not
   considered as potency factors by the TS, although these values are provided in Tables 15 and 16 for
- 10 comparison purposes.

### 11 4.2.4 Potency Estimate Selected to Represent Excess Leukemia Mortality Risk

The cancer potency estimates and  $10^{-5}$ -risk air concentrations using URFs (MLE) in Table 16 range from 12 7.184E-04/ppm (13.92 ppb) to 8.083E-05/ppm (123.7 ppb). The cancer potency estimates and 10<sup>-5</sup>-risk air 13 14 concentrations using URFs (95% UCL) in Table 16 range from 1.296E-03/ppm (7.715 ppb) to 2.125E-04/ppm (47.05 ppb). Of the various estimates presented in Table 16, the potency estimate of 3.466E-15 04/ppm (10<sup>-5</sup>-risk air concentrations of 28.85 ppb) from the Cox regression model, mean-scored deciles, 16 17 age + number of BD HITs as covariates, based on the URF (95% UCL) and a linear default approach is 18 selected to represent the excess leukemia mortality risk from the occupational data. However, refer to 19 Sections 4.2.4.1 and 4.2.4.2 for additional adjustments to the URF (95% UCL) and 10<sup>-5</sup>-risk air 20 concentrations. 21

A linear default was used to extrapolate to lower concentrations and the URF (95% UCL) was preferred
 to account for the following uncertainties:

- Uncertainty of calculating potency estimates for BD from mortality data, not incidence data (i.e., to protect against developing leukemia) (Section 4.2.5.1)
- Uncertainty of calculating potency estimates for the general population when the SBR cohort was comprised primarily of males (Section 4.2.5.2)

30 The URFs (MLE) based on mean-scored deciles were preferred because the impact of exposure estimate

- 31 errors/misclassification may be somewhat alleviated by the use of categorical deciles of BD exposure
- 32 (Section 4.2.5.3). The Cox regression model was preferred because it potentially provides optimal control
- of confounding by age. While the cancer potency estimate from the Poisson regression (mean-scored
- deciles, age + number of BD HITs) of 3.341E-04/ppm (10<sup>-5</sup>-risk air concentrations of 29.93 ppb) is
- 35 similar to the values from the Cox regression, it is slightly less conservative.
- 36

25

26

27

28

29

- 37 The models that use age + number of BD HITs > 100 ppm as covariates are preferred because once age is
- in the model, inclusion of number of BD HITs results in a significant improvement in the fit (likelihood).
- In addition, the general population is not expected to be exposed to BD concentrations greater than 100
- 40 ppm, so adjusting for BD HITs > 100 ppm as a covariate produces cancer potency estimates more
- 41 relevant to BD exposures experienced by the general population. Slikker et al. (2004) provides a
- 42 discussion of the role of dose-dependent transitions in mechanisms of toxicity for BD as well as several 42 at the schemicale. For every table to be a several time scheme a from the hydroletic
- 43 other chemicals. Exposure to BD at high concentrations may result in a change from the hydrolytic
- 44 pathways that are normally used by humans to form EBD to the formation of the more toxic metabolite, 45 DEB (i.e. metabolic any may be acturated). In addition, protective any may and other protective
- 45 DEB (i.e., metabolic enzymes may be saturated). In addition, protective enzymes and other protective

1 cellular constituents may be depleted which could result in mechanisms of toxic tissue injury that are not

2 relevant at exposures significantly less than 100 ppm. In a study conducted by Albertini et al. (2001), a

3 clear NOAEL for biomarkers of effect (hypoxanthine-guanine phosphoribosyltransferase (HPRT)

4 mutations and chromosome aberrations) at mean BD exposure concentrations of 0.800 ppm was reported

5 in a study of workers in the Czech Republic (see Section 4.5 for additional information).

### 6 4.2.4.1 Evaluating Susceptibility from Early-Life Exposures

USEPA (2005b) provides default, age-dependent adjustment factors (ADAFs) to account for potential
increased susceptibility in children due to early-life exposure when a chemical has been identified as
acting through a mutagenic MOA for carcinogenesis and the cancer assessment did not include exposures
at an early age (generally before age 16). This is the case for the epidemiological leukemia data utilized in
this evaluation. BD is currently identified by USEPA as having a mutagenic MOA. USEPA (2005b)
states:

- "The following adjustments represent a practical approach that reflects the results of the preceding analysis, which concluded that cancer risks generally are higher from early-life exposure than from similar exposure durations later in life:
  - For exposures before 2 years of age (i.e., spanning a 2-year time interval from the first day of birth up until a child's second birthday), a 10-fold adjustment.
  - For exposures between 2 and <16 years of age (i.e., spanning a 14-year time interval from a child's second birthday up until their sixteenth birthday), a 3-fold adjustment
- 2324 For exposures after the
  - For exposures after turning 16 years of age, no adjustment."

The ADAF is an adjustment to the slope factor (as opposed to an adjustment to the dose metric). The ADAF is to be applied on an age-specific basis. That is, the ADAFs are applied to each relevant year in a life and the risks for all years summed to get the lifetime risk, as opposed to calculating a lifetime excess risk without ADAFs and then multiplying this calculated value by a constant ADAF.

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31 When the dose metric is cumulative exposure and when using a life-table analysis BEIR IV approach

- 32 (NRC 1988), an implementation consistent with USEPA guidelines is to calculate the excess risk in each
- 33 year using the age-specific dose (cumulative dose) for that year and multiply the slope by the age-specific

ADAF for that year (age). This is consistent with USEPA's guidelines from the point of view of both

excess risk being calculated using age-specific exposures and ADAFs being age-specific modifiers of the

slope (potency). That is, the excess risk in year "i" is calculated with the  $\beta$  or 95% UCL multiplied by ADAF(i). Refer to Appendix 6 *Calculating Excess Risk with Age-Dependent Adjustment Factors using a* 

ADAF(i). Refer to Appendix 6 Calculating Excess Risk with Age-Dependent Adjustment Factors using a
 Life-Table Analysis.

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40 The TS calculated potency factors both with and without ADAFs. When the ADAFs are not applied, the

- 41 selected potency estimate is 3.466E-04/ppm. When the ADAFs are incorporated into the life-table
- 42 analyses using the BEIR IV approach (NRC 1988), the selected potency estimate is 3.505E-04/ppm.
- 43

#### 1 4.2.4.2 Relevance of Estimated Risks to the Texas General Population

2 There is uncertainty about whether potency estimates are representative of the mortality risks that might 3 be associated with environmental BD exposures in Texas because potency estimates were developed 4 based on the leukemia mortality experience of predominantly male workers in the styrene-butadiene 5 rubber industry, total US rates of mortality from leukemia and total US survival rates (Appendix 5). In 6 order to address this uncertainty. Texas-specific mortality rates for 1999-2003 for all leukemia and Texas-7 specific survival rates for 2003 were kindly provided by the Texas Department of State Health Services, 8 Cancer Epidemiology and Surveillance Branch, Texas Cancer Registry. There were minor differences in 9 calculated air concentrations when Texas versus US all leukemia mortality rates and survival rates were 10 used because the Texas-specific rates are very similar to US rates (Appendix 5). The selected potency estimate is 3.505E-04/ppm using US rates of mortality from leukemia and total US survival rates when 11 12 ADAFS are incorporated (Section 4.2.4.1) and is 3.622E-04/ppm using Texas-specific mortality rates for 13 1999-2003 for all leukemia and Texas-specific survival rates for 2003 when ADAFS are incorporated. 14 15 The adjusted URF of 3.622E-04/ppm using Texas-specific rates is used because, in addition to being

16 slightly more conservative, the residents of Texas are the target population of this DSD. It becomes 3.6E-

17 04/ppm when the URF is rounded to the least number of significant figures for a measured value at the

end of all calculations. The <sup>chronic</sup>ESL<sub>linear(c)</sub> or air concentration at 1 in 100,000 excess cancer risk is 28

19 ppb (62  $\mu$ g/m<sup>3</sup>).

### 20 4.2.5 Uncertainty Analysis

#### 21 4.2.5.1 Use of Mortality Rates to Predict Incidence

The URF (95% UCL) was used instead of the URF (MLE) to account for the uncertainty of calculating a
potency estimate for BD from mortality data, not incidence data (i.e., to protect against developing
leukemia). This is consistent with guidance in TCEQ (2006) and USEPA (2005a). In addition, a linear

25 default was used to extrapolate to lower concentrations, which is a conservative procedure.

26

USEPA (2002) used leukemia incidence rates instead of mortality rates to calculate air concentrations

based on a life-table analyses using the BEIR IV approach (NRC 1988) in an attempt to account for the

uncertainty that potency estimates were based on mortality and not incidence. The BEIR IV methodology

- 30 for calculating excess risk is mathematically correct when the specified response is mortality and
- 31 leukemia mortality rates are used but not when the specified response is mortality and leukemia incidence
- rates are used as was done by USEPA (2002). This error is demonstrated in Appendix 7 *Issues in*

33 *Quantitative Epidemiology: Calculating Excess Risk When Specified Response is Mortality versus* 

*Incidence*. Appendix 7 also shows that if the specified response is incidence, then the BEIR IV

35 methodology for mortality cannot be used. Teta et al. (2004) investigated the validity and implications of

36 using a mortality-based leukemia relative rate model with background leukemia incidence rates, rather

than mortality rates. They concluded that a biased estimate of excess lifetime risk will result, and the

- direction of the bias will vary by potency and the type of leukemia being modeled. Therefore, the TS did
- 39 not use leukemia incidence rates to account for the uncertainty of calculating potency estimates for BD
- 40 from mortality data.

#### 1 4.2.5.2 Adjustments for Other Potentially Sensitive Subpopulations

Leukemia mortality was evaluated based on male workers employed at North American plants that manufactured SBR. It is unknown whether workers with genetic polymorphisms as discussed in Section 3.1.2 (i.e., genes that regulate the metabolism of BD to mutagenic intermediates and genes that regulate the detoxification of those metabolites) were represented in the cohort. Populations with certain lifestyle choices may be more sensitive to health effects caused by BD. Children may also be more sensitive to mutagenic carcinogens (Section 4.2.4.1).

9 Since the UAB cohort was comprised primarily of males, a linear default was used to extrapolate to lower 10 concentrations, and the URF (95% UCL) was used instead of the URF (MLE) to account for the 11 uncertainty of calculating potency estimates for the general population. Studies in which animals were 12 exposed to high BD concentrations suggest that female animals may be more sensitive than male animals

13 for cancer effects after exposure to BD (USEPA 2002). Initial studies conducted in humans by Albertini

et al. (2007) indicate that except for lower production of both urine BD metabolites in females, no female-

15 male differences in response to low BD exposures were detected (mean 8-h TWA exposure levels were

16 0.180 ppm for BD-exposed female workers and 0.370 ppm for BD-exposed male workers as discussed in

17 Section 4.5).

19 The UAB group has analyzed mortality results for 4,863 female workers employed in the SBR industry

20 from 1943 to 2002 (Sathiakumar and Delzell 2007a, b). Preliminary results indicate that standard

21 mortality rates (SMRs) for lung and bladder cancer were elevated in female workers. Both excesses were

- 22 concentrated among ever-hourly employees and among ever-hourly employees with 20+ years since hire,
- 23 but neigher cancer displayed a pattern of increasing SMRs with increasing duration of employment. For

24 lung cancer, analyses of cumulative exposure indices were conducted. Results for lung cancer indicated a

25 moderately positive association with each agent, without exposure-response. The SMRs for leukemia, 26 non-Hodgkin lymphoma or other forms of lymphohematopoietic cancers, breast cancer and ovarian

non-Hodgkin lymphoma or other forms of lymphohematopoietic cancers, breast cancer and ovarian
 cancer were not elevated (Sathiakumar and Delzell 2007b). For lung and bladder cancer, the absence of

any tread of increasing SMRs with increasing duration of employment, the lack of any exposure-response

trend for cumulative exposure to BD, styrene or DMDTC and the absence of positive results in studies of

male employees indicate that these occupational exposures may not have been responsible for the

31 observed excesses of lung and bladder cancers among women in the industry (Sathiakumar and Delzell

- 32 2007b).
- 33

### 34 4.2.5.3 Effect of Occupational Exposure Misclassification

35 One of the limitations of most epidemiological studies is potential exposure misclassification. Health

Canada (2000) and USEPA (2002) expressed concerns about the validity of exposure estimates from the

37 Delzell (1995, 1996) study. In the updated exposure estimates, Macaluso et al. (2004) used a more in-

38 depth job, task, and exposure classification for the cohort, and exposure estimates were developed using 39 exposure modeling, historical exposure data, and plant equipment analysis. Recently, Sathiakumar et al.

exposure modeling, historical exposure data, and plant equipment analysis. Recently, Sathiakumar et al.

(2007) assessed the validity of the BD exposure estimates by measuring the differences and correlations
 between calendar year- and job-specific estimates and measurements of BD concentrations at the

41 between calendar year- and job-specific estimates and measurements of BD concentrations at the 42 Canadian Sarnia plant (a latex operation), one plant included in the UAB cohort. BD measurements from

42 Canadian Sama plant (a fatex operation), one plant included in the OAB conort. BD measurements from
 43 the late 1970s onward were available. Before 1984, estimated concentrations were lower than measured

44 concentrations, whereas after 1984 an opposite pattern was observed. This pattern reversal does not

suggest that exposure values before and after 1984 balance out because exposure estimates before 1984
 were the larger exposures (in absolute value) and contributed more to the estimation of the slopes in the

dose-response models. Increasing the larger exposure estimates will tend to decrease the estimated slopes
 and increase the estimated doses (ppb) corresponding to specified risk levels.

5

6 At lower concentrations, there was reasonably good agreement between measured versus estimated BD

7 exposures; whereas at higher exposures, the estimates tended to be less than the measured values. On

8 average, estimates were about 10% lower than measurements. The Cox regression analysis using mean-

9 scored deciles was used to calculate the URF (95% UCL) to account for concerns about potential

10 exposure misclassification and exposure estimates and as a health-protective policy decision.

### 11 4.2.5.4 Estimated Risks from Occupational Worker

12 There is uncertainty regarding the extrapolation of risks from occupational workers exposed to high BD

13 concentrations and to BD HITs > 100 ppm to risks for the general population who are exposed to much

14 lower BD concentrations and not exposed to BD HITs > 100 ppm. There are no reliable data linking BD

15 exposures at low concentrations typical for the general population to increased mortality from any cause

- 16 in Texas (Grant et al. 2007). The inclusion of age and number of HITs > 100 ppm BD as covariates in the
- 17 Cox regression modeling may result in cancer potency estimates that are more relevant to BD exposures
- experienced by the general population. Figure 3 shows the  $5^{\text{th}}$ ,  $50^{\text{th}}$ , and  $95^{\text{th}}$  percentiles of the distribution of the cumulative number of BD HITs > 100 ppm in the UAB cohort study indicating SBR workers were
- frequently exposed to BD HITs > 100 ppm. In contrast, air monitoring data in Texas do not indicate the
- general population is exposed to BD HITs > 100 ppm. For example, Figure 4 provides 40-min BD
- 22 concentrations (ppbv) at a monitoring site at Milby Park (2005 present) which is located predominantly
- downwind of nearby major industrial sources of BD emissions. There were only four times in a two-year
- 24 period that the concentration of BD exceeded 0.2 ppm and the maximum peak BD concentration was 1.6
- 25 ppm. Maximum 40-min BD concentration data from 25 other ambient air monitoring sites in Texas
- indicate peak concentrations have not approached 1.6 ppm; in fact, maximum concentrations are less than
   0.15 ppm. Other exposure studies indicate that the general population is exposed to concentrations of BD

much lower than occupational workers (USEPA 2002, Gordon et al. 1999; Sapkota and Buckley 2003;

29 Sapkota et al. 2005; Grant et al. 2007).

### 30 4.2.5.5 Dose-Response Modeling

The use of the linear Poisson regression and exponential Cox regression models introduces uncertainty in the dose-response analyses since the MOA of BD is not sufficiently understood to use a more appropriate

biologically-based model. These models are commonly used to investigate dose-response relationships

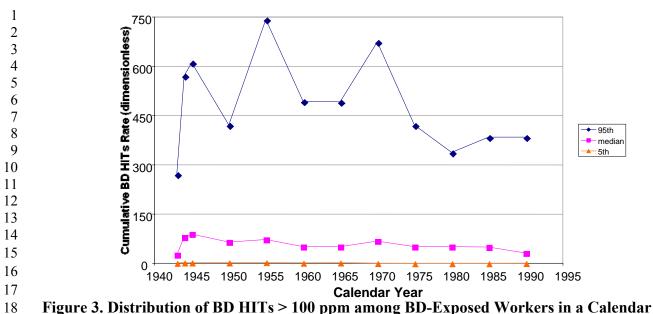
34 derived from occupational cohort epidemiologic studies based on mortality. Both  $\beta$  and upper 95% UCL

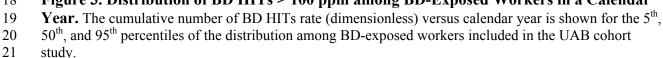
35 estimates were reported in order to provide information on the residual uncertainty in the relative risk

estimates. Generally, there was less than a factor of two difference between  $10^{-5}$ -risk air concentrations calculated with URFs (MLE) versus URFs (95% UCL) except for a four-fold difference for Poisson

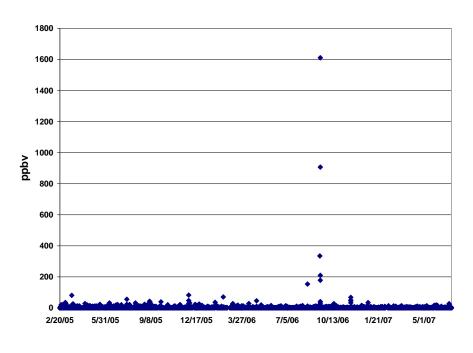
calculated with URFs (MLE) versus URFs (95% UCL) except for a four-fold differ
 regression, mean-scored deciles, age + number of HITs as covariates (Table 16).

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Figure 4. Forty-Minute BD Concentrations (ppbv) at Milby Park (2005 – present). Milby

27 Park is located predominantly downwind of nearby major industrial sources of BD emissions (Grant et al. 2007). Forty minute outpace of predominantly dots

28 2007). Forty-minute auto gas chromatography data.

### 1 4.2.6 Comparison of TCEQ's URF to USEPA's URF

USEPA published an inhalation URF of 0.08/ppm which was last reviewed in 2002. The URF is based on
 a Health Canada analysis of data from Delzell et al. (1995, 1996) using a linear relative rate model and

- 4 was calculated for up to 85 years. Relative risks were evaluated with leukemia incidence rates, which is
- 5 not mathematically correct as demonstrated in Appendix 7. Using the  $LEC_{01}$  (i.e., the 95% lower
- 6 confidence limit of the exposure concentration associated with a 1% increased risk) of 0.254 ppm as the
- 7 POD and a linear extrapolation to zero yields a URF of 0.04/ppm. An adjustment factor of 2 was applied
- 8 to the URF to yield a final URF of 0.08/ppm. This adjustment was applied to reflect evidence from
- 9 studies in mice which suggest that extrapolating leukemia risks from a male-only occupational cohort
- 10 may underestimate the cancer risks for the general public.
- 11
- 12 The TCEQ derived an inhalation URF of 0.00036/ppm based on the most current exposure estimates and
- 13 updated epidemiological study conducted by the UAB group (Macaluso et al. 2004; Sathiakumar et al.
- 14 2005; Graff et al. 2005; HEI 2006). Relative risks were evaluated with Texas-specific leukemia mortality
- rates and survival rates and were calculated for up to 70 years. The URF is based on the 95% UCL
- 16 estimate derived with a exponential Cox regression model, age + number of HITs > 100 ppm as
- 17 covariates, and mean-scored deciles (Cheng et al. 2007). Using the  $LEC_{001}$  (i.e., the 95% lower
- 18 confidence limit of the exposure concentration associated with a 0.1% increased risk) as the POD, a linear
- 19 extrapolation to zero, and adjusting for the increased susceptibility of children using a life-table approach
- 20 (Appendix 6) yields a URF of 0.00036/ppm.

### 21 4.3. Welfare-Based Chronic ESL

22 No data were found regarding long-term vegetative effects.

### 23 4.4 Long-Term ESL and Values for Air Monitoring Evaluation

The chronic evaluation resulted in the derivation of the following values:

<ul> <li>Chronic ReV</li> <li><sup>chronic</sup>ESL<sub>nonlinear(nc)</sub></li> <li>URF</li> <li><sup>chronic</sup>ESL<sub>linear(c)</sub></li> </ul>	= 33 $\mu$ g/m <sup>3</sup> (15 ppb) = 10 $\mu$ g/m <sup>3</sup> (4.5 ppb) = 3.6E-04/ppm (1.6E-04/mg/m <sup>3</sup> ) = 62 $\mu$ g/m <sup>3</sup> (28 ppb)
• ESLlinear(c)	$- 02 \mu g/m (28 \mu p)$

The long-term ESL for air permit reviews is the <sup>chronic</sup>ESL<sub>nonlinear(nc)</sub> of 10  $\mu$ g/m<sup>3</sup> (4.5 ppb) based on ovarian atrophy because it is lower than the <sup>chronic</sup>ESL<sub>linear(c)</sub> of 62  $\mu$ g/m<sup>3</sup> (28 ppb) (Table 1). For evaluation of long-term ambient air monitoring data, the chronic ReV of 33  $\mu$ g/m<sup>3</sup> (15 ppb) based on ovarian atrophy is

- 34 lower than the <sup>chronic</sup>ESL<sub>linear(c)</sub> of 62  $\mu$ g/m<sup>3</sup> (28 ppb), although both values will be used for the evaluation
- of air data as well as the URF of  $3.6E-04/\text{ppm}(1.6E-04/\text{mg/m}^3)$ .

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### 37 4.5 Other Relevant Information

38 The proceedings of the International Symposium on Evaluation of Butadiene and Chloroprene Health

- 39 Risks, held in Charleston, South Carolina on September 20-22, 2005 have recently been published, and
- 40 the findings and results from many of these articles have been cited in the Development Support

1 Document (DSD). Refer to Himmelstein et al. (2007), which provides an excellent summary the main 2 findings of the symposium. A summary of the molecular epidemiology findings from Albertini et al. 3 (2007) as summarized by Himmelstein et al. (2007) is reproduced here because of the significance of their 4 findings. The references, which are in numerical format in the journal, have been supplemented with the 5 author(s) names and year of publication. 6 7 "1.1.3. Molecular epidemiology 8 Albertini [9 (Albertini et al. 2001)] reported that the initial study of workers in the Czech 9 Republic demonstrated a clear no-observed-adverse-effect level (NOAEL) for biomarkers of 10 effect (hypoxanthine-guanine phosphoribosyltransferase (HPRT) mutations and chromosome aberrations) at mean BD exposure concentrations of 0.800 ppm. 11 12 This NOAEL reflects the maximum average exposure level experienced by these workers and 13 was based on extensive external exposure assessments and a comprehensive series of biomarker responses, which included urine metabolites (M1 and M2) and hemoglobin adducts of 14 epoxybutene and EBD (N-[2-dihydroxy-3-butenyl]valine = HB-Val and N- [2,3,4-15 16 trihydroxybutyl]valine = THB-Val, respectively), HPRT mutations, sister-chromatid-exchange 17 frequencies and chromosomal aberrations determined by traditional methods and chromosome 18 painting (fluorescence in situ hybridization). Both the urine metabolite and hemoglobin adduct 19 concentrations proved to be excellent biomarkers of exposure. A second study of Czech 20 workers was conducted at this same facility to compare biomarker responses in female and male employees [10 (Albertini et al. 2007)]. Mean BD exposure concentrations were lower in 21 22 this second study than in the first, being 0.180 ppm and 0.370 ppm for females and males, 23 respectively. Again, there were no BD-associated elevations of HPRT mutation or chromosome 24 aberration frequencies above background in either sex. Similarly, there was no difference 25 between genders in the pattern of BD detoxification, as evidenced by urinary M1 and M2 levels. Females, however, appeared to absorb less BD per unit of exposure, as reflected by 26 27 urine metabolite concentrations. Concentrations of the N,N-(2,3-dihydroxy-1,4-butadiyl)valine 28 (pyr-Val) hemoglobin adduct, which is specific for the highly genotoxic 1,2:3,4-diepoxybutane 29 (DEB) metabolite of BD, were measured in this second study and found to be below the level 30 of quantification for all workers. Later presentations by Swenberg [11 (Swenberg et al. 2007)] 31 and Boysen [12 (Boysen et al. 2007)] in this Symposium described extensive studies of pyr-Val 32 concentrations in BD exposed rodents that, coupled with the results of this Czech worker study, indicate that DEB production in humans is below levels produced in mice or rats exposed to as 33 little as 1.0 ppm BD by inhalation." 34

# 35 Chapter 5. References

#### 36 5.1 References Cited in the Development Support Document

37

Albertini, RJ, RJ Sram, PM Vacek, et al. 2007. Molecular epidemiological studies in 1,3-butadiene
 exposed Czech workers: Female–male comparisons. *Chem Biol Inter* 166: 63-77.

Acute Exposure Guideline Levels (AEGLs). 2005. Acute Exposure Guideline Levels (AEGLs) for 1,3 butadiene (CAS Reg. No. 106-99-0). Interim. Available from: <u>www.epa.gov/oppt/aegl</u>.

1	
2 3	Albertini, RJ, RJ Sram, PM Vacek, et al. 2001. Biomarkers for assessing occupational exposures to 1,3- butadiene. <i>Chem Biol Interact</i> 135-136: 429-53.
4 5 6 7	Alder, N, J Fenty, F Warren, et al. 2006. Meta-analysis of mortality and cancer incidence among workers in the synthetic rubber-producing industry. <i>Am J Epidemiol</i> 164: 405-20.
7 8 9 10	American Conference of Governmental Industrial Hygienists (ACGIH 2001). 1,3-Butadiene. Documentation of the Threshold Limit Values for Chemical Substances 7th Edition. American Conference of Governmental Industrial Hygienists, Inc., Cincinnati, OH.
11 12 13 14	American Chemistry Council (ACC). 2003. An inhalation reproduction/developmental toxicity screening study of 1,3-butadiene in rats. WIL Research Laboratories. OLF-68.0-BD-HPV-WIL.
14 15 16	Arias, E. 2002. United States life tables, 2000. National Vital Statistics Reports. 51(3): p. 3, Table B.
17 18 19	Barnes, DG, GP Daston, JS Evans, et al. 1995. Benchmark dose workshop: criteria for use of a benchmark dose to estimate a reference dose. <i>Regul Toxicol Pharmacol</i> 21: 296-306.
20 21 22	Boysen, G, NI Georgieva, PB Upton, et al. 2007. N-terminal globin adducts as biomarkers for formation of butadiene derived epoxides. <i>Chem Biol Inter</i> 166: 84-92.
23 24 25	Brochot, C, TJ Smith, and FY Bois. 2007. Development of a physiologically based toxicokinetic model for butadiene and four major metabolites in humans: global sensitivity analysis for experimental design issues. <i>Chem Biol Interact</i> in press.
26 27 28 29	Carpenter, CP, CB Shaffer, CSWeil, and HF Smyth, Jr. 1944. Studies on the inhalation of 1:3-butadiene; with a comparison of its narcotic effect with benzol, toluol, and styrene, and a note on the elimination of styrene by the human. <i>J Ind Hyg Toxicol</i> 26: 69-78.
30 31 32	Cheng, H, N Sathiakumar, J Graff, et al. 2004. 1,3-Butadiene and leukemia among synthetic rubber industry workers: Exposure–response relationships. <i>Chem Biol Inter</i> 166:15-24.
33 34 35 36	Christian, MS. 1996. Review of reproductive and developmental toxicity of 1,3-butadiene. <i>Toxicology</i> 113: 137-143.
37 38 39 40	Cochrane, JE and TR Skopek. 1994. Mutagenicity of butadiene and its epoxide metabolites: I. Mutagenic potential of 1,2-epoxybutene, 1,2,3,4-diepoxybutane and 3,4-epoxy-1,2-butanediol in cultured human lymphoblasts. <i>Carcinogenesis</i> 15: 713-17.
41 42 43 44	Csanády, GA, F P Guengerich, and JA Bond. 1992. Comparison of the biotransformation of 1,3- butadiene and its metabolite, butadiene monoepoxide, by hepatic and pulmonary tissues from humans, rats, and mice. <i>Carcinogenesis</i> 13: 1143-53.
44 45 46 47	Delzell, E, N Sathiakumar, and M Macaluso. 1995. A follow-up study of synthetic rubber workers. Final report prepared under contract to International Institute of Synthetic Rubber Producers.

1 2 3	Delzell, E, N Sathiakumar, and M Hovinga. 1996. A follow-up study of synthetic rubber workers. <i>Toxicology</i> 113: 182-189.
4 5	Department of Health and Human Sevices (DHHS). 2000. The ninth report on carcinogens. U.S. Public Health Services, National Toxicology Program, Research Triangle Park, NC.
6 7 8 9	Doerr, JK, SB Hooser, BJ Smith, et al. 1995. Ovarian toxicity of 4-vinylcyclohexene and related olefins in B6C3F1 mice: Role of diepoxides. <i>Chem Res Toxicol</i> 8:963-69
10 11 12	Doerr, JK, EA Hollis, and IG Sipes. 1996. Species difference in the ovarian toxicity of 1,3-butadiene epoxides in B6C3F1 mice and Sprague-Dawley rats. <i>Toxicology</i> 113:128-36.
12 13 14 15 16	Duescher, RJ and AA Elfarra. 1994. Human liver microsomes are efficient catalysts for 1,3-butadiene oxidation: evidence for major roles by cytochrome P450 2A6 and 2E1. <i>Arch Biochem Biophys</i> 311: 342-49.
17 18 19 20	Filipsson, AF, S Sand, J Nilsson, and K Victorin. 2003. The benchmark dose method - review of available models, and recommendations for application in health risk assessment. <i>Crit Rev Toxicol</i> 33: 505- 42.
20 21 22 23	Filser, JG, C Hutzler, V Meischner, et al. 2007. Metabolism of 1,3-butadiene to toxicologically relevant metabolites in single-exposed mice and rats. <i>Chem Biol Inter</i> 166: 93-103.
24 25	Fowles, JR, GV Alexeeff, and D Dodge. 1999. The use of benchmark dose methodology with acute inhalation lethality data. <i>Reg Toxicol Pharmacol</i> 29: 262-278.
26 27 28	Georgieva, NL, G Boysen, P Upton, et al. 2007. Analysis of 1,2;3, 4-diepoxybutane specific protein adduct in occupationally exposed workers. <i>The Toxicologist</i> Abstact #428: 89.
29 30 31 32	Gordon, SM, PJ Callahan, MG Nishioka, et al. 1999. Residential environmental measurements in the national human exposure assessment survey (NHEXAS) pilot study in Arizona: preliminary results for pesticides and VOCs. <i>J Expo Anal Environ Epidemiol</i> 9:456-70.
33 34 35 36	Graff, JJ, N Sathiakumar, M Macaluso, et al. 2005. Chemical exposures in the synthetic rubber industry and lymphohematopoietic cancer mortality. <i>J Occup Environ Med</i> 47:916-32.
37 38 39	Grant, RL, V Leopold, D McCant, and M Honeycutt. 2007. Spatial and temporal trend evaluation of ambient concentrations of 1,3-butadiene and chloroprene in Texas. <i>Chem Biol Inter</i> 166: 44-51.
40 41 42	Green, JW. 2003. Statistical analysis of butadiene mouse data from Hackett et al. (1987) for American Chemistry Council. Laboratory Project ID: Dupont-13474. Sponsor Contract ID: OLF-114.0-BD- stat-DHL. pp 1-151.
43 44 45 46 47	<ul> <li>Hackett, PL, MR Sikov, TJ Mast, et al. 1987a. Inhalation developmental toxicology studies of 1,3- butadiene in the rat (final report). Richland, W.A.: Pacific Northwest Laboratory; PNL Report No. PNL-6414 UC-48; NIH Report No. NIH- 401-ES-410311 101 p. Prepared for NIEHS, NTP, under a Related Services Agreement with the U.S. Department of Energy under contract DE-</li> </ul>

1	AC06-76RLO-1830.
2	
3 4	Hackett, PL, MR Sikov, TJ Mast, et al. 1987b. Inhalation developmental toxicology studies: Teratology study of 1,3-butadiene in mice (final report). Richland, W.A.: Pacific Northwest Laboratory; PNL
5 6	Report No. PNL-6412 UC-48; NIH Report No. NIH- 401-ES-410311 92 p. Prepared for NIEHS, NTP, under a Related Services Agreement with the U.S. Department of Energy under contract
7 8	DE-AC06-76RLO-1830.
9	Health Canada. 2000. Environment Canada, Priority substances list assessment report: 1, 3-butadiene.
10	May 2000. http://www.ec.gc.ca/substances/ese/eng/psap/final/butadiene.cfm, accessed June 25,
11	2007.
12	
13 14 15	Health Effects Institute (HEI). 2006. E Delzell, N Sathiakuman, J Graff, et al. An updated study of mortality among North American synthetic rubber industry workers. Health Effects Institute Research Report Number 132.
16	
17 18	Himmelstein, MW, JF Acquavella, L Recio, et al. 1997. Toxicology and epidemiology of 1,3-butadiene, <i>Crit Rev Toxicol</i> 27: 1-108.
19 20	International Agency for Research on Cancer. 2007. 1,3-Butadiene. Report to be published in 2007 at
20 21 22	following website: <u>http://monographs.iarc.fr/</u>
23 24 25	International Institute of Synthetic Rubber Producers (IISRP). 1982. 1,3-Butadiene: Inhalation teratogenicity in the rat (final report with cover letter dated 08/11/82). Report no. 2788-522/3; submission 8EHQ-0382-0441. Harrowgate, England: Hazleton Laboratories Europe, Ltd.
26 27 28 20	Irons, RD and DW Pyatt. 1998. Dithiocarbamates as potential confounders in butadiene epidemiology. <i>Carcinogenesis</i> 19: 539-42.
29 30 31 32	Irons, RD,WS Stillman, DW Pyatt, et al. 2001. Comparative toxicity of dithiocarbamates and butadiene metabolites in human lymphoid and bone marrow cells. <i>Chem Biol Interact</i> 135-136: 615-25.
33 34 35	Larionov, LF, TA Shtessel', EI Nusel'man. 1934. The physiological action of butadiene, butene-2 and isoprene. <i>Kazanskii Meditsinkii Zhurnal</i> 30:440-45 (HSE translation no. 10855).
36 37 38	Lewis, RJ. 1992. <u>Sax's Dangerous Properties of Industrial Materials</u> , 8th Edition. Van Nostrand Reinhold Company, New York.
39 40 41	Lewis, RJ. 1993. <u>Hawley's Condensed Chemical Dictionary</u> , 12th Edition. Van Nostrand Reinhold Company, New York.
42 43 44	Macaluso, M., R Larson, J Lynch, et al. 2004. Historical estimation of exposure to 1, 3-butdiene, styrene, and dimethyldithiocarbamate among synthetic rubber workers. <i>J Occup Environ Med</i> 1: 371-90.
45 46 47	Miller, LM. 1978. Investigation of selected potential environmental contaminants: Butadiene and its oligomers. Philadelphia, PA: Franklin Research Center. As reported in USEPA 1985.

1 2	Nagata, Y. 2003. Measurement of odor threshold by triangular odor bag method. Odor Measurement Review, Japan Ministry of the Environment. pp. 118-127.
3 4 5 6 7	National Institute for Occupational Safety and Health. 1997. NIOSH pocket guide into 119 carcinogens (http://cdfc.rug.ac.be/HealthRisk/Butadiene/specific_classification/NIOSH.htm). U.S. Department of Health and Human Services, Washington, DC.
7 8 9 10 11	National Research Council (NRC). 1988. Health risks of radon and other internally deposited alpha- emitters. Committee on the biological effects of ionizing radiation. Biological effects of ionizing radiation IV (BEIR IV). Washington DC: National Academy Press.
12 13 14 15 16	<ul> <li>National Toxicology Program (NTP) 1993. NTP technical report on the toxicology and carcinogenesis studies of 1,3-butadiene (CAS No. 106-99-0) in B6C3F1 mice (inhalational studies), NTP TR 434, NIH Publication No. 93-3165, US Department of Health and Human Services Public Health Service. National Institute of Health, Research Triangle Park, NC.</li> </ul>
17 18 19	Occupational Safety and Health Administration (OSHA) 1996. Occupational exposure to 1,3-butadiene Final Rule. 29CFR Part 1910.
20 21 22	Pohl, HR, C Smith-Simon, and H Hicks. 1998. Health effects classification and its role in the derivation of minimal risk levels: Developmental effects. <i>Reg Toxic Pharm</i> 28: 55–60.
22 23 24 25	Preston, RJ. 2007. Cancer risk assessment for 1,3-butadiene: Data integration opportunities. <i>Chem Biol Inter</i> 166: 150-155.
23 26 27 28 29 30	Ripp, GK. 1967. Sanitary validation of the maximum permissible concentration of divinyl in atmosphericair. In: VA Ryazanova (ed.). Biologicheskoe deystvie i gigienicheskoe znachenie atmosfernykh zagryazneniy. Moscow: Izdatel'stvo Meditsina 33-54 (translation prepared for US Environmental Protection Agency PB-212 599).
31 32 33	Ruth, JH. 1986. Odor thresholds and irritation levels of several chemical substances: A review. <i>Am Ind Hyg J</i> 47:A142-A151.
34 35 36 37	Sapkota, A and TJ Buckley. 2003. The mobile source effect on curbside 1,3-butadiene, benzene, and particle-bound polycyclic aromatic hydrocarbons assessed at a tollbooth. <i>J Air Waste Manan Assoc</i> 53: 740-48.
38 39 40	Sapkota, A, D Williams, and TJ Buckley. 2005. Tollbooth workers and mobile source-related hazardous air pollutants: How protective is the indoor environment? <i>Environ Sci Technol</i> 39: 2936-43.
41 42 43	Sathiakumar, N, and E Delzell. 2007a. A follow-up study of women in the synthetic rubber industry: Study methods. <i>Chem Biol Inter</i> 166: 25-28.
44 45 46 47	Sathiakumar, N, and E Delzell. 2007b. A follow-up study of women in the synthetic rubber industry. Draft report submitted to International Institute of Synthetic Rubber Producers (IISRP). Under review.

1 2 3	Sathiakumar, N, J Graff, M Macaluso, et al. 2005. An updated study of mortality among North American synthetic rubber industry workers. <i>Occup Environ Med</i> 62: 822-29.
3 4 5 6 7	Seaton, MJ, MH Follansbee, and JA Bond.1995. Oxidation of 1,2-epoxy-3-butene to 1,2:3,4- diepoxybutane by cDNA-expressed human cytochrome P450 2E1 and 3A4 and human, mouse and rat liver microsomes. <i>Carcinogenesis</i> 16: 2287-92.
8 9 10	Sielken, RL, C Valdez-Flores, ML Gargas, et al. 2007. Cancer risk assessment for 1,3-butadiene: Dose- response modeling from an epidemiological perspective. <i>Chem Biol Inter</i> 166: 140-49.
10 11 12 13	Slikker, Jr, W, ME Andersen, MS Bogdanffy, et al. 2004. Dose-dependent transitions in mechanisms of toxicity: Case studies. <i>Tox Appl Pharm</i> 201: 226-94.
13 14 15 16	Smith, TJ, Y Lin, M Mezzetti, et al. 2001. Genetic and dietary factors affecting human metabolism of 1,3- butadiene. <i>Chem Biol Interact</i> 135-136: 407-28.
17 18	Surveillance, Epidemiology, and End Results (SEER). 2006. Crude total US mortality rates, leukemia, for 1998-2003 by race and sex. <u>www.seer.cancer.gov</u> , accessed June 25, 2007.
19 20 21	Swenberg, JA, G Boysen, N Georgieva, et al. 2007. Future directions in butadiene risk assessment and the role of cross-species internal dosimetry. <i>Chem Biol Inter</i> 166: 78-83.
22 23 24 25	Teta, MJ, NL Tran, PJ Mink, and LM Barraj. 2004. Validity of using background leukemia incidence rates with cohort mortality-based potency estimates to calculate excess lifetime risk . <i>Human and Eco Risk Assess</i> 10: 923-38.
26 27 28 29 30	Texas Risk Reduction Program (TRRP). 2006. Chemical/physical properties table. <u>www.tceq.state.tx.us/assets/public/remediation/trrp/trrptoxchph_2006.xls</u> , accessed June 25, 2007.
31 32	Texas Commission on Environmental Quality (TCEQ). 2006. Guidelines to develop effects screening levels, reference values, and unit risk factors. Chief Engineer's Office. RG-442.
33 34 35 36 37	United States Department of Health, Education, and Welfare (USDHEW). 1970. Chapter 6 Effects of hydrocarbons and certain aldehydes on vegetation in air quality criteria for hydrocarbons. Nat Air Poll Control Admin. Pub. No. AP-64.
38 39 40 41	United States Environmental Protection Agency (USEPA). 1994. Methods for derivation of inhalation reference concentrations and application of inhalation dosimetry. Office of Research and Development. Washington, D.C. EPA/600/8-90/066F.
42 43 44 45 46	United States Environmental Protection Agency (USEPA). 1998. Review of the Office of Research and Development's draft health risk assessment of 1,3-butadiene prepared by the environmental health committee of the Science Advisory Board (SAB), EPA-SAB-EHC-99-003 (November 1998) (available at www.epa.gov/sab/pdf/ehc9903.pdf)

1 2 3	United States Environmental Protection Agency (USEPA). 2000. Benchmark dose technical guidance document. Risk Assessment Forum. Washington, D.C. EPA/630/R-00/001.
4 5 6 7	United States Environmental Protection Agency (USEPA). 2001. National-Scale Air Toxics Assessment (NATA) of emissions from the 1996 National Toxics Inventory (NTI). http://www.epa.gov/ttn/atw/sab/sabrev.html#A1.
8 9 10 11	United States Environmental Protection Agency (USEPA). 2002. Health Assessment of 1,3-Butadiene. EPA/600/P-98/001F. National Center for Environmental Assessment, Office of Research and Development, Washington D.C.
12 13 14	United States Environmental Protection Agency (USEPA). 2005a. Guidelines for carcinogen risk assessment. Risk Assessment Forum. Washington, DC. EPA/630/P-03/001B.
15 16 17	United States Environmental Protection Agency (USEPA). 2005b. Supplemental guidance for assessing susceptibility from early-life exposure to carcinogens. Washington, DC. EPA/630/R-03/003F.
18 19 20 21 22 23	Weincke, JK and KT Kelsey. 1993. Susceptibility to induction of chromosomal damage by metabolites of 1,3-butadiene and its relationship to 'spontaneous' sister chromatid exchange frequencies in human lymphocytes. In Butadiene and styrene: assessment of health hazards, IARC Scientific Publications Vol. 127. (Sorsa, M., Peltonen, K., Vainio, H., et al., eds.). Lyon, France: International Agency for Research on Cancer, pp. 265-273.
24	5.2 Other Studies and Documents Reviewed by the TS
25 26 27 28	Abdel-Rahman, SZ, RA El-Zein, MM Ammenheuser, et al. 2003. Variability in human sensitivity to 1,3- butadiene: Influence of the allelic variants of the microsomal epoxide hydrolase gene. <i>Environ</i> <i>Mol Mutagen</i> 41:140-6.
29 30 31 32	Abdel-Rahman, SZ, MM Ammenheuser, CJ Omiecinski, et al. 2005. Variability in human sensitivity to 1,3-butadiene: Influence of polymorphisms in the 5'-flanking region of the microsomal epoxide hydrolase gene (EPHX1). <i>Toxicol Sci</i> 85: 624-31.
33 34 35	Acquavella, JF, and RC Leonard. 2001. A review of the epidemiology of 1,3-butadiene and chloroprene. <i>Chem Biol Interact</i> 135-136: 43-52.
36 37 38	Adler, ID, J Cao, JG Filser, et al. 1994. Mutagenicity of 1,3-butadiene inhalation in somatic and germinal cells of mice. <i>Mutat Res</i> 309:307-14.
39 40 41 42	Adler, ID, J Filser, H Gonda, and G Schriever-Schwemmer. 1998. Dose response study for 1,3-butadiene- induced dominant lethal mutations and heritable translocations in germs cells of male mice. <i>Mutat</i> <i>Res</i> 397: 85-92.
43	Adler, ID and D Anderson. 1994. Dominant lethal effects after inhalation exposure to 1,3-butadiene.

1 2 3	Agency for Toxic Substances and Disease Registry (ATSDR). 1992. Toxicological profile for 1,3- butadiene. Available from ATSDR, Public Health Service, U.S. Department of Health and Human Services, Atlanta, GA. Available from: www.atsdr.cdc.gov/toxprofiles/tp28.html.
4	
5 6 7 8	Albertini, R, H Clewell, MW Himmelstein, et al. 2003. The use of non-tumor data in cancer risk assessment: Reflections on butadiene, vinyl chloride, and benzene. <i>Regul Toxicol Pharmacol</i> 37:105-32.
9 10 11 12	Ammenheuser, MM, WE Bechtold, SZ Abdel-Rahman, et al. 2001. Assessment of 1,3-butadiene exposure in polymer production workers using HPRT mutations in lymphocytes as a biomarker. <i>Environ Health Perspect</i> 109: 1249-55.
13 14 15	Anderson, D. 2001. Genetic and reproductive toxicity of butadiene and isoprene. <i>Chem Biol Interact</i> 135-136: 65-80.
15 16 17	Anderson, D. 2005. Male-mediated developmental toxicity. Tox Appl Pharm 207: S506-13.
18 19 20	Anderson, D, AJ Edwards, MH Brinkworth and JA Hughes. 1996. Male-mediated F1 effects in mice exposed to 1,3-butadiene. <i>Toxicology</i> 113: 120-27.
21 22 23 24	Anderson, D, MH Brinkworth, TW Yu, et al. 1997. Somatic and germ cell effects in rats and mice after treatment with 1,3-butadiene and its metabolites, 1,2-epoxybutene and 1,2,3,4-diepoxybutane. <i>Mutat Res</i> 391: 233-42.
25 26 27	Anderson, D, JA Hughes, AJ Edwards, and MH Brinkworth. 1998. A comparison of male-mediated effects in rats and mice exposed to 1,3-butadiene. <i>Mutat Res</i> 397: 77-84.
28 29 30	Anderson, D. 1998. Butadiene: Species comparison for metabolism and genetic toxicology. <i>Mutat Res</i> 405: 247-58.
31 32 33	Baker, J, J Arey, and R Atkinson. 2005. Formation and reaction of hydroxycarbonyls from the reaction of OH radicals with 1,3-butadiene and isoprene. <i>Environ Sci Technol</i> 39: 4091-9.
34 35 36 37	Barshteyn, N, RJ Krause and AA Elfarra. 2007. Mass spectral analyses of hemoglobin adducts formed after in vitro exposure of erythrocytes to hydroxymethylvinyl ketone. <i>Chem Biol Interact</i> 166: 176-181.
38 39 40 41	Bechtold, WE, MR Strunk, IY Chang, et al. 1994. Species differences in urinary butadiene metabolites: Comparisons of metabolite ratios between mice, rats, and humans. <i>Toxicol Appl Pharmacol</i> 127: 44-9.
41 42 43 44 45	Begemann, P, PB Upton, A Ranasinghe, et al. 2001. Hemoglobin adducts as biomarkers of 1,3-butadiene in occupationally low exposed Italian workers and a few diesel-exposed miners. <i>Chem Biol Interact</i> 135-136: 675-678.

1 2 3	Bernardini, S, K Pelin, K Peltonen, et al. 1996. Induction of sister chromatid exchange by 3,4- expoxybutane-1,2-diol in cultured human lymphocytes of different GSTT1 and GSTM1 genotypes. <i>Mutat Res</i> 361: 121-7.
4 5 6 7	Bernardini, S, A Hirvonen, K Pelin, and H Norppa. 1998. Induction of sister chromatid exchange by 1,2- epoxy-3-butene in cultured human lymphocytes: Influence of GSTT1 genotype. Carcinogenesis 19: 377-80.
8 9 10 11	Bird, MG, JM Rice, and JA Bond. 2001. Evaluation of 1,3-butadiene, isoprene and chloroprene health risks. <i>Chem Biol Interact</i> 135-136: 1-7.
12 13 14	Bird, MG, DFV Lewis, FT Whitman, et al. 2001. Application of process chemistry and SAR modelling to the evaluation of health findings of lower olefins. <i>Chem Biol Interact</i> 135-136: 571-84.
15 16 17	Bois, FY, TJ Smith, A Gelman, et al. 1999. Optimal design for a study of butadiene toxicokinetics in humans. <i>Toxicol Sci</i> 49: 213-24.
18 19 20	Bond, JA and MA Medinsky. 2001. Insights into the toxicokinetics and toxicodynamics of 1,3-butadiene. <i>Chem Biol Interact</i> 135-136: 599-614.
20 21 22 23 24	Bond, JA, MW Himmelstein, M Seaton, et al. 1996. Metabolism of butadiene by mice, rats, and humans: A comparison of physiologically based toxicokinetic model predictions and experimental data. <i>Toxicology</i> 113: 48-54.
25 26	Bond, JA, L Recio, and MA Medinsky. 1997, Letter to the editor: Importance of mechanistic data in human health assessment, <i>J Clean Technol Environ Toxicol Occup Med</i> 6: 101-6.
27 28 29 30	Bond, JA, L Recio, and D Andjelkovich. 1995. Epidemiological and mechanistic data suggest that 1,3- butadiene will not be carcinogenic to humans at exposures likely to be encountered in the environment or workplace. <i>Carcinogenesis</i> 16: 165-71.
31 32 33 34 35	Boogaard, PJ, KP de Kloe, ED Booth, and WP Watson. 2004. DNA adducts in rats and mice following exposure to [4-[1][4]C]-1,2-epoxy-3-butene and to [2,3-[1][4]C]-1,3-butadiene. <i>Chem Biol Interact</i> 148: 69-92.
36 37 38 39	Boogaard, PJ, NJ van Sittert, and HJJJ Megens. 2001. Urinary metabolites and haemoglobin adducts as biomarkers of exposure to 1,3-butadiene: A basis for 1,3-butadiene cancer risk assessment. <i>Chem Biol Interact</i> 135-136: 695-701.
40 41 42 43	Boogaard, PJ, NJ van Sittert, WP Watson and KP de Kloe. 2001. A novel DNA adduct, originating from 1,2-epoxy-3,4-butanediol, is the major DNA adduct after exposure to [2,3-14C]-1,3-butadiene, but not after exposure to [4-14C]-1,2-epoxy-3-butene. <i>Chem Biol Interact</i> 135-136: 687-693.
43 44 45 46 47	Booth, ED, JD Kilgour, and WP Watson. 2004. Dose responses for the formation of hemoglobin adducts and urinary metabolites in rats and mice exposed by inhalation to low concentrations of 1,3-[2,3-(14)C]-butadiene. <i>Chem Biol Interact</i> 147: 213-32.

1 2 2	Booth, ED, JD Kilgour, SA Robinson, and WP Watson. 2004. Dose responses for DNA adduct formation in tissues of rats and mice exposed by inhalation to low concentrations of 1,3-[2,3- <sup>14</sup> C]-butadiene.
3 4	Chem Biol Interact 147: 195-211.
5 6	Boysen, G, NI Georgieva, PB Upton, et al. 2004. Analysis of diepoxide-specific cyclic n-terminal globin adducts in mice and rats after inhalation exposure to 1,3-butadiene. <i>Cancer Res</i> 64: 8517-20.
7 8 9	Boysen, G, CO Scarlett, B Temple, et al. 2007. Identification of covalent modifications in P450 2E1 by 1,2-epoxy-3-butene in vitro. <i>Chem Biol Interact</i> 166: 170-75.
10 11 12 13	Brinkworth, MH, D Anderson, JA Hughes, et al. 1998. Genetic effects of 1,3-butadiene on the mouse testis. <i>Mutat Res</i> 397: 67-75.
14 15 16	Brondeau MT, A Hesbert, C Beausoleil, and O Schneider. 1999. To what extent are biomonitoring data available in chemical risk assessment? <i>Hum Exp Toxicol</i> 18: 322-6.
17 18	Catallo, WJ, CH Kennedy, W Henk, et al. 2001. Combustion products of 1,3-butadiene are cytotoxic and genotoxic to human bronchial epithelial cells. <i>Environ Health Perspect</i> 109: 965-71.
19 20 21	Crouch, CN, DH Pullinger, and IF Gaunt. 1979. Inhalation toxicity studies with 1,3-butadiene 2. 3 month toxicity study in rats. <i>Am Ind Hyg Assoc J</i> 40: 796-802.
22 23 24 25 26	Csanády, GA, PE Kreuzer, C Baur, and JG Filser. 1996. A physiological toxicokinetic model for 1,3- butadiene in rodents and man: Blood concentrations of 1,3-butadiene, its metabolically formed epoxides, and of haemoglobin adductsrelevance of glutathione depletion. <i>Toxicology</i> 113: 300- 5.
27 28 29 30	Dahl, AR, and RF Henderson. 2000. Comparative metabolism of low concentrations of butadiene and its monoepoxide in human and monkey hepatic microsomes. <i>Inhal Toxicol</i> 12: 439-51.
31 32	Dahl, AR, WE Bechtold, JA Bond, et al. 1990. Species differences in the metabolism and disposition of inhaled 1,3-butadiene and isoprene. <i>Environ Health Perspect</i> 86: 65-9.
33 34 35 36 37	Dale, M. Walker, JD McDonald, Q Meng, et al. 2007. Measurement of plasma or urinary metabolites and Hprt mutant frequencies following inhalation exposure of mice and rats to 3-butene-1,2-diol. <i>Chem Biol Interact</i> 166: 191-206.
37 38 39 40 41	Delzell, E, M Macaluso, N Sathiakumar, and R Matthews. 2001. Leukemia and exposure to 1,3- butadiene, styrene and dimethyldithiocarbamate among workers in the synthetic rubber industry. <i>Chem Biol Interact</i> 135-136: 515-34.
42 43 44	Divine, BJ and CM Hartman. 2001. A cohort mortality study among workers at a 1,3 butadiene facility. <i>Chem Biol Interact</i> 135-136: 535-53.
45 46 47	Dollard, G J, CJ Dore, and ME Jenkin. 2001. Ambient concentrations of 1,3-butadiene in the UK. <i>Chem</i> <i>Biol Interact</i> 135-136: 177-206.

1 2 3	Dorr, DQ, K Murphy and N Tretyakova. 2007. Synthesis of DNA oligodeoxynucleotides containing structurally defined N6-(2-hydroxy-3-buten-1-yl)-adenine adducts of 3,4-epoxy-1-butene. <i>Chem Biol Interact</i> 166: 104-11.
4	
5 6	Doyle, M, KG Sexton, H Jeffries and I Jaspers. 2007. Atmospheric photochemical transformations enhance 1,3-butadiene-induced inflammatory responses in human epithelial cells: The role of
7 8	ozone and other photochemical degradation products. Chem Biol Interact 166: 163-69.
9	Doyle, M, KG Sexton, H Jeffries, et al. 2004. Effects of 1,3-butadiene, isoprene, and their photochemical
10	degradation products on human lung cells. <i>Environ Health Perspect</i> 112: 1488-95.
11	Durachen DL and AA Elfanza 1002 Chlanamaridara madiatad aridation of 1.2 hatadiana ta 2 hatanal
12 13	Duescher, RJ, and AA Elfarra. 1993. Chloroperoxidase-mediated oxidation of 1,3-butadiene to 3-butenal, a crotonaldehyde precursor. <i>Chem Res Toxicol</i> 6: 669-73.
14	
15 16	Elfarra, AA, RJ Krause, and RA Kemper. 2001. Cellular and molecular basis for species, sex and tissue differences in 1,3-butadiene metabolism. <i>Chem Biol Interact</i> 135-136: 239-48.
17	
18	Evelo, CT, JG Oostendorp, WF ten Berge, and PJ Borm. 1993. Physiologically based toxicokinetic
19	modeling of 1,3-butadiene lung metabolism in mice becomes more important at low doses.
20 21	Environ Health Perspect 101: 496-502.
22 23	Farmer, PB. 2004. DNA and protein adducts as markers of genotoxicity. <i>Toxicol Lett</i> 149: 3-9.
24 25	Fernandes, PH, LC Hackfeld, ID Kozekov, et al. 2006. Synthesis and mutagenesis of the butadiene- derived n3 2'-deoxyuridine adducts. <i>Chem Res Toxicol</i> 19: 968-76.
26 27 28 20	Filser, JG, TH Faller, S Bhowmik, et al. 2001. First-pass metabolism of 1,3-butadiene in once-through perfused livers of rats and mice. <i>Chem Biol Interact</i> 135-136: 249-65.
29 30 31 32	Filser, JG, G Johanson, W Kessler, et al. 1993. A pharmacokinetic model to describe toxicokinetic interactions between 1,3-butadiene and styrene in rats: Predictions for human exposure. <i>IARC Scientific Publications</i> 127: 65-78.
33 34 35 36	Fraser, I. 2001. Butadiene – progress under the European Union Existing Substances Regulation. <i>Chem</i> <i>Biol Interact</i> 135-136: 103-107.
37 38 39	Fred, C, A Kautiainen, I Athanassiadis, and M Tornqvist. 2004. Hemoglobin adduct levels in rat and mouse treated with 1,2:3,4-diepoxybutane. <i>Chem Res Toxicol</i> 17: 785-94.
40 41 42	Fustinoni, S, L Soleo, M Warholm, et al. 2002. Influence of metabolic genotypes on biomarkers of exposure to 1,3-butadiene in humans. <i>Cancer Epidemiology Biomarkers &amp; Prev</i> 11: 1082-1090.
43 44 45	Fustinoni, S, L Perbellini, L Soleo, et al. 2004. Biological monitoring in occupational exposure to low levels of 1,3-butadiene. <i>Tox Letters</i> 149: 353-60.
43 46 47	Genter, MB, and L Recio. 1994, Absence of detectable P450 2E1 in bone marrow of B6C3F1 mice: Relevance to butadiene-induced bone marrow toxicity. <i>Fundam Appl Toxicol</i> 22: 469-73.

1	
2	Georgieva, NI, G Boysen, PB Upton, et al. 2007. Quantitative analysis of N-terminal valine peptide
3	adducts specific for 1,2-epoxy-3-butene. Chem Biol Interact 166: 219-25.
4	
5	Green, T, A Toghill, and R Moore. 2001. The influence of co-exposure to dimethyldithiocarbamate on
6	butadiene metabolism. Chem Biol Interact 135-136: 585-98.
7	
8	Goggin, M, R Loeber, S Park, et al. 2007. HPLC-ESI+-MS/MS analysis of N7-guanine-N7-guanine DNA
9	cross-links in tissues of mice exposed to 1,3-butadiene. Chem Res Toxicol 20: 839-47.
10	
11	Hackett, PL, BJ McClanahan, TJ Mast, et al. 1988b. Dominant lethal study in CD-1 mice following
12	inhalation exposure to 1,3-butadiene (final technical report). Richland, W.A.: Pacific Northwest
13	Laboratory; PNL Report No. PNL-6545 UC-408; NIH Report No. NIH-Y01-ES-70153; 85 p.
14	Prepared for NIEHS, NTP, under a Related Services Agreement with the U.S. Department of
15	Energy under contract DE-AC06-76RLO-1830.
16	
17	Hackett, PL, BJ McClanahan, MG Brown, et al. 1988a. Sperm-head morphology study in B6C3F1 mice
18	following inhalation exposure to 1,3-butadiene (final technical report). Richland, W.A.: Pacific
19	Northwest Laboratory; PNL Report No. PNL-6459 UC-48; NIH Report No. NIH-Y01-ES-70153;
20	51 p. Prepared for NIEHS, NTP, under a Related Services Agreement with the U.S. Department
21	of Energy under contract DE-AC06-76RLO-1830.
22 23	Hallberg, LM, WE Bechtold, J Grady, et al. 1997. Abnormal DNA repair activities in lymphocytes of
23 24	workers exposed to 1,3-butadiene. <i>Mutat Res</i> 383: 213-21.
24	workers exposed to 1,3-butadiene. Mutat Res 585. 213-21.
26	Hallenbeck, WH. 1992. Cancer risk assessment for the inhalation of 1,3-butadiene using PBPK modeling.
27	Bulletin Environ Cont Tox 49: 66-70.
28	Dute tin Environ Control 47.00-70.
29	Hayes, RB, L Zhang, JA Swenberg, et al. 2001. Markers for carcinogenicity among butadiene-polymer
30	workers in China. Chem Biol Interact 135-136:455-64.
31	
32	Hayes, RB, L Zhang, S Yin, et al. 2000. Genotoxic markers among butadiene polymer workers in China.
33	Carcinogenesis 21: 55-62.
34	
35	Health Effects Institute (HEI). 1999. A partnership to examine emerging health effects: EC/HEI
36	workshop on 1,3-butadiene. Health Effects Institute Communications Number 6.
37	
38	Health Effects Institute (HEI). 2000. 1,3-Butadiene: Cancer, mutations, and adducts. Health Effects
39	Institute Research Report Number 92.
40	
41	Health Effects Institute (HEI). 2003. Albertini RJ, RJ Sram, PM Vacek, et al. Biomarkers in Czech
42	workers exposed to 1,3-butadiene: A transitional epidemiologic study. Health Effects Institute
43	Research Report Number 116.
44	
45	Henderson, RF, FF Hahn, EB Barr, et al. 1999. Carcinogenicity of inhaled butadiene diepoxide in female
46	B6C3F1 mice and Sprague-Dawley rats. Toxicol Sci 52: 33-44.
47	

1 2	Henderson, RF, FF Hahn, JM Benson, et al. 1999. Dosimetry and acute toxicity of inhaled butadiene diepoxide in rats and mice. <i>Toxicol Sci</i> 51: 146-52.
3 4 5	Henderson, RF 2001. Species differences in the metabolism of olefins: implications for risk assessment. <i>Chem Biol Interact</i> 135-136: 53-64.
5 6	Chem Biol Interact 155-150. 55-04.
7 8	Henderson, RF, WE Bechtold, JR Thornton-Manning, AR Dahl. 2001. Urinary butadiene diepoxide: a potential biomarker of blood diepoxide. Toxicology 160:81-6.
8 9	potential biomarker of blood diepoxide. Toxicology 100.81-0.
10 11	Higashino, H, K Mita, H Yoshikado, et al. 2007. Exposure and risk assessment of 1,3-butadiene in Japan. <i>Chem Biol Interact</i> 166: 52-62.
12	
13 14	Himmelstein, MW, RA Baan, RJ Albertini, et al. 2007. International Symposium on the Evaluation of Butadiene and Chloroprene Health Risks. <i>Chem Biol Interact</i> 166: 1-9.
15 16 17	Hoyer, PB, PJ Devine, X Hu, et al. 2001. Ovarian toxicity of 4-vinylcyclohexene diepoxide: A mechanistic model. <i>Toxicol Pathol</i> 29: 91-9.
18 19 20	Hughes, K, ME Meek, M Walker. 2001. Health risk assessment of 1,3-butadiene as a Priority Substance in Canada. <i>Chem Biol Interact</i> 135-136:109-35.
21 22 23	Hughes, K, ME Meek, M Walker, R Beauchamp. 2003. 1,3-Butadiene: Exposure estimation, hazard characterization, and exposure-response analysis. <i>J Toxicol Environ Health B Crit Rev</i> 6:55-83.
24 25 26	Iba, MM and MG Bird. 2007. Effect of n-hexane on the disposition and toxicity of the 1,3-butadiene metabolite 3-butene-1,2-diol. <i>Chem Biol Interact</i> 166: 232-38.
27 28 29	Jackson, TE, PD Lilly, L Recio, et al. 2000. Inhibition of cytochrome P450 2E1 decreases, but does not eliminate, genotoxicity mediated by 1,3-butadiene. <i>Toxicol Sci</i> 55: 266-73.
30 31 32 33	Johanson, G, and JG Filser. 1993. A physiologically based pharmacokinetic model for butadiene and its metabolite butadiene monoxide in rat and mouse and its significance for risk extrapolation. <i>J Arch Toxicol</i> 67: 151-163.
34 35 36	Johanson, G, and JG Filser. 1996. PBPK model for butadiene metabolism to epoxides: Quantitative species differences in metabolism. <i>Toxicology</i> 113: 40-7.
37 38 39	Kemper, RA, RJ Krause, and AA Elfarra. 2001. Metabolism of butadiene monoxide by freshly isolated hepatocytes from mice and rats: Different partitioning between oxidative, hydrolytic, and
40	conjugation pathways. Drug Metab Dispos 29: 830-6.
41 42 43	Kim, Y, HH Hong, Y Lachat, et al. 2005. Genetic alterations in brain tumors following 1,3-butadiene exposure in B6C3F1 mice. <i>Toxicol Pathol</i> 33: 307-12.
43 44	exposure in boest 1 mile. <i>Toxicol 1 unol 55</i> , 507-12.
45 46 47	Kim, MY, N Tretyakova, and GN Wogan. 2007. Mutagenesis of the supF gene by stereoisomers of 1,2,3,4-diepoxybutane. Chem Res Toxicol 20: 790-97.

1 2 3	Kirman, CR and ML Gargas. 2006. Benchmark dose analyses for reproductive and developmental endpoints for 1,3-butadiene. Submitted to Olefins Panel, American Chemistry Council, Arlington, VA.
4	
5 6	Kligerman, AD, and Y Hu. 2007. Some insights into the mode of action of butadiene by examining the genotoxicity of its metabolites. <i>Chem Biol Interact</i> 166: 132-139.
7	
8 9	Kohn, MC. 1997. The importance of anatomical realism for validation of physiological models of disposition of inhaled toxicants. <i>Tox Appl Pharm</i> 147: 448-458.
10	
11 12 13	Kohn, MC, and RL Melnick. 1993. Species differences in the production and clearance of 1,3-butadiene metabolites: A mechanistic model indicates predominantly physiological, not biochemical control. <i>Carcinogenesis</i> 14: 619-28.
	control. Carcinogenesis 14. 019-28.
14 15 16	Kohn, MC, and RL Melnick. 1996. Effects of the structure of a toxicokinetic model of butadiene inhalation exposure on computed production of carcinogenic intermediates. <i>Toxicology</i> 113: 31-9.
17 18 19	Kohn, MC, and RL Melnick. 2000. The privileged access model of 1,3-butadiene disposition. <i>Environ Health Perspect</i> 108 Suppl 5: 911-7.
20	
21 22 23	Kohn, MC, and RL Melnick. 2001. Physiological modeling of butadiene disposition in mice and rats. <i>Chem Biol Interact</i> 135-136: 285-301.
24 25 26	Koivisto, P and K Peltonen. 2001. N7-guanine adducts of the epoxy metabolites of 1,3-butadiene in mice lung. <i>Chem Biol Interact</i> 135-136: 363-372.
27 28 29	Koivisto, P, ID Adler, F Pacchierotti, and K Peltonen. 1998. DNA adducts in mouse testis and lung after inhalation exposure to 1,3-butadiene. <i>Mutat Res</i> 397: 3-10.
29 30 31 32	Koppikar, AM. 2001. Future research needs for the health assessment of 1,3-butadiene. <i>Chem Biol Interact</i> 135-136: 629-36.
32 33 34 35	Leavens, TL, and JA Bond. 1996. Pharmacokinetic model describing the disposition of butadiene and styrene in mice. <i>Toxicology</i> 113: 310-13.
36 37 38 39	Lee, JH, HS Kang, D Han. 2005. Ratios of N-(2,3,4-trihydroxybutyl) valine and N-(2-hydroxy-3-butenyl) valine formed hemoglobin adducts in female mice inhalation exposure with 1,3-butadiene. <i>Tox Ind Health</i> 21: 15-20.
40 41 42	Legator, MS. 1997. Response to letter to the editor. <i>J Clean Technol Environ Toxicol Occup Med</i> 6: 107-12.
43 44 45	Legator, MS. 1997. Underestimating risk for three important human carcinogens: Vinyl chloride, benzene, and butadiene. <i>Ann N Y Acad Sci</i> 837: 170-5.
43 46 47	Lin, Y, TJ Smith, D Wypij, et al. 2002. Association of the blood/air partition coefficient of 1,3-butadiene with blood lipids and albumin. <i>Environ Health Pers</i> 110: 165-68.

1	
2 3	Lindbohm, ML, K Hemminki, P Kyyronen, et al. 1983. Spontaneous abortions among rubber workers and congenital malformations in their offspring. <i>Scand J Work Environ Health</i> 9 <i>Suppl</i> 2:85-90.
4	
5 6 7	Loughlin, JE, KJ Rothman, and NA Dreyer. 1999. Lymphatic and haematopoietic cancer mortality in a population attending school adjacent to styrene-butadiene facilities, 1963-1993. <i>J Epidemiol Community Health</i> 53: 283-7.
8	
9 10	Lynch, J. 2001. Occupational exposure to butadiene, isoprene and chloroprene. <i>Chem Biol Interact</i> 135-136: 207-14.
11	
12 13	Macaluso, M, R Larson, E Delzell. 1996. Leukemia and cumulative exposure to butadiene, styrene and benzene among workers in the synthetic rubber industry. <i>Toxicology</i> 113: 190-202.
14 15 16	Maniglier-Poulet, C, X Cheng, JA Ruth, and D Ross. 1995. Metabolism of 1,3-butadiene to butadiene monoxide in mouse and human bone marrow cells. <i>Chem Biol Interact</i> 97: 119-29.
17 18 19 20	Medinsky, MA, TL Leavens, GA Csanády, et al. 1994. In vivo metabolism of butadiene by mice and rats: A comparison of physiological model predictions and experimental data. <i>Carcinogenesis</i> 15: 1329-40.
21 22 23 24 25	Mehlman, MA, and MS Legator. 1991. Dangerous and cancer-causing properties of products and chemicals in the oil refining and petrochemical industry - Part II: Carcinogenicity, mutagenicity, and developmental toxicity of 1,3-butadiene. <i>Toxicol Ind Health</i> 7: 207-20.
26 27	Melnick, RL, and RC Sills. 2001. Comparative carcinogenicity of 1,3-butadiene, isoprene, and chloroprene in rats and mice. <i>Chem Biol Interact</i> 135-136: 27-42.
28 29 30 31	Meng, Q, RF Henderson, L. Long, et al. 2001. Mutagenicity at the Hprt locus in T cells of female mice following inhalation exposures to low levels of 1,3-butadiene. <i>Chem Biol Interact</i> 135-136: 343- 61.
32 33 34 35	Meng Q, DM Walker, BR Scott, et al. 2004. Characterization of Hprt mutations in cDNA and genomic DNA of T-cell mutants from control and 1,3-butadiene-exposed male B6C3F1 mice and F344 rats. <i>Environ Mol Mutagen</i> 43:75-92.
36 37 38 39	Meng, Q, DL Redetzke, LC Hackfeld, et al. 2007. Mutagenicity of stereochemical configurations of 1,2- epoxybutene and 1,2:3,4-diepoxybutane in human lymphblastoid cells. <i>Chem Biol Interact</i> 166: 207-18.
40 41 42	Meng, Q., DM Walker, JD McDonald, et al. 2007. Age-, gender-, and species-dependent mutagenicity in T cells of mice and rats exposed by inhalation to 1,3-butadiene. <i>Chem Biol Interact</i> 166: 121-131.
43 44 45 46 47	Merritt, WK, A Kowalczyk, T Scholdberg et al. 2005. Dual roles of glycosyl torsion angle conformation and stereochemical configuration in butadiene oxide-derived N1 β-hydroxyalkyl deoxyinosine adducts : A structural perspective. <i>Chem Res Toxicol</i> 18: 1098-1107.

1 2 3	Moll, TS, AC Harms, and AA Elfarra. 2001. Advances in the mass spectrometry of hemoglobin adducts: global analysis of the covalent binding of butadiene monoxide. <i>Chem Biol Interact</i> 135-136: 667-674.
4	
5 6 7	Morrissey, RE, BA Schwetz, PL Hackett, et al. 1990, Overview of reproductive and developmental toxicity studies of 1,3-butadiene in rodents. <i>Environ Health Perspect</i> 86: 79-84.
7 8	Morrow NI 2001 Significance of 1.2 but diana to the US air toxics regulatory effort Cham Piel
o 9	Morrow, NL. 2001. Significance of 1,3-butadiene to the US air toxics regulatory effort. <i>Chem Biol Interact</i> 135-136: 137-143.
9 10	Interact 155-150. 157-145.
11	Murphy, CF and DT Allen. 2005. Hydrocarbon emissions from industrial release events in the Houston-
12 13	Galveston area and their impact on ozone formation. <i>Atmos Environ</i> 39: 3785–98.
14 15	National Institute Occupation Safety and Health (NIOSH). 1996. NIOSH Study of the Health Effects of Exposure to 1,3-butadiene. Available from:
16	http://www.cdc.gov/niosh/pgms/worknotify/butadiene.html.
17	
18	Office of Environmental Health Hazard Assessment (OEHHA). 2005. Air Toxics Hot Spots Program
19	Risk Assessment Guidelines, Part II, Technical Support Document for Describing Available
20	Cancer Potency Factors. May 2005. 1,3-butadiene. B116-B123. Available from:
21	http://www.oehha.org/air/hot_spots/pdf/May2005Hotspots.pdf.
22 23	Office of Environmental Health Hazard Assessment (OEHHA). 2000. Chronic toxicity summary 1,3-
23 24	butadiene. Available from: http://www.oehha.org/air/chronic_rels/pdf/106990.pdf.
25	
26 27	Onaran, I, A Ozaydin, F Akbas, et al. 2000. Are individuals with glutathione S-transferase GSTT1 null genotype more susceptible to in vitro oxidative damage? <i>J Toxicol Environ Health A</i> 59: 15-26.
28	
29 30	Ottensmeier, C. 2001. The classification of lymphomas and leukemias. <i>Chem Biol Interact</i> 135-136: 653-64.
31	Developmenti D. C. Timmer, D. Develdi et al. 1000. Deven develop terrisite eff. 2 herte diversite the mercer
32	Pacchierotti, F, C Tiveron, R Ranaldi, et al. 1998. Reproductive toxicity of 1,3-butadiene in the mouse:
33	Cytogenetic analysis of chromosome aberrations in first-cleavage embryos and flow cytometric evaluation of spermatogonial cell killing. <i>Mutat Res</i> 397: 55-66.
34 35	evaluation of spermatogonial cen kining. Mulai Res 397. 33-66.
33 36	Pacchierotti, F, ID Adler, D Anderson, et al. 1998. Genetic effects of 1,3-butadiene and associated risk for
30 37	heritable damage. <i>Mutat Res</i> 397:93-115.
38	licittable damage. Mutat Kes 337.93-115.
39	Park, S, J Hodge, C Anderson, and N Tretyakova. 2004. Guanine-adenine DNA cross-linking by 1,2,3,4-
40	diepoxybutane: Potential basis for biological activity. <i>Chem Res Toxicol</i> 17: 1638-51.
40 41	depoxybutane. Totential basis for biological activity. Chem Res Toxicol 17. 1056-51.
42	Powley, MW, K Jayaraj, A Gold, et al. 2003. 1,N <sup>2</sup> -propanodeoxyguanosine adducts of the 1,3-butadiene
43	metabolite, hydroxymethylvinyl ketone. <i>Chem Res Toxicol</i> 16:1448-54.
44	
45	Powley, MW, Y Li, PB Upton, et al. 2005. Quantification of DNA and hemoglobin adducts of 3,4-epoxy-
46	1,2-butanediol in rodents exposed to 3-butene-1,2-diol. Carcinogenesis 26:1573-80.
47	

1 2 3 4	Powley, MW, Vernon E. Walker, Yutai Li, Patricia B. Upton and James A. Swenberg. 2007. The importance of 3,4-epoxy-1,2-butanediol and hydroxymethylvinyl ketone in 3-butene-1,2-diol associated mutagenicity. <i>Chem Biol Interact</i> 166: 182-190.
5 6 7	Recio, L, A Steen, LJ Pluta, et al. 2001. Mutational spectrum of 1,3-butadiene and metabolites 1,2- epoxybutene and 1,2,3,4-diepoxybutane to assess mutagenic mechanisms. <i>Chem Biol Interact</i> 135-136: 325-41.
8 9 10 11	Reiss, R. 2006. Temporal trends and weekend-weekday differences for benzene and 1,3-butadiene in Houston, Texas. <i>Atmos Environ</i> 40: 4711-24.
12 13 14 15	Rice, JM, and P Boffetta. 2001. 1,3-Butadiene, isoprene and chloroprene: Reviews by the IARC monographs programme, outstanding issues, and research priorities in epidemiology. <i>Chem Biol Interact</i> 135-136: 11-26.
16 17 18 19	Richardson, KA, MM Peters, BA Wong, et al. 1999. Quantitative and qualitative differences in the metabolism of <sup>14</sup> C-1,3-butadiene in rats and mice: relevance to cancer susceptibility. <i>Toxicol Sci</i> 49: 186-201.
20 21 22	Sathiakumar, N, E Delzell, M Hovinga, et al. 1998. Mortality from cancer and other causes of death among synthetic rubber workers. <i>Occup Environ Med</i> 55: 230-35.
23 24 25	Sexton, KG, HE Jeffries, M Jang, et al. 2004. Photochemical products in urban mixtures enhance inflammatory responses in lung cells. <i>Inhal Toxicol</i> 16 Suppl 1: 107-14.
23 26 27 28 29	Sexton, KG, ML Doyle, HE Jeffries and S Ebersviller. 2007. Development and testing of a chemical mechanism for atmospheric photochemical transformations of 1,3-butadiene. <i>Chem Biol Interact</i> 166: 156-162.
30 31 32	Shelby, MD. 1990. Results of NTP-sponsored mouse cytogenetic studies on 1,3-butadiene, isoprene, and chloroprene. <i>Environ Health Perspect</i> 86: 71-3.
32 33 34 35	Shugaev, BB. 1969. Concentrations of hydrocarbons in tissues as a measure of toxicity. <i>Arch Environ Health</i> 18: 878-82.
36 37 38	Sielken, RL, RH Reitz, and SM Hays. 1996. Using PBPK modeling and comprehensive realism methodology for the quantitative cancer risk assessment of butadiene. Toxicology 113: 231-37.
39 40 41 42	Sills, RC, HL Hong, GA Boorman, et al. 2001. Point mutations of K-ras and H-ras genes in forestomach neoplasms from control B6C3F1 mice and following exposure to 1,3-butadiene, isoprene or chloroprene for up to 2 -years. <i>Chem Biol Interact</i> 135-136: 373-86.
43 44 45	Sorsa, M, K Peltonen, D Anderson, et al. 1996. Assessment of environmental and occupational exposures to butadiene as a model for risk estimation of petrochemical emissions. <i>Mutagenesis</i> 11: 9-17.
43 46 47	Spano, M, C Bartoleschi, E Cordelli, et al. 1996. Flow cytometric and histological assessment of 1,2:3,4- diepoxybutane toxicity on mouse spermatogenesis. <i>J Toxicol Environ Health</i> 47: 423-41.

1	
2 3 4	Spencer, F, L Chi, and M Zhu. 2001. A mechanistic assessment of 1,3-butadiene diepoxide-induced inhibition of uterine deciduoma proliferation in pseudopregnant rats. <i>Reprdo Toxicol</i> 15: 253-60.
5 6 7 8	Sprague, CL, LA Phillips, KM Young, and AA Elfarra. 2004. Species and tissue differences in the toxicity of 3-butene-1,2-diol in male Sprague-Dawley rats and B6C3F1 mice. <i>Toxicol Sci</i> 80: 3- 13.
9 10 11 12	Sprague, CL, and AA Elfarra. 2004. Mercapturic acid urinary metabolites of 3-butene-1,2-diol as <i>in vivo</i> evidence for the formation of hydroxymethylvinyl ketone in mice and rats. <i>Chem Res Toxicol</i> 17:819-26.
13 14 15	Sprague, CL, and AA Elfarra. 2005. Protection of rats against 3-butene-1,2-diol-induced hepatotoxicity and hypoglycemia by N-acetyl-l-cysteine. <i>Toxicol Appl Pharmacol</i> 207: 266-74.
16 17 18 19	Sram, RJ, P Rössner, O Beskid, et al. 2007. Chromosomal aberration frequencies determined by conventional methods: Parallel increases over time in the region of a petrochemical industry and throughout the Czech Republic. <i>Chem Biol Interact</i> 166: 239-44.
20 21 22 23	Sram, RJ, P Rossner, K Peltonen, et al. 1998. Chromosomal aberrations, sister-chromatid exchanges, cells with high frequency of SCE, micronuclei and comet assay parameters in 1,3-butadiene-exposed workers. <i>Mutat Res</i> 419: 145-54.
24 25 26	Stephanou, G, A Russo, D Vlastos, et al. 1998. Micronucleus induction in somatic cells of mice as evaluated after 1,3-butadiene inhalation. <i>Mutat Res</i> J397: 11-20.
20 27 28 29 30	Swain, CM, ED Booth, and WP Watson. 2003. Metabolic distribution of radioactivity in Sprague-Dawley rats and B6C3F1 mice exposed to 1,3-[2,3- <sup>14</sup> C]-butadiene by whole body exposure. <i>Chem Biol</i> <i>Interact</i> 145: 175-189.
31 32 33 34	Sweeney, LM, MW Himmelstein and ML Gargas. 2001. Development of a preliminary physiologically based toxicokinetic (PBTK) model for 1,3-butadiene risk assessment. <i>Chem Biol Interact</i> 135- 136: 303-22.
35 36 37 38	Sweeney, LM, MW Himmelstein, PM Schlosser, and MA Medinsky. 1996. Physiologically based pharmacokinetic modeling of blood and tissue epoxide measurements for butadiene. <i>Toxicology</i> 113:318-21.
<ul> <li>39</li> <li>40</li> <li>41</li> <li>42</li> </ul>	Sweeney, LM, PM Schlosser, MA Medinsky, and JA Bond. 1997. Physiologically based pharmacokinetic modeling of 1,3-butadiene, 1,2-epoxy-3-butene, and 1,2:3,4-diepoxybutane toxicokinetics in mice and rats. <i>Carcinogenesis</i> 18: 611-25.
43 44 45	Swenberg, JA, H Koc, PB Upton, et al. 2001. Using DNA and hemoglobin adducts to improve the risk assessment of butadiene. <i>Chem Biol Interact</i> 135-136: 387-403.
43 46 47	Swenberg, JA, N Gorgeiva, A Ham, et al. 2002. Linking pharmacokinetics and biomarker data to mechanism of action in risk assessment. <i>Human Ecol Risk Assess</i> 8: 1315-38

1	
2	Tates, AD, FJ van Dam, CM van Teylingen, et al. 1998. Comparison of induction of hprt mutations by
3	1,3-butadiene and/or its metabolites 1,2-epoxybutene and 1,2,3,4-diepoxybutane in lymphocytes
4	from spleen of adult male mice and rats in vivo. Mutat Res 397: 21-36.
5	
6	Thornton-Manning, JR, AR Dahl, WE Bechtold, et al. 1995. Disposition of butadiene monoepoxide and
7	butadiene diepoxide in various tissues of rats and mice following a low-level inhalation exposure
8	to 1,3-butadiene. Carcinogenesis 16: 1723-31.
9	
10	Thornton-Manning, JR, AR Dahl, ML Allen, et al. 1998. Disposition of butadiene epoxides in Sprague-
11	Dawley rats following exposures to 8000 ppm 1,3-butadiene: Comparisons with tissue epoxide
12	concentrations following low-level exposures. <i>Toxicol Sci</i> 41: 167-173.
12	concentrations following low-level exposures. Toxicor Ser 41. 107-175.
13	Tommasi, AM, S de Conti, MM Dobrzynska, and A Russo. 1998. Evaluation and characterization of
14	micronuclei in early spermatids of mice exposed to 1,3-butadiene. <i>Mutat Res</i> 397: 45-54.
16	incronucier in early spermatics of nice exposed to 1,5-butadiene. <i>Mutua</i> Res 597. 45-54.
	Ton T. H. Hong, T. Doverson, et al. 2007 Evolution of constinuitions in concernational concerns
17	Ton, T, H Hong, TR Devereux, et al. 2007. Evaluation of genetic alterations in cancer-related genes in
18	lung and brain tumors from B6C3F1 mice exposed to 1,3-butadiene or chloroprene. <i>Chem Biol</i>
19	Interact 166: 112-120.
20	
21	Tsai, SP, JK Wendt, and JD Ransdell. 2001. A mortality, morbidity, and hematology study of
22	petrochemical employees potentially exposed to 1,3-butadiene monomer. Chem Biol Interact
23	135-136: 555-67.
24	
25	van Sittert, NJ, HJJJ Megens, WP Watson, and PJ Boogaard. 2000. Biomarkers of exposure to 1,3-
26	butadiene as a basis for cancer risk assessment. Toxicol Sci 56: 189-202.
27	
28	Vodicka, P, R Kumar, R Stetina, et al. 2004. Markers of individual susceptibility and DNA repair rate in
29	workers exposed to xenobiotics in a tire plant. Environ Mol Mutagen 44: 283-92.
30	
31	Ward, Jr, JB, SZ Abdel-Rahman, RF Henderson, et al. 2001. Assessment of butadiene exposure in
32	synthetic rubber manufacturing workers in Texas using frequencies of hprt mutant lymphocytes
33	as a biomarker. Chem Biol Interact 135-136: 465-483.
34	
35	White, WC. 2007. Butadiene production process overview. Chem Biol Interact 166: 10-14.
36	
37	Wickliffe, JK, SM Herring, LM Hallberg, et al. 2007. Detoxification of olefinic epoxides and nucleotide
38	excision repair of epoxide-mediated DNA damage: Insights from animal models examining
39	human sensitivity to 1,3-butadiene. Chem Biol Interact 166: 226-31.
40	
41	Wickliffe, JK, MM Ammenheuser, JJ Salazar, et al. 2003. A model of sensitivity: 1,3-Butadiene increases
42	mutant frequencies and genomic damage in mice lacking a functional microsomal epoxide
43	hydrolase gene. Environ Mol Mutagen 42: 106-10.
44	
45	
46	Yadavilli, S, and PM Muganda. 2004. Diepoxybutane induces caspase and p53-mediated apoptosis in
47	human lymphoblasts. Toxicol Appl Pharmacol 195:154-65.

1	
2	Zhang, X, and AA Elfarra. 2005. Reaction of 1,2,3,4-diepoxybutane with 2'-deoxyguanosine: initial
3 4	products and their stabilities and decomposition patterns under physiological conditions. <i>Chem Res Toxicol</i> 18: 1316 -23.
5	
6	Zhang, L, RB Hayes, W Guo, et al. 2004. Lack of increased genetic damage in 1,3-butadiene-exposed
7 8	Chinese workers studied in relation to EPHX1 and GST genotypes. <i>Mut Res</i> 558: 63-74.
9	Zhang, XY, and AA Elfarra. 2003. Identification and characterization of a series of nucleoside adducts
10	formed by the reaction of 2'-deoxyguanosine and 1,2,3,4-diepoxybutane under physiological
11	conditions. Chem Res Toxicol 16: 1606-15.
12	
13	Zhao, C, P Vodicka, RJ Sram, and K Hemminki. 2001. DNA adducts of 1,3-butadiene in humans:
14	relationships to exposure, GST genotypes, single-strand breaks, and cytogenetic end points.
15	Environ Mol Mutagen 37: 226-30.
16	
17	Zhao, C, P Vodicka, RJ Sram, and K Hemminki. 2000. Human DNA adducts of 1,3-butadiene, an
18	important environmental carcinogen. Carcinogenesis 21: 107-11.
19	
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1	<b>Appendix 1. Statistical Analyses of Developmental Endpoints</b>
2	
3	Robert L. Sielken Jr., Ph.D., and Ciriaco Valdez Flores, Ph.D., P.E.
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11	<b>TCEQ Contract 582-7-81521</b>
12	
13	
14	The analyses performed by Hackett et al. (1987) did have some important statistical flaws that needed to
15	be corrected. The statistical analyses reported by Green (2003) are valid and correct the flaws of Hackett
16	et al. analyses. We have focused on the analyses of fetal body weights. The NOAEL based on the fetal
17	weights for this study is 40 ppm.
18	
19	Hackett et al. (1987) conducted analyses of variance (ANOVA) on the average pup weight followed up
20	by Student's t-tests comparing the average pup weight for different treatment groups. Their pairwise
21	comparisons using Student's t-test do not adjust significance levels that occur for the number of multiple tests. In addition, their analyses did not adjust for well-known important covariate effects such as litter
22 23	size. Hackett et al. analyses were based on dam's average pup weights instead of analyzing the individual
23 24	pup weights and treating the dam as a random effect, which would result in a more powerful statistical
25	test.
26	
27	The Green (2003) reanalysis was based on analysis of covariance (ANCOVA) on the average pup weight
28	and adjusting for covariates. In this context, Green used the Dunnett-Hsu test to compare the mean
29	weights for each of the exposed groups to the mean weight for the control group after both are adjusted
30	for the effects of the covariates. This is the specific situation for which the Dunnett-Hsu test was
31	designed. Furthermore, the Dunnett-Hsu test is the appropriate test to use here to determine a NOAEL.
32	Green considered the p-values in the Dunnett-Hsu test to draw his conclusions of significant effects.
33	Green's discussion in A. Evaluation of Earlier Methods and B. Method of Re-Analysis is appropriate.
34	Crean's analysis were based on dam's sugress num visishts instead of enclusing the individual num
35 36	Green's analyses were based on dam's average pup weights instead of analyzing the individual pup weights and treating the dam as a random effect, which would result in a more powerful statistical test.
30 37	The statistical conclusions reached by Green (2003) hold even when the more powerful statistical
38	analyses where the individual pup weights are analyzed and the dams are treated as random effects.
39	analyses where the individual pup weights are analyzed and the dams are treated as fandom effects.
40	Thus, the Green (2003) conclusions are reasonable and based on standard statistical analyses practices
41	that were overlooked by Hackett et al. (2003). The NOAEL based on the fetal weights for this study is 40
42	ppm.
43	
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1

#### Statistical Analyses Performed by Sielken & Associates

2

3 In addition to reviewing the methodology used in Hackett et al. (1987) and Green (2003), Sielken &

4 Associates re-analyzed the fetal weight data. This was to confirm the numerical results obtained by

5 Green, do a sensitivity analysis with respect to the effects of covariates, and determine the outcome of the

6 more powerful statistical analyses where the individual pup weights are analyzed and the dams are treated 7 as random effects. These analyses support the finding that the NOAEL based on the fetal weights for this

- 7 as random effects8 study is 40 ppm.
- 9

10 Table 1 contains an overview of the results in Tables 2 to 10 which contain the detailed analyses. The raw

11 data used are given in Table 11. The statistical analyses were done in SAS Ver. 9. In the overview in

- 12 Table 1, all comparisons to control were based on Dunnett-Hsu tests and were one-sided tests for a
- 13 decrease in fetal weight compared to control. The outcomes of the more powerful statistical analyses

14 where the individual pup weights are analyzed and the dams are treated as random effects were

- 15 comparable to the outcomes obtained with the Green ANCOVA model. The results for 1 Covariate (Litter
- 16 Size) are highlighted since this covariate was always statistically significant at the 5% significance level –
- 17 the 2nd Covariate (% Males in Litter) was significant for the Males Only analyses.
- 18 19

1 Table 1. Overview of Statistical Analyses of Fetal Weight Data: The results for 1 Covariate (Litter Size)

- 2 are highlighted since this covariate was always statistically significant at the 5% significance level – the 2nd Covariate (% Males in Litter) was significant for the Males Only analyses
- 3 4

Table #	Model: Mixed Model: (1) Based on Mean Data (2) Based on Individual Data and Dam as Random Effect	Sex	# of Covariates	Covariates (1) Litter Size (2) % Males in Litter	p-value in one-sided control dose=40		
2	(1)	M&F	2	(1) & (2)	0.1354	< 0.0001	< 0.0001
2	(2)	M&F	2	(1) & (2)	0.1383	< 0.0001	< 0.0001
3	(1)	M&F	1	(1)	0.1120	< 0.0001	< 0.0001
3	(2)	M&F	1	(1)	0.1184	< 0.0001	< 0.0001
4	(1)	M&F	0	None	0.0832	< 0.0001	< 0.0001
4	(2)	M&F	0	None	0.0849	< 0.0001	< 0.0001
5	(1)	F	2	(1) & (2)	0.2091	< 0.0001	< 0.0001
5	(2)	F	2	(1) & (2)	0.2373	< 0.0001	< 0.0001
6	(1)	F	1	(1)	0.1919	< 0.0001	< 0.0001
6	(2)	F	1	(1)	0.2298	< 0.0001	< 0.0001
7	(1)	F	0	None	0.1427	< 0.0001	< 0.0001
7	(2)	F	0	None	0.1854	< 0.0001	< 0.0001
8	(1)	М	2	(1) & (2)	0.0687	< 0.0001	< 0.0001
8	(2)	М	2	(1) & (2)	0.0795	< 0.0001	< 0.0001
9	(1)	М	1	(1)	0.0603	< 0.0001	< 0.0001
9	(2)	М	1	(1)	0.0695	< 0.0001	< 0.0001
10	(1)	М	0	None	0.0408	< 0.0001	< 0.0001
10	(2)	Μ	0	None	0.0479	< 0.0001	< 0.0001

#### Table 2.

1

- Males & Females Combined
- Litter Size & %Males as Covariates
- Mixed Model Based on Mean Data

	Mixed Model Based on Mean Data									
Data Set						WORK.MEANDATA				
_						ь£ г.	und FEE	-+-		
	Type 3 Tests of Fixed Effects									
					Num	Den			-	
				Effect	DF	DF	F Valu			
				dose	3	72				
				PercMales	1	72		54 0.06		
				LitterSize	e 1	72	19.3	10 <.00	01	
					Least	Squares	s Means			
						Standard				
		Effec	t d	ose Est	timate	Erro	n DF	t Valu	e Pr> t	
		dose		0 1	L.3348	0.02034	4 72	65.6	3 <.0001	
		dose		40 1	L.2898	0.01984	4 72	65.0	2 <.0001	
		dose		200 1	L.1243	0.01879	9 72	59.8	3 <.0001	
		dose	1	000 1	L.0378	0.01926	5 72	53.8	8 <.0001	
					Standard					
	Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P
	dose	40	0	-0.04497	0.02849	72	-1.58	0.1188	5	0.2701
	dose	200	0	-0.2104	0.02767	72	-7.60	<.0001	Dunnett-Hsu	<.0001
	dose	1000	0	-0.2969	0.02801	72	-10.60	<.0001	Dunnett-Hsu	<.0001
					Standard					
	Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P
	dose	0	40	0.04497	0.02849	72	1.58	0.0594	Dunnett-Hsu	0.1354
	dose	0	200	0.2104	0.02767	72	7.60	<.0001	Dunnett-Hsu	<.0001
	dose	0	1000	0.2969	0.02801	72	10.60	<.0001	Dunnett-Hsu	<.0001

Mixed Model Based on Individual Data and Dam as Random Effect

4										
5			Dat	a Set		W	IORK.ANDAT	A		
6 7				-	una 2 Taatu		ad FEEaa			
8				I	ype 3 Test: Num	Den	еа Еттест	S		
9				Effect	DF	Den	F Valu	ie Pr>	-	
0										
1				dose PercMales	3	69.3	52.7			
1						72.9	4.1			
$\frac{2}{2}$				LitterSiz	e 1	83.5	14.6	69 0.00	02	
2 3 4 5						~				
4						Squares				
				-		Standard				
6		Effect	: a		timate	Error		t Valu		
7		dose			1.3265	0.02009		66.0		
8		dose			1.2829	0.01930		66.4		
9		dose			1.1145	0.01847		60.3		
0		dose	1	000	1.0306	0.01886	68.9	54.6	4 <.0001	
1										
2 3					Standard					
3	Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P
4 5	dose	40	0	-0.04357	0.02782	69.1	-1.57	0.1218	Dunnett-Hsu	0.2759
5	dose	200	0	-0.2120	0.02714	70	-7.81	<.0001	Dunnett-Hsu	<.0001
6	dose	1000	0	-0.2959	0.02739	69.5	-10.81	<.0001	Dunnett-Hsu	<.0001
7					Standard					
8	Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P
9	dose	0	40	0.04357	0.02782	69.1	1.57	0.0609	Dunnett-Hsu	0.1383
0	dose	0	200	0.2120	0.02714	70	7.81	<.0001	Dunnett-Hsu	<.0001
1	dose	0	1000	0.2959	0.02739	69.5	10.81	<.0001	Dunnett-Hsu	<.0001

#### Table 3.

Males & Females Combined

LitterSize as Covariate (%Males not included as a covariate)

Mixed Model Based on Mean Data

		Dat	a Set				h	IORK.	MEAN	DATA					
				Тур	oe 3 Te Num		of Fi Den	xed	Effe	cts					
			Effect		DF		DEN	E	Val		Pr ≻	с			
					3		73	Г	48.		<.00				
			dose		-		-								
			LitterSi	ze	1		73		16.	36	0.00	01			
					Leas	t S	quares	Mea	ns						
							andard								
	Effect	t d	ose E	stim	nate		Error	•	DF	t	: Valu	e	Pr >  t		
	dose		0	1.3	3358	e	.02068	3	73		64.6	0	<.0001		
	dose		40	1.2	2871	e	.02013	3	73		63.9	5	<.0001		
	dose		200	1.1	L249	e	.01911		73		58.8	5	<.0001		
	dose	1	000	1.0	9388	e	.01959	)	73		53.0	3	<.0001		
				-	Standar				_	_	1.1				_
Effect	dose	dose	Estimat	-	Erro			t Va			•  t		stment	Adj	
dose	40	0	-0.0487		0.0289		73		.68		0963		ett-Hsu	0.223	
dose	200	0	-0.210	9	0.0281	5	73	-7	.49	<.	0001	Dunn	ett-Hsu	<.000	ð1
dose	1000	0	-0.297	1	0.0284	9	73	-10	.43	<.	0001	Dunn	ett-Hsu	<.000	ð1
				S	Standar	d									
Effect	dose	dose	Estimat	e	Erro	r	DF	t Va	lue	Pr	`≻t	Adju	stment	Adj	Р
dose	0	40	0.0487	0	0.0289	1	73	1	.68	0.	0482	Dunn	ett-Hsu	0.112	20
dose	0	200	0.210	9	0.0281	5	73	7	.49	<.	0001	Dunn	ett-Hsu	<.000	<b>)</b> 1
dose	0	1000	0.297	1	0.0284	9	73	10	.43	<.	0001	Dunn	ett-Hsu	<.000	<b>)</b> 1

Mixed Model Based on Individual Data and Dam as Random Effect

		Dat	a Set		h	IORK . ANDA	ТА		
			т	ype 3 Test	ts of Fi	ved Effe	rts		
				Num	Den				
			Effect	DF	DF	F Valu	ue Pr>	F	
			dose	3	71	49.			
			LitterSize		86.7	12.0			
				Loost	Causna	Maana			
					Squares Standaro				
	Effec	+ 4	lose Est	imate	Error		t Valu	e Pr> t	
	dose	L U			0.02058		64.5		
	dose				0.01974				
	dose				0.01972				
	dose	T	.000	.0318	0.01932	/0./	53.4	.0001	
				Standard					
Effec	t dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P
dose	40	0	-0.04706	0.02846	71	-1.65	0.1026	Dunnett-Hsu	0.2365
dose	200	0	-0.2112	0.02781	71.7	-7.60	<.0001	Dunnett-Hsu	<.0001
dose	1000	0	-0.2956	0.02807	71.2	-10.53	<.0001	Dunnett-Hsu	<.0001
				Standard					
Effec	t dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P
dose	0	40	0.04706	0.02846	71	1.65	0.0513	Dunnett-Hsu	0.1184
dose	0	200	0.2112	0.02781	71.7	7.60	<.0001	Dunnett-Hsu	<.0001
dose	0	1000	0.2956	0.02807	71.2	10.53	<.0001	Dunnett-Hsu	<.0001

1 2 

#### Table 4.

Males & Females Combined

No Covariates

Model Based on Mean Data

		Dat	a Set			WOF	RK.MEANI	DATA		
			-	Type 3 Tes <sup>.</sup> Num	ts of F Den		ed Effe	cts		
			Effect	DF	DF		F Val	ue Pr>	F	
			dose	3	74		40.	30 <.00	01	
				Least	Square	s I	leans			
				1	Standar	d				
	Effect	t d	ose Est	timate	Erro	r	DF	t Valu	e Pr> t	
	dose		0 3	1.3407	0.0226	9	74	59.1	0 <.0001	
	dose		40 3	1.2824	0.0220	8	74	58.0	8 <.0001	
	dose		200 2	1.1259	0.0210	0	74	53.6	0 <.0001	
	dose	1	000	1.0379	0.0215	2	74	48.2	2 <.0001	
				Standard						
Effect	dose	dose	Estimate	Error	DF	t	Value	Pr >  t	Adjustment	Adj P
dose	40	0	-0.05832	0.03166	74		-1.84	0.0695	Dunnett	0.1664
dose	200	0	-0.2148	0.03092	74		-6.95	<.0001	Dunnett	<.0001
dose	1000	0	-0.3028	0.03127	74		-9.68	<.0001	Dunnett	<.0001
				Standard						
Effect	dose	dose	Estimate	Error	DF	t	Value	Pr > t	Adjustment	Adj P
dose	0	40	0.05832	0.03166	74		1.84	0.0347	Dunnett	0.0832
dose	0	200	0.2148	0.03092	74		6.95	<.0001	Dunnett	<.0001
dose	0	1000	0.3028	0.03127	74		9.68	<.0001	Dunnett	<.0001

#### Mixed Model Based on Individual Data and Dam as Random Effect

		Dat	a Set		k	ORK.ANDA	ТА		
			1	Type 3 Test Num	ts of Fi Den	ixed Effe	cts		
			Effect	DF	DF	F Val	ue Pr>	F	
			dose	3	68.5	44.	45 <.00	01	
				Least	Squares	s Means			
					Standard	t			
	Effect	: d	ose Est	timate	Error	n DF	t Valu	e Pr> t	
	dose		0 1	L.3377	0.02163	69.1	61.8	4 <.0001	
	dose		40 1	L.2825	0.02095	5 67.9	61.2	3 <.0001	
	dose		200 1	L.1217	0.02001	L 68.7	56.0	4 <.0001	
	dose	1	.000 1	L.0377	0.02044	4 68.2	50.7	8 <.0001	
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P
dose	40	0	-0.05520	0.03011	68.5	-1.83	0.0711	Dunnett	0.1696
dose	200	0	-0.2160	0.02947	68.9	-7.33	<.0001	Dunnett	<.0001
dose	1000	0	-0.3001	0.02976	68.6	-10.08	<.0001	Dunnett	<.0001
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P
dose	0	40	0.05520	0.03011	68.5	1.83	0.0355	Dunnett	0.0849
dose	0	200	0.2160	0.02947	68.9	7.33	<.0001	Dunnett	<.0001
dose	0	1000	0.3001	0.02976	68.6	10.08	<.0001	Dunnett	<.0001

#### Table 5. 1

# Females Only LitterSize & %Males as Covariates

Mixed Model Based on Mean Data	
--------------------------------	--

		Dat	a Set		١	WORK.MEAN	DATABYSEX		
			I	Type 3 Tes		ixed Effe	cts		
				Num	Den				
			Effect	DF	DF	F Val	ue Pr>	F	
			dose	3	72	45.	71 <.00	01	
			PercMales	1	72	0.	47 0.49	36	
			LitterSize	e 1	72	13.	89 0.00	04	
				Least	Square	s Means			
					Standar				
	Effect	t d	ose Est	imate	Erro	r DF	t Valu	e Pr> t	
	dose		0 1	L.2949	0.0202	9 72	64.0		
	dose		40 1	L.2579	0.0197	1 72	63.8	3 <.0001	
	dose		200 1	L.0991	0.0186	7 72	58.8	7 <.0001	
	dose	1	.000 1	L.0155	0.0191	3 72	53.0	7 <.0001	
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P
dose	40	0	-0.03706	0.02830	72	-1.31	0.1945	Dunnett-Hsu	0.4148
dose	200	0	-0.1958	0.02749	72	-7.12	<.0001	Dunnett-Hsu	<.0001
dose	1000	0	-0.2794	0.02783	72	-10.04	<.0001	Dunnett-Hsu	<.0001
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P
dose	0	40	0.03706	0.02830	72	1.31	0.0973	Dunnett-Hsu	0.2091
dose	0	200	0.1958	0.02749	72	7.12	<.0001	Dunnett-Hsu	<.0001
dose	0	1000	0.2794	0.02783	72	10.04	<.0001	Dunnett-Hsu	<.0001

Mixed Model Based on Individual Data and Dam as Random Effect

		Dat	a Set		L	WORK.ANDA	ТА		
			Т	ype 3 Tes	ts of Fi	ixed Effe	cts		
				Num	Den				
			Effect	DF	DF	F Val	ue Pr>	• F	
			dose	3	67.9	48.	10 <.00	001	
			PercMales	1	77.9	0.	65 0.42	28	
			LitterSize	e 1	85.2	10.	64 0.00	916	
				Least	Squares	s Means			
					Standard	t			
	Effec	t d	lose Est	imate	Error	n DF	t Valu	ıe Pr> t	
	dose		0 1	.2850	0.02019	9 67.3	63.6	6 <.0001	
	dose		40 1	.2514	0.01897	7 65.2	65.9	97 <.0001	
	dose		200 1	.0881	0.01853	67.3	58.7	′2 <.0001	
	dose	1	.000 1	.0063	0.01889	66.8	53.2	<.0001	
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Ad
dose	40	0	-0.03367	0.02761	67.1	-1.22	0.2269	Dunnett-Hsu	0.4
dose	200	0	-0.1969	0.02714	69.4	-7.26	<.0001	Dunnett-Hsu	<.0
dose	1000	0	-0.2788	0.02738	68.9	-10.18	<.0001	Dunnett-Hsu	<.0
				Standard					
Effect	dose	dose	Estimate	Error		t Value	Pr > t	5	Ad
dose	0	40	0.03367	0.02761		1.22	0.1134		0.2
dose	0	200	0.1969	0.02714		7.26	<.0001		<.00
dose	0	1000	0.2788	0.02738	68.9	10.18	<.0001	Dunnett-Hsu	<.00

#### 1 Table 6.

- Females Only
- LitterSize as Covariate (%Males not included as a covariate) Mixed Model Based on Mean Data

Mixed Model Based on Mean Data													
		Dat	a Set		WORK.MEANDATABYSEX								
			Т	ype 3 Test	ts of F	ixed Effe	cts						
				Num	Den								
			Effect	DF	DF	F Val	ue Pr>	F					
			dose	3	73	45.	91 <.00	001					
			LitterSize	e 1	73	13.	51 0.00	004					
Least Squares Means													
Standard													
	Effec	t d	ose Est	imate	Erro		t Valu	e Pr> t					
	dose			.2953	0.0201								
	dose			.2569	0.0195								
	dose			.0993	0.0186								
	dose			.0159	0.0190								
				Standard									
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P				
dose	40	0	-0.03841	0.02813	73	-1.37	0.1763	Dunnett-Hsu	0.3813				
dose	200	0	-0.1960	0.02739		-7.15	<.0001	Dunnett-Hsu	<.0001				
dose	1000	0	-0.1900	0.02733		-10.08	<.0001	Dunnett-Hsu	<.0001				
uose	1000	0	-0.2795	Standard	/ 5	-10.08	1.0001	Duffiele	1.0001				
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P				
dose	uose 0	40 40	0.03841	0.02813		1.37	0.0881	Dunnett-Hsu	0.1919				
dose	0	200	0.1960	0.02739		7.15	<.0001	Dunnett-Hsu	<.0001				
dose	0	1000	0.1900	0.02733	73	10.08	<.0001	Dunnett-Hsu	<.0001				
uuse	0	1000	0.2795	0.02//5	, ,	10.00	1.0001	Dunnett-nsu					

Mixed Model Based on Individual Data and Dam as Random Effect

		Dat	a Set		h	ORK.ANDA	ГА		
			1	Type 3 Test	ts of Fi	ixed Effe	ts		
				Num	Den				
			Effect	DF	DF	F Valu	ue Pr>	F	
			dose	3	69.2	47.9	97 <.00	01	
			LitterSize	e 1	87.8	10.1	L1 0.00	20	
				Least	Squares	Means			
					Standard				
	Effect	: d	ose Est	timate	Error	DF	t Valu	ie Pr> t	
	dose		0 1	L.2864	0.02010	9 70	63.9	9 <.0001	
	dose		40 1	L.2522	0.01893	66.9	66.1	.5 <.0001	
	dose		200 1	L.0904	0.01830	69.3	59.5	8 <.0001	
	dose	1	.000 1	L.0084	0.01869	69.2	53.9	<.0001	
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P
dose	40	0	-0.03424	0.02758	68.7	-1.24	0.2186	Dunnett-Hsu	0.4550
dose	200	0	-0.1960	0.02709	70.5	-7.23	<.0001	Dunnett-Hsu	<.0001
dose	1000	0	-0.2780	0.02734	70.1	-10.17	<.0001	Dunnett-Hsu	<.0001
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P
dose	0	40	0.03424	0.02758	68.7	1.24	0.1093	Dunnett-Hsu	0.2298
dose	0	200	0.1960	0.02709	70.5	7.23	<.0001	Dunnett-Hsu	<.0001
dose	0	1000	0.2780	0.02734	70.1	10.17	<.0001	Dunnett-Hsu	<.0001

#### Table 7.

- 2
  - Females Only
- No Covariates
- Model Based on Mean Data

		Dat	a Set		WORK.MEANDATABYSEX					
			Т	Type 3 Test Num	e 3 Tests of Fixed Effects Num Den					
			Effect	DF	DF		F Valu	ue Pr>	F	
			dose	3	74		39.0	52 <.00	01	
				Least	Square	s١	leans			
					Standar					
	Effect	: d	ose Est	imate	Erro	r	DF	t Valu	e Pr> t	
	dose		0 1	L.2996	0.0217	2	74	59.8	3 <.0001	
	dose		40 1	L.2527	0.0211	4	74	59.2	5 <.0001	
	dose		200 1	1.1001	0.0201	1	74	54.7	1 <.0001	
	dose	1	.000 1	1.0151	0.0206	1	74	49.2	6 <.0001	
				Standard						
Effect	dose	dose	Estimate	Error	DF	t	Value	Pr >  t	Adjustment	Adj P
dose	40	0	-0.04692	0.03031	74		-1.55	0.1259	Dunnett	0.2846
dose	200	0	-0.1995	0.02960	74		-6.74	<.0001	Dunnett	<.0001
dose	1000	0	-0.2846	0.02994	74		-9.50	<.0001	Dunnett	<.0001
				Standard						
Effect	dose	dose	Estimate	Error	DF	t	Value	Pr > t	Adjustment	Adj P
dose	0	40	0.04692	0.03031	74		1.55	0.0630	Dunnett	0.1427
dose	0	200	0.1995	0.02960	74		6.74	<.0001	Dunnett	<.0001
dose	0	1000	0.2846	0.02994	74		9.50	<.0001	Dunnett	<.0001

Mixed Model Based on Individual Data and Dam as Random Effect

4												
4 5 6 7			Dat	a Set				WOI	RK.ANDA	ТА		
0					<b>.</b>					- 4 -		
8					Тy	pe 3 Test			ed Ette	cτs		
9				F C C +		Num	Den		F 1/- 1		-	
9				Effect		DF	DF		F Val			
1				dose		3	67.1		43.	81 <.00	1001	
1 2 3 4 5 6 7						1	C					
2							Square		rieans			
5 1							Standar		DF	+ \/-1.	الحل م	
+		Effect	τ α			mate	Erro		DF	t Valu		
5		dose		0		2935	0.0209		68.7			
2		dose		40		2536	0.0198		64.9			
8		dose		200		0947	0.0191		67.7			
9		dose	1	.000	1.	0130	0.0195	1	67.2	51.9	·2 <.0001	
0						ام م م م م						
		4	4	E - to day and		Standard			N - 1		<b>A d d d d d d d d d d</b>	
1 7	Effect	dose	dose	Estimat		Error		τ	Value	Pr >  t	5	Adj P
2	dose	40	0	-0.0399		0.02882	66.9		-1.39	0.1706	Dunnett	0.3688
5 4	dose	200	0	-0.198		0.02833	68.3		-7.02	<.0001	Dunnett	<.0001
1 2 3 4 5 6 7	dose	1000	0	-0.286		0.02860	68		-9.81	<.0001	Dunnett	<.0001
5		4	4	E - to day and		Standard			N - 1	Du i t	<b>A d d d d d d d d d d</b>	
2	Effect	dose	dose	Estimat		Error	DF	τ	Value	Pr > t	Adjustment	Adj P
/	dose	0	40	0.0399		0.02882	66.9		1.39	0.0853	Dunnett	0.1854
8 9	dose	0	200	0.198		0.02833	68.3		7.02	<.0001	Dunnett	<.0001
-	dose	0	1000	0.280	15	0.02860	68		9.81	<.0001	Dunnett	<.0001
0												

1 2 

#### Table 8.

1

Males Only

#### LitterSize & %Males as Covariates

Mixed Model Based on Mean Data
--------------------------------

		Dat	a Set		WORK.MEANDATABYSEX						
			1	Type 3 Tes	ts of Fi	ixed Effe	cts				
				Num	Den						
			Effect	DF	DF	F Val	ue Pr>	F			
			dose	3	71	51.	81 <.00	01			
	PercMales				71	6.	19 0.01	52			
	LitterSize				71	5.	31 0.02	41			
				Least	Square	s Means					
				:	Standar	d					
	Effec	t d	lose Est	imate	Erro	r DF	t Valu	e Pr> t			
	dose		0 1	L.3704	0.0211	3 71	64.8	6 <.0001			
	dose		40 1	L.3131	0.0205	3 71	63.9	5 <.0001			
	dose		200 1	L.1321	0.0201	1 71	56.3	0 <.0001			
	dose	1	.000 1	L.0582	0.0199	3 71	53.0	9 <.0001			
				Standard							
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P		
dose	40	0	-0.05724	0.02950	71	-1.94	0.0563	Dunnett-Hsu	0.1372		
dose	200	0	-0.2382	0.02934	71	-8.12	<.0001	Dunnett-Hsu	<.0001		
dose	1000	0	-0.3122	0.02901	71	-10.76	<.0001	Dunnett-Hsu	<.0001		
				Standard							
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P		
dose	0	40	0.05724	0.02950	71	1.94	0.0282	Dunnett-Hsu	0.0687		
dose	0	200	0.2382	0.02934	71	8.12	<.0001	Dunnett-Hsu	<.0001		
dose	0	1000	0.3122	0.02901	71	10.76	<.0001	Dunnett-Hsu	<.0001		

Mixed Model Based on Individual Data and Dam as Random Effect

		Dat	a Set		h				
			Т	ype 3 Tes	ts of Fi	ixed Effe	cts		
				Num	Den				
			Effect	DF	DF	F Val	ue Pr>	F	
			dose	3	69.5	52.	24 <.00	001	
			PercMales	1	73	5.	56 0.02	10	
			LitterSize	e 1	74.4	4.	65 0.03	43	
				Least	Squares	5 Means			
					Standard	t			
	Effect	: d	ose Est	imate	Error	ר DF	t Valu	ie Pr> t	
	dose		0 1	.3704	0.02132	2 69.9	64.2	.0001	
	dose		40 1	.3158	0.02065	69.2	63.7	'3 <.0001	
	dose		200 1	.1346	0.01953	67.4	58.0	9 <.0001	
	dose	1	000 1	.0607	0.01992	69.1	53.2	<.0001	
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Ac
dose	40	0	-0.05466	0.02928	70.7	-1.87	0.0661	Dunnett-Hsu	0.1
dose	200	0	-0.2359	0.02892	68.9	-8.16	<.0001	Dunnett-Hsu	<.(
dose	1000	0	-0.3098	0.02879	70.6	-10.76	<.0001	Dunnett-Hsu	<.6
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	5	Ac
dose	0	40	0.05466	0.02928		1.87			0.0
dose	0	200	0.2359	0.02892	68.9	8.16	<.0001	Dunnett-Hsu	<.6
dose	0	1000	0.3098	0.02879	70.6	10.76	<.0001	Dunnett-Hsu	<.6

#### Table 9.

1

- Males Only
- LitterSize as Covariate (%Males not included as a covariate)
- Mixed Model Based on Mean Data

1000

0

viixed iv		based	on Mean	Data								
		Dat	a Set		WORK.MEANDATABYSEX							
			-									
			I	ype 3 Tes			ects					
				Num	Den							
			Effect	DF	DF	F Val	.ue Pr>	F				
			dose	3	72	47.	18 <.00	01				
			LitterSize	e 1	72	5.	88 0.01	.78				
				least	Square	s Means						
					Standar							
	Effec	ь ,	aca Ect				+ 1/21.					
		L U		imate	Erro							
	dose			.3697	0.0218							
	dose			.3086	0.0211		61.7					
	dose			.1368	0.0207		54.8					
	dose	1	.000 1	.0583	0.0206	4 72	51.2	8 <.0001				
			Diff	erences o	f Least	Squares	Means					
				Standard		·						
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P			
dose	40	0	-0.06107	0.03050	72	-2.00	0.0490	Dunnett-Hsu	0.1206			
dose	200	0	-0.2329	0.03029	72	-7.69	<.0001	Dunnett-Hsu	<.0001			
dose	1000	0	-0.3114	0.03003	72	-10.37	<.0001	Dunnett-Hsu	<.0001			
0.000	2000	Ū	010111	Standard		20107						
Effect	dose	dose	Estimate	Error		t Value	Pr \ t	Adjustment	Adj P			
dose	0	40	0.06107	0.03050		2.00		-	0.0603			
dose	0	200	0.2329	0.03029	72	7.69	<.0001	Dunnett-Hsu	<.0001			

72

10.37

<.0001 Dunnett-Hsu

<.0001

Mixed Model Based on Individual Data and Dam as Random Effect

0.3114

0.03003

		Dat	a Set		W	IORK.ANDA			
			T	Type 3 Test Num	ts of Fi Den	xed Effe	cts		
			Effect	DF	DF	F Valu	ue Pr>	F	
			dose	3	69.6	48.	38 <.00	01	
			LitterSize	e 1	74.2	5.2	26 0.02	46	
				Least	Squares	Means			
					Standard				
	Effect	: d	ose Est	timate	Error	DF	t Valu	e Pr >  t	
	dose		0 1	L.3647	0.02176	70.3	62.7	1 <.0001	
	dose		40 1	L.3066	0.02084	70.2	62.6	9 <.0001	
	dose		200 1	L.1334	0.02007	67.5	56.4	7 <.0001	
	dose	1	000 1	L.0560	0.02037	69.6	51.8	4 <.0001	
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr >  t	Adjustment	Adj P
dose	40	0	-0.05810	0.03005	70.6	-1.93	0.0572	Dunnett-Hsu	0.1389
dose	200	0	-0.2312	0.02965	69	-7.80	<.0001	Dunnett-Hsu	<.0001
dose	1000	0	-0.3087	0.02958	70.5	-10.44	<.0001	Dunnett-Hsu	<.0001
				Standard					
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P
dose	0	40	0.05810	0.03005	70.6	1.93	0.0286	Dunnett-Hsu	0.0695
dose	0	200	0.2312	0.02965	69	7.80	<.0001	Dunnett-Hsu	<.0001
dose	0	1000	0.3087	0.02958	70.5	10.44	<.0001	Dunnett-Hsu	<.0001

dose

#### Table 10.

- Males Only
- No Covariates
- Model Based on Mean Data

		Dat	a Set		1					
				Type 3 Tes <sup>.</sup> Num	Tests of Fixed Effects Num Den					
			Effect	DF	Den			-		
			dose	3	73	45.6	58 <.000	1		
				Least	Square	s Means				
					Standar					
	Effect	t d	ose Es	timate	Erro	r DF	t Valu	e Pr> t		
	dose		0	1.3754	0.0224	6 73	61.2	3 <.0001		
	dose		40	1.3070	0.0218	6 73	59.7	8 <.0001		
	dose		200	1.1319	0.0213	1 73	53.1	2 <.0001		
	dose	1	.000	1.0596	0.0213	1 73	49.7	2 <.0001		
				Standard			- 1.1			
Effect	dose	dose	Estimate			t Value	Pr >  t	Adjustment	Adj P	
dose	40	0	-0.06845	0.03135	73	-2.18	0.0322	Dunnett	0.0815	
dose	200	0	-0.2435	0.03096	73	-7.86	<.0001	Dunnett	<.0001	
dose	1000	0	-0.3158	0.03096	73	-10.20	<.0001	Dunnett	<.0001	
				Standard						
Effect	dose	dose	Estimate	Error	DF	t Value	Pr > t	Adjustment	Adj P	
dose	0	40	0.06845	0.03135	73	2.18	0.0161	Dunnett	0.0408	
dose	0	200	0.2435	0.03096	73	7.86	<.0001	Dunnett	<.0001	
dose	0	1000	0.3158	0.03096	73	10.20	<.0001	Dunnett	<.0001	

Mixed Model Based on Individual Data and Dam as Random Effect

t t										
5			Dat	a Set			WORK.ANDA	TA		
2						_				
(					Туре 3 Т	ests of F	ixed Effe	ects		
3					Nu	m Der	1 IIII			
)				Effect	D	F DF	F Val	ue Pr >	> F	
)				dose		3 69.9	47.	22 <.00	001	
2					Lea	st Square	es Means			
3						Standar	'nd			
1		Effect	t d	lose E	stimate	Erro	or DF	t Valu	ue Pr> t	
5		dose		0	1.3729	0.0220	2 71.3	62.3		
5		dose		40	1.3081	0.021				
7		dose		200	1.1326	0.0205				
Ŝ		dose		.000	1.0604	0.0208				
)			-		210001	010200				
)					Standa	rd				
1	Effect	dose	dose	Estimat	e Err	or DF	t Value	Pr >  t	Adjustment	Adj P
2	dose	40	0	-0.0647	8 0.030	68 70.9	-2.11	0.0382	Dunnett	0.0957
3	dose	200	0	-0.240	4 0.030	15 69.6	-7.97	<.0001	Dunnett	<.0001
1	dose	1000	0	-0.312			-10.32	<.0001	Dunnett	<.0001
5					Standa					
5	Effect	dose	dose	Estimat	e Err	or DF	t Value	Pr > t	Adjustment	Adj P
7	dose	0	40	0.0647	8 0.030	68 70.9	2.11	0.0191	Dunnett	0.0479
3	dose	0	200	0.240	4 0.030	15 69.6	7.97	<.0001	Dunnett	<.0001
)	dose	0	1000	0.312		29 70.8	10.32	<.0001	Dunnett	<.0001
)										

#### 1 Table 11. Fetal Weight Data

2

(These data are the same as provided by TCEQ except that a few errors in going from the Hackett et al. (1987) data sheets to the electronic copy have been corrected.)

- 3 4 5

Index

2	Dam	SITE	Status	FetalSex	dose	feta	lwt
	1	228	1	1	1	0	1.611
	2	228	2	1	2	0	1.393
	3	228	3	1	1	0	1.524
	4	228	4	1	1	0	1.512
	5	228	5	1	2	0	1.573
	6	228	6	1	1	0	1.526
	7	228	7	1	1	0	1.563
	8	228	8	2		0	
	9	228	9	1	2	0	1.311
	10	228	10	1	1	0	1.55
	11	256	1	1	1	0	1.406
	12	256	2	1	2	0	1.277
	13	256	3	1	2	0	1.272
	14	256	4	1	1	0	1.22
	15	256	5	2		0	
	16	256	6	1	1	0	1.362
	17	256	7	1	2	0	1.273
	18	256	8	1	2	0	1.293
	19	256	9	2		0	
	20	256	10	1	1	0	1.336
	21	256	11	1	1	0	1.312
	22	256	12	1	1	0	1.316
	23	270	1	1	2	0	1.433
	24	270	2	1	1	0	1.763
	25	270	3	2		0	
	26	270	4	2		0	
	27	270	5	1	1	0	1.613
	28	270	6	2		0	
	29	273	1	1	2	0	1.352
	30	273	2	2		0	
	31	273	3	1	2	0	1.215
	32	273	4	1	2	0	1.181
	33	273	5	1	1	0	1.425
	34	273	6	1	2	0	1.204
	35	273	7	1	2	0	1.183
	36	273	8	1	1	0	1.106
	37	273	9	1	2	0	1.372
	38	273	10	1	1	0	1.37
	39	273	11	1	1	0	1.379
	40	273	12	1	2	0	1.355

41	273	13	1	2	0	0.664
42	273	14	1	1	0	1.436
43	304	1	1	2	0	1.189
44	304	2	1	1	0	1.165
45	304	3	1	2	0	1.14
46	304	4	1	1	0	1.172
47	304	5	1	1	0	1.289
48	304	6	1	2	0	1.179
49	304	7	1	1	0	1.098
50	304	8	1	1	0	1.105
51	304	9	1	2	0	1.231
52	304	10	1	2	0	1.183
53	304	11	1	1	0	1.349
54	304	12	1	1	0	1.118
55	320	1	1	1	0	1.322
56	320	2	1	2	0	1.132
57	320	3	1	2	0	1.281
58	320	4	1	1	0	1.354
59	320	5	1	1	0	1.383
60	320	6	1	1	0	1.338
61	320	7	1	2	0	1.016
62	320	8	1	2	0	1.273
63	320	9	1	2	0	1.39
64	320	10	1	2	0	1.249
65	320	11	1	1	0	1.444
66	320	12	1	2	0	1.31
67	320	13	1	2	0	1.381
68	321	1	1	2	0	1.294
69	321	2	1	1	0	1.299
70	321	3	1	2	0	1.342
71	321	4	1	2	0	1.294
72	321	5	1	1	0	1.308
73	321	6	1	1	0	1.336
74	321	7	1	1	0	1.285
75	321	8	1	2	0	1.153
76	321	9	1	2	0	1.151
77	321	10	1	1	0	1.3
78	321	11	1	1	0	1.459
79	321	12	1	1	0	1.477
80	321	13	1	2	0	1.259
81	321	14	1	1	0	1.276
82	321	15	1	2	0	1.184
83	341	1	1	1	0	1.493
84	341	2	1	1	0	1.492
85	341	3	1	1	0	1.469
86	341	4	1	2	0	1.379

87	341	5	1	1	0	1.429
88	341	6	1	2	0	1.361
89	341	7	1	2	0	1.269
90	341	8	1	1	0	1.429
91	341	9	1	2	0	1.381
92	341	10	1	1	0	1.404
93	341	11	1	2	0	1.311
94	341	12	1	2	0	1.403
95	341	13	1	1	0	1.426
96	351	1	1	2	0	1.285
97	351	2	1	2	0	1.18
98	351	3	1	1	0	1.18
99	351	4	1	1	0	1.148
100	351	5	1	2	0	1.117
101	351	6	1	1	0	1.234
102	351	7	1	2	0	1.128
103	351	8	1	2	0	1.218
104	351	9	1	2	0	1.169
105	351	10	1	1	0	0.932
106	351	11	1	2	0	1.214
107	351	12	2	-	0	
108	351	13	1	2	0	1.158
109	351	13	1	2	0	1.214
110	372	1	1	2	0	1.43
111	372	2	1	2	0	1.45
112	372	3	1	1	0	1.252
112	372	4	1	1	0	1.354
114	372	5	1	2	0	1.322
115	372	6	1	2	0	1.322
115	372	0 7	1	1	0	1.451
117	372	8	1	1	0	1.316
118	372	9	1	2	0	1.262
119	372	10	1	1	0	1.353
120	372	10	1	2	0	1.333
120	372	12	4	2	0	1.27
121	372	12	1	2	0	1.305
122	372	13	1	1	0	1.41
123	378	14	1	1	0	1.338
124	378	2	1	1	0	1.402
125	378	3	1	1	0	1.462
120	378	4	1	1	0	1.46
127	378	5	1	2	0	1.348
128	378	6	1 2	2	0	1.340
129		0 7	2 1	1	0	1.35
130	378 378	8	1 2	1	0	1.33
131	378 378	8 9	2 1	1		1 216
132	378	9	1	1	0	1.346

133	378	10	1	1	0	1.398
134	378	11	1	1	0	1.4
135	378	12	1	1	0	1.347
136	378	13	1	1	0	1.332
137	378	14	1	1	0	1.245
138	380	1	1	2	0	1.337
139	380	2	1	1	0	1.36
140	380	3	2		0	
141	380	4	1	2	0	1.276
142	380	5	1	1	0	1.429
143	380	6	1	2	0	1.295
144	380	7	1	2	0	1.284
145	380	8	1	1	0	1.482
146	380	9	1	2	0	1.334
147	380	10	1	2	0	1.236
148	380	11	1	1	0	1.365
149	380	12	1	1	0	1.357
150	380	13	1	2	0	1.36
151	380	14	1	1	0	1.275
152	388	1	2	1	0	1.270
152	388	2	1	1	0	1.511
155	388	3	1	2	0	1.37
155	388	4	1	1	0	1.459
155	388	5	1	2	0	1.428
150	388	6	1	2	0	1.345
158	388	7	1	2	0	1.441
158	388	8	1	2	0	1.376
160	388	9	1	1	0	1.279
161	388	10	1	2	0	1.419
162	391	1	1	2	0	1.419
162	391	2	1	1	0	1.341
164	391	3	1	2	0	1.341
165	391	4	1	1	0	1.302
165	391	4 5	1	2	0	1.482
167	391	6				
			1	1	0	1.281
168	391 201	7	1	1	0	1.179
169	391 201	8	2	2	0	1.07
170	391	9	1	2	0	1.07
171	391	10	1	2	0	1.261
172	391	11	1	1	0	1.269
173	391	12	1	1	0	1.344
174	391	13	1	1	0	1.489
175	391	14	1	1	0	1.502
176	415	1	1	1	0	1.459
177	415	2	2		0	
178	415	3	2		0	

179	415	4	1	1	0	1.364
180	415	5	2		0	
181	415	6	1	2	0	1.288
182	415	7	1	1	0	1.226
183	415	8	1	2	0	1.332
184	415	9	1	2	0	1.137
185	415	10	1	1	0	1.333
186	415	11	1	1	0	1.217
187	415	12	1	1	0	1.456
188	418	1	1	2	0	1.154
189	418	2	1	2	0	1.281
190	418	3	1	2	0	1.383
191	418	4	1	1	0	1.354
192	418	5	1	2	0	1.318
193	418	6	1	2	0	0.957
194	418	7	1	2	0	1.311
195	418	8	1	2	0	1.3
196	418	9	1	1	0	1.37
197	418	10	1	1	0	1.296
198	418	11	1	2	0	1.218
199	418	12	2	_	0	
200	418	13	1	2	0	1.22
201	418	14	1	2	0	1.328
202	422	1	1	2	0	1.475
203	422	2	1	2	0	1.511
204	422	3	1	1	0	1.49
205	422	4	1	2	0	1.405
206	422	5	1	1	0	1.5
207	422	6	1	2	0	1.413
208	422	7	1	1	0	1.518
209	422	8	1	1	0	1.524
210	422	9	1	1	0	1.498
211	422	10	1	2	0	1.368
212	422	11	1	2	0	1.36
212	422	12	1	2	0	1.351
213	422	13	1	1	0	1.478
215	422	13	2	1	0	1.170
215	422	15	1	1	0	1.497
210	444	1	1	2	0	1.343
217	444	2	1	2	0	1.345
210	444	3	1	1	0	1.372
219	444	4	1	2	0	1.311
220	444	4 5	1	1	0	1.311
221	444	6	1	2	0	1.259
222	444	0 7	1	1	0	1.259
223 224	444	8	1	2	0	1.33
<i>∠∠</i> 4	+++	0	1	2	U	1.2/3

225	444	9	1	1	0	1.31
226	444	10	1	1	0	1.138
227	444	11	1	2	0	1.278
228	444	12	1	1	0	1.444
229	444	13	1	2	0	1.304
230	444	14	1	2	0	1.332
231	242	1	1	1	40	1.47
232	242	2	2		40	
233	242	3	1	1	40	1.377
234	242	4	1	1	40	1.429
235	242	5	1	2	40	1.363
236	242	6	1	2	40	1.325
237	242	7	1	1	40	1.269
238	242	8	1	1	40	1.319
230	242	9	1	2	40	1.33
240	242	10	1	1	40	1.381
240 241	242	10	1	2	40	1.214
241	242	11	1	2	40 40	1.302
242	242	12	2	2	40 40	1.302
243 244	242	13	1	2	40 40	1.422
244 245	240 246	2	1	2	40 40	1.394
243 246	240 246	2 3		2	40 40	1.394
240 247		3 4	1 1	2 1		
	246				40	1.329
248	246	5	1	2	40	1.372
249	246	6	1	2	40	0.94
250	246	7	1	2	40	1.287
251	246	8	1	1	40	1.356
252	246	9	1	2	40	1.29
253	246	10	1	2	40	1.304
254	246	11	1	1	40	1.168
255	263	1	1	2	40	1.308
256	263	2	1	1	40	1.313
257	263	3	1	2	40	1.373
258	263	4	1	2	40	1.275
259	263	5	1	2	40	1.378
260	263	6	1	2	40	1.295
261	263	7	1	2	40	1.301
262	263	8	1	1	40	1.267
263	263	9	2		40	
264	263	10	1	2	40	1.326
265	263	11	1	1	40	1.363
266	263	12	1	2	40	1.312
267	263	13	1	2	40	1.321
268	263	14	1	2	40	1.048
269	286	1	1	1	40	1.429
270	286	2	1	2	40	1.233

271	286	3	1	1	40	1.32
272	286	4	1	1	40	1.326
273	286	5	1	1	40	1.359
274	286	6	1	2	40	1.334
275	286	7	1	2	40	1.321
276	286	8	1	1	40	1.426
277	286	9	1	1	40	1.407
278	286	10	1	2	40	1.283
279	286	11	1	2	40	1.356
280	286	12	1	2	40	1.142
281	286	13	1	1	40	1.409
282	286	14	1	1	40	1.313
283	295	1	1	1	40	1.426
284	295	2	1	1	40	1.292
285	295	3	1	1	40	1.25
286	295	4	1	1	40	1.443
287	295	5	1	2	40	1.241
288	295	6	1	2	40	1.23
289	295	7	1	1	40	1.289
290	295	8	4	1	40	1.209
291	295	9	1	1	40	1.376
292	295	10	1	1	40	1.287
293	295	11	1	2	40	1.157
294	295	12	1	2	40	1.291
295	295	13	1	1	40	1.349
296	295	14	1	2	40	1.264
297	302	1	1	1	40	1.133
298	302	2	1	1	40	1.14
299	302	3	1	2	40	1.065
300	302	4	1	1	40	1.193
301	302	5	1	2	40	1.079
302	302	6	1	1	40	1.108
303	302	° 7	4	1	40	1.100
304	302	8	1	1	40	1.183
305	302	9	1	2	40	1.191
306	302	10	1	1	40	1.172
307	302	11	1	2	40	1.172
308	302	12	1	1	40	1.038
309	302	12	1	1	40	1.13
310	302	13	1	1	40	1.13
311	302	15	1	1	40	1.167
312	302	16	1	1	40	1.173
312	307	1	1	2	40	1.343
313	307	1 2	1	2	40 40	1.343
314	307	2 3	1	2 1	40 40	1.227
315	307	4	1	1	40 40	1.330
310	307	4	1	1	40	1.423

317	307	5	1	1	40	1.351
318	307	6	1	2	40	1.179
319	307	7	1	2	40	1.364
320	307	8	1	1	40	1.397
321	307	9	1	1	40	1.362
322	307	10	1	1	40	1.384
323	307	11	1	2	40	1.252
324	307	12	1	2	40	1.265
325	307	13	1	2	40	1.35
326	311	1	1	1	40	1.378
327	311	2	1	1	40	1.337
328	311	3	1	1	40	1.4
329	311	4	1	2	40	1.315
330	311	5	1	2	40	1.297
331	311	6	1	1	40	1.43
332	311	7	1	1	40	1.38
333	311	8	1	2	40	1.294
334	311	8 9	1	2	40	1.294
335	311	10	1	2	40 40	1.290
336	311		1	2 1	40 40	1.31
330 337		11 12		1 2	40 40	
	311		1			1.063
338	312	1	1	1	40	1.344
339	312	2	1	1	40	1.239
340	312	3	1	2	40	1.273
341	312	4	1	2	40	1.249
342	312	5	1	2	40	1.259
343	312	6	1	2	40	1.149
344	312	7	1	1	40	1.312
345	312	8	1	2	40	1.217
346	312	9	1	2	40	1.386
347	312	10	1	1	40	1.235
348	312	11	1	2	40	1.151
349	312	12	1	1	40	1.215
350	312	13	1	1	40	1.291
351	312	14	1	1	40	1.146
352	312	15	1	2	40	1.199
353	312	16	4		40	
354	312	17	1	1	40	1.305
355	314	1	1	1	40	1.405
356	314	2	1	2	40	1.184
357	314	3	1	1	40	1.184
358	314	4	1	1	40	1.424
359	314	5	1	2	40	1.3
360	314	6	1	1	40	1.313
361	314	7	1	1	40	1.416
362	314	8	1	1	40	1.437

363	314	9	1	2	40	1.288
364	314	10	2		40	
365	314	11	1	2	40	1.287
366	314	12	1	2	40	1.321
367	318	1	1	1	40	1.482
368	318	2	1	2	40	1.289
369	318	3	1	1	40	1.245
370	318	4	1	1	40	1.379
371	318	5	1	2	40	1.256
372	318	6	1	1	40	1.217
373	318	7	1	2	40	1.339
374	318	8	1	2	40	1.308
375	318	9	4		40	
376	318	10	1	2	40	1.205
377	318	11	1	1	40	1.49
378	318	12	1	2	40	1.284
379	318	13	1	1	40	1.321
380	346	1	1	2	40	1.092
381	346	2	2	-	40	1.07
382	346	3	2		40	
383	346	4	2		40	
384	346	5	1	1	40	1.31
385	346	6	1	1	40	1.322
386	346	7	1	1	40	1.048
387	346	8	1	2	40	1.238
388	346	9	1	2	40	1.167
389	349	1	1	2	40	1.015
390	349	2	1	2	40	1.227
391	349	3	1	1	40	1.249
392	349	4	1	2	40	1.394
393	349	5	1	2	40	1.334
394	349	6	1	1	40	1.404
395	349	7	1	1	40	1.344
396	349	8	1	1	40	1.395
397	349	9	1	1	40	1.391
398	349	10	1	1	40	1.246
399	349	11	1	1	40	1.411
400	349	12	1	2	40	1.349
401	349	13	1	2	40	1.354
402	368	1	1	2	40	1.283
403	368	2	1	2	40	1.396
404	368	$\frac{2}{3}$	1	1	40	1.421
405	368	4	1	2	40	1.253
406	368	5	1	1	40	1.355
400	368	6	1	1	40	1.391
408	368	0 7	1	1	40	1.379
100	200	'	1	1	10	1.217

409	368	8	1	1	40	1.48
410	368	9	1	2	40	1.365
411	368	10	1	1	40	1.235
412	368	11	1	1	40	1.369
413	369	1	1	2	40	1.286
414	369	2	1	2	40	1.237
415	369	3	1	1	40	1.292
416	369	4	1	2	40	1.216
417	369	5	1	1	40	1.23
418	369	6	2	-	40	
419	369	7	1	1	40	1.276
420	369	8	1	1	40	1.127
421	369	9	1	2	40	1.345
422	369	10	2	-	40	1.5 10
423	369	11	1	2	40	1.251
424	369	12	1	$\frac{2}{2}$	40	1.287
425	373	1	1	1	40	1.421
426	373	2	1	2	40	1.307
427	373	3	1	2	40	1.26
428	373	4	1	1	40	1.342
429	373	5	1	2	40	1.312
430	373	6	1	1	40	1.313
430	373	7	1	1	40	1.302
432	373	8	1	1	40	1.338
433	373	9	1	1	40	1.301
434	373	10	1	1	40	1.289
435	373	11	1	1	40	1.266
436	373	12	1	2	40	1.27
437	373	12	1	2	40	1.308
438	373	13	1	2	40	1.268
439	373	15	1	2	40	1.259
440	381	15	2	2	40	1.20)
441	381	2	1	1	40	1.401
442	381	3	1	2	40	1.243
443	381	4	1	2	40	1.077
444	381	5	1	1	40	1.278
445	381	6	1	1	40	1.270
446	381	7	1	1	40	1.289
447	381	8	1	1	40	1.399
448	381	9	1	2	40	1.238
448 449	381	10	1	2 1	40 40	1.238
449 450	381	10	1	1	40 40	1.234
430 451	381	11	1	1 2	40 40	1.344
451 452	381	12	1	2	40 40	1.41
452 453		13		2	40 40	
	381		1	2		0.902
454	381	15	1	2	40	1.37

455	390	1	1	2	40	1.277
456	390	2	1	1	40	1.338
457	390	3	1	2	40	1.25
458	390	4	1	2	40	1.211
459	390	5	1	2	40	1.215
460	390	6	1	2	40	1.058
461	390	7	1	2	40	1.082
462	390	8	1	2	40	1.078
463	390	9	1	2	40	1.085
464	390	10	1	1	40	1.009
465	390	11	1	1	40	1.187
466	390	12	1	2	40	1.351
467	390	13	1	2	40	1.303
468	390	14	1	2	40	1.298
469	433	1	1	1	40	1.314
470	433	2	1	1	40	1.225
471	433	3	1	2	40	1.115
472	433	4	1	1	40	1.141
473	433	5	1	2	40	1.202
474	433	6	1	2	40	1.214
475	433	7	1	1	40	1.23
476	433	8	1	1	40	1.194
477	433	9	1	1	40	1.293
478	433	10	1	1	40	1.358
479	433	11	1	2	40	1.168
480	433	12	2		40	
481	433	13	4		40	
482	433	14	1	2	40	1.239
483	433	15	1	2	40	1.252
484	251	1	1	2	200	1.124
485	251	2	1	1	200	1.228
486	251	3	1	1	200	1.142
487	251	4	1	1	200	1.183
488	251	5	1	1	200	1.08
489	251	6	1	1	200	1.19
490	251	7	1	1	200	1.061
491	251	8	1	2	200	1.127
492	251	9	1	2	200	1.064
493	251	10	1	1	200	1.123
494	251	11	1	2	200	1
495	251	12	1	2	200	1.068
496	251	13	1	2	200	0.984
497	258	1	1	1	200	1.198
498	258	2	1	2	200	1.122
499	258	3	1	1	200	1.141
500	258	4	1	1	200	1.157

501	258	5	1	1	200	1.146
502	258	6	2		200	
503	258	7	1	1	200	1.169
504	258	8	1	1	200	1.18
505	258	9	1	2	200	1.127
506	258	10	1	2	200	1.178
507	258	11	1	1	200	1.164
508	258	12	1	1	200	1.121
509	258	13	1	2	200	1.14
510	260	1	1	1	200	1.229
511	260	2	4		200	1
512	260	3	1	1	200	1.255
512	260	4	1	2	200	1.224
515	260	5	1	1	200	1.137
515	260	6	2	1	200	1.157
515	260 260	0 7	1	1	200	1.294
517	260 260	8	1	2	200	1.294
518	260 260	8 9	1	2	200	1.088
518	260 260	10		2	200	1.175
			1	2		
520	260	11	1		200	1.181
521	260	12	1	2	200	1.13
522	260	13	1	1	200	1.186
523	260	14	1	1	200	1.217
524	265	1	1	2	200	1.075
525	265	2	1	1	200	1.049
526	265	3	1	2	200	1.131
527	265	4	1	1	200	1.139
528	265	5	1	1	200	1.118
529	265	6	1	2	200	1.038
530	265	7	1	2	200	1.078
531	265	8	1	2	200	1.064
532	265	9	1	2	200	0.988
533	265	10	1	1	200	0.974
534	265	11	1	2	200	0.978
535	265	12	1	2	200	0.921
536	265	13	1	2	200	1.051
537	272	1	1	1	200	1.041
538	272	2	1	1	200	0.953
539	272	3	1	2	200	1.051
540	272	4	1	2	200	1.016
541	272	5	1	2	200	1.037
542	272	6	1	2	200	1.01
543	272	7	1	1	200	0.953
544	272	8	1	2	200	0.962
545	272	9	1	2	200	1.026
546	272	10	1	1	200	1.127
0.10	212	10	1	1	200	1.14/

547	272	11	1	1	200	0.993
548	272	12	1	1	200	1.122
549	272	13	1	2	200	0.905
550	272	14	1	2	200	1.06
551	274	1	1	1	200	1.135
552	274	2	1	1	200	1.192
553	274	3	1	1	200	1.13
554	274	4	1	2	200	0.983
555	274	5	1	1	200	1.187
556	274	6	1	2	200	0.995
557	274	7	1	2	200	1.115
558	274	8	1	1	200	0.826
559	274	9	1	2	200	0.967
560	274	10	1	1	200	1.15
561	274	11	1	1	200	1.176
562	274	12	1	2	200	1.106
563	274	12	1	1	200	1.167
565 564	274	13	1	2	200	1.038
565	274	15	1	1	200	1.138
565 566	296	1	1	2	200	1.138
567	296	2	1	1	200	1.1
568	290 296	2 3	1	1	200	1.159
569	290 296	4	1	2	200	1.139
570	290 296	4 5	1	1	200	1.124
570 571	290 296	6	1	1	200	1.18
	296 296	8 7	1	1 2	200	
572		8		2		1.063
573	296 296		1	2	200	1.113
574		9	1		200	1.097
575	296	10	1	1	200	1.094
576	296	11	1	2	200	1.06
577	319	1	1	2	200	1.071
578	319	2	1	2	200	1.207
579	319	3	1	2	200	1.175
580	319	4	1	2	200	1.139
581	319	5	1	1	200	1.148
582	319	6	1	1	200	1.144
583	319	7	1	2	200	1.092
584	319	8	2		200	
585	319	9	1	2	200	0.951
586	319	10	1	1	200	1.182
587	319	11	1	1	200	1.146
588	319	12	1	1	200	1.186
589	319	13	1	2	200	0.973
590	319	14	1	1	200	1.073
591	319	15	1	1	200	1.121
592	328	1	2		200	

593	328	2	1	2	200	0.975
594	328	3	1	1	200	1.028
595	328	4	1	2	200	1.007
596	328	5	1	1	200	1.033
597	328	6	4		200	
598	328	7	1	1	200	1.1
599	328	8	1	2	200	0.906
600	328	9	1	1	200	0.843
601	328	10	4		200	
602	328	11	1	1	200	0.99
603	328	12	1	2	200	1.064
604	328	12	1	2	200	1.026
605	328	13	1	1	200	1.002
606	337	1	1	1	200	1.205
607	337	2	1	2	200	1.102
608	337	3	1	1	200	1.231
609	337	4	1	2	200	1.112
610	337	5	1	2	200	1.098
611	337	6	1	2	200	1.098
612	337	0 7	1	1	200	1.087
613	337	8	1	1	200	1.207
613 614		8 9	1	2	200	1.048
	337		1	2		
615	337	10			200	1.173
616	337	11	1	2	200	0.945
617	337	12	1	1	200	1.141
618	339	1	1	1	200	1.163
619	339	2	1	1	200	1.207
620	339	3	1	2	200	1.072
621	339	4	1	1	200	1.09
622	339	5	1	1	200	0.993
623	339	6	1	2	200	1.049
624	339	7	1	2	200	1.073
625	339	8	1	1	200	1.11
626	339	9	1	2	200	1.034
627	339	10	1	1	200	1.056
628	339	11	1	1	200	0.73
629	342	1	1	2	200	0.979
630	342	2	1	1	200	1.148
631	342	3	1	1	200	1.028
632	342	4	1	1	200	1.116
633	342	5	1	1	200	1.162
634	342	6	1	1	200	1.135
635	342	7	1	2	200	1.026
636	342	8	1	2	200	1.071
637	342	9	1	1	200	1.119
638	342	10	1	1	200	1.116

639	342	11	1	1	200	1.134
640	343	1	2		200	
641	343	2	1	2	200	1.097
642	343	3	1	2	200	1.118
643	343	4	1	1	200	1.052
644	343	5	1	1	200	1.133
645	343	6	1	2	200	1.087
646	343	7	1	2	200	0.981
647	343	8	1	1	200	1.049
648	343	9	1	1	200	1.077
649	343	10	1	2	200	1.036
650	343	11	1	2	200	1.159
651	343	12	1	2	200	0.94
652	343	13	1	1	200	1.101
653	343	14	1	2	200	1.057
654	348	1	1	2	200	1.257
655	348	2	1	2	200	1.162
656	348	3	1	1	200	1.255
657	348	4	1	2	200	1.181
658	348	5	1	2	200	1.154
659	348	6	1	1	200	1.161
660	348	7	1	2	200	1.167
661	348	8	1	2	200	1.177
662	348	9	1	2	200	1.142
663	348	10	1	1	200	1.186
664	348	11	1	1	200	1.107
665	348	12	1	2	200	1.149
666	348	13	1	1	200	1.242
667	348	14	1	1	200	1.209
668	353	1	1	1	200	1.198
669	353	2	4		200	
670	353	3	1	1	200	1.181
671	353	4	1	1	200	1.236
672	353	5	1	1	200	1.167
673	353	6	1	1	200	1.104
674	353	7	1	2	200	1.182
675	353	8	1	2	200	1.187
676	353	9	1	2	200	1.158
677	353	10	1	1	200	1.167
678	353	11	1	2	200	1.151
679	353	12	1	1	200	1.182
680	353	13	1	1	200	1.226
681	366	1	1	2	200	1.212
682	366	2	1	1	200	1.263
683	366	3	1	1	200	1.378
684	366	4	1	2	200	1.178

685	366	5	1	2	200	1.27
686	366	6	1	1	200	1.21
687	366	7	1	1	200	1.192
688	366	8	1	1	200	1.226
689	366	9	1	2	200	1.133
690	366	10	2		200	
691	366	11	1	1	200	1.232
692	366	12	1	1	200	1.144
693	366	13	2		200	
694	366	14	1	2	200	1.189
695	371	1	1	1	200	1.091
696	371	2	1	1	200	1.093
697	371	3	1	2	200	0.902
698	371	4	1	2	200	0.976
699	371	5	1	1	200	1.012
700	371	6	1	2	200	0.935
701	371	7	1	2	200	0.987
702	371	8	1	2	200	1.002
703	371	9	1	1	200	1.022
704	371	10	1	2	200	0.984
705	371	11	1	2	200	1.001
706	371	12	1	1	200	1.055
707	371	12	1	1	200	0.973
708	371	19	1	1	200	1.068
709	371	15	1	2	200	0.897
710	382	1	1	2	200	1.548
711	382	2	1	2	200	1.325
712	392	1	1	1	200	1.206
712	392	2	1	1	200	1.253
713	392	3	1	2	200	1.235
715	392	4	1	1	200	1.295
716	392	5	1	2	200	1.087
717	392	6	1	2	200	1.052
718	392	0 7	1	1	200	1.032
719	392	8	1	2	200	1.135
720	392 392	8 9	1	2	200	1.133
720	392 392	10	1	1	200	1.125
721	392 392	10	1	1	200	1.232
722	392 392	11	1	1 2		
				1	200	1.132
724 725	392 202	13	1	1 2	200	1.168
725 726	392 202	14	1		200	1.176
726 727	392 202	15	1	1	200	1.24
727 728	392 402	16	1	2	200	1.198
728 720	402	1	1	2	200	0.903
729 720	402	2	1	1	200	1.208
730	402	3	1	1	200	1.093

731	402	4	1	2	200	1.078
732	402	5	1	1	200	1.052
733	402	6	1	2	200	1.098
734	402	7	1	2	200	0.941
735	402	8	1	1	200	1.109
736	402	9	1	2	200	1.051
737	402	10	1	2	200	1.164
738	402	11	1	1	200	1.135
739	402	12	1	2	200	0.933
740	402	13	2	-	200	0.900
741	402	14	1	2	200	1.061
742	402	15	1	1	200	1.071
743	402	16	1	1	200	1.072
744	420	1	1	1	200	1.294
745	420	2	1	1	200	1.289
746	420	3	2	1	200	1.20)
747	420	4	1	1	200	1.284
748	420	5	2	1	200	1.204
749	420	6	1	2	200	1.16
750	420	7	1	1	200	1.144
751	420	8	1	1	200	1.263
752	420	8 9	1	1	200	1.203
753	420	10	1	1	200	1.239
754	420	10	1	2	200	1.143
755	420	11	1	2 1	200	1.143
756	420 420	12	1	1	200	1.192
757	420 231	13	2	1	1000	1.300
758	231	1 2	1	1	1000	1 1 1 2
758 759	231	2 3	1		1000	1.112 0.932
7 <i>59</i> 760	231	3 4	1	2 2	1000	1.063
	231	4 5	1	2	1000	1.003
761 762		5 6		2		
762	231		1	2	1000	0.955
763	231	7 8	1 1	1 2	1000	1.051
764 765	231 231				1000	1.036
765		9	1	1	1000	1.038
766	231	10	1	1	1000	1.046
767	243	1	1	2	1000	0.982
768	243	2	1	2	1000	0.96
769	243	3	1	2	1000	1.016
770	243	4	1	1	1000	1.13
771	243	5	1	2	1000	0.949
772	243	6	1	2	1000	1.046
773	243	7	1	1	1000	1.003
774	243	8	1	1	1000	0.998
775	243	9	1	1	1000	1.001
776	243	10	1	2	1000	1.077

777	243	11	1	2	1000	1.028
778	243	12	1	2	1000	1.041
779	243	13	1	2	1000	1.018
780	243	14	1	1	1000	1.037
781	244	1	1	1	1000	0.934
782	244	2	1	2	1000	0.537
783	244	3	1	2	1000	0.862
784	244	4	1	2	1000	0.746
785	244	5	1	2	1000	0.889
786	244	6	1	1	1000	0.942
787	244	7	1	2	1000	0.948
788	244	8	1	1	1000	0.885
789	244	9	1	2	1000	1
790	244	10	1	1	1000	0.925
791	244	11	1	1	1000	1.08
792	244	12	2	1	1000	1.00
793	255	1	1	2	1000	1.067
794	255	2	1	2	1000	1.13
795	255	3	1	1	1000	1.081
796	255	4	1	1	1000	1.073
797	255	5	1	1	1000	1.056
798	255	6	1	2	1000	1.034
799	255	7	1	1	1000	1.087
800	255	8	1	2	1000	1.078
800	255	9	1	2	1000	1.078
802	255	10	1	1	1000	1.038
802	255	10	1	1	1000	1.133
803	255	11	1	1	1000	1.133
804 805	233	12	1	1	1000	1.067
805	264 264	2	1	1	1000	0.901
800 807	264	3	1	1 2	1000	1.057
807	264 264	3 4		2	1000	0.98
	264 264	4 5	1	2		
809 810	264 264	5 6	1 1	2	1000	1.036 0.856
				2	1000	
811	264	7	1		1000	0.937
812	264	8	1	1	1000	0.95
813	264	9	1	2	1000	1.09
814	264	10	1	1	1000	1.162
815	264	11	1	2	1000	1.074
816	264	12	1	1	1000	1.004
817	264	13	1	1	1000	1.083
818	264	14	1	2	1000	1.001
819	276	1	1	1	1000	0.862
820	276	2	1	2	1000	0.93
821	276	3	1	2	1000	0.786
822	276	4	1	2	1000	0.783

823	276	5	4		1000	
824	276	6	1	2	1000	0.809
825	276	7	1	1	1000	0.737
826	276	8	1	1	1000	0.964
827	276	9	1	1	1000	1
828	276	10	1	1	1000	0.84
829	276	11	1	1	1000	0.85
830	276	12	1	1	1000	0.695
831	276	13	1	1	1000	0.934
832	276	14	1	2	1000	0.79
833	276	15	1	1	1000	0.915
834	294	1	1	1	1000	1.047
835	294	2	1	1	1000	1.164
836	294	3	1	1	1000	1.091
837	294	4	1	2	1000	0.917
838	294	5	1	1	1000	1.065
839	294	6	1	1	1000	1.059
840	294	7	1	2	1000	0.934
841	294	8	1	1	1000	1.042
842	294	9	1	2	1000	0.956
843	294	10	1	2	1000	1.029
844	294	11	1	1	1000	0.958
845	294	11	1	2	1000	0.938
843 846	294	12	1	2	1000	1.032
840 847	294	13	1	2	1000	0.951
847 848	294 294	14	1	2	1000	1.02
848 849	305	13		1		
			1		1000	1.197
850	305	2 3	1	1	1000	1.021
851	305		1	2	1000	1.031
852	305	4	1	2	1000	0.924
853	305	5	1	2	1000	1.106
854	305	6	1	1	1000	1.029
855	305	7	1	1	1000	1.127
856	305	8	1	1	1000	1.23
857	305	9	1	1	1000	1.054
858	305	10	1	2	1000	0.996
859	305	11	1	2	1000	0.952
860	305	12	1	1	1000	0.991
861	309	1	1	2	1000	1.045
862	309	2	1	2	1000	1.062
863	309	3	1	1	1000	1.165
864	309	4	1	1	1000	1.076
865	309	5	1	1	1000	1.106
866	309	6	1	2	1000	1.054
867	309	7	1	2	1000	1.097
868	309	8	1	2	1000	1.09

869	309	9	1	1	1000	1.086
870	309	10	1	1	1000	1.097
871	309	11	1	1	1000	1.153
872	309	12	1	2	1000	1.183
873	309	13	1	2	1000	1.019
874	309	14	1	1	1000	1.072
875	309	15	1	1	1000	1.017
876	309	16	1	1	1000	1.105
877	317	1	1	1	1000	1.046
878	317	2	1	1	1000	1.071
879	317	3	1	1	1000	
880	317	4	1	1	1000	1.055
881	317	5	1	1	1000	1.054
882	317	6	1	1	1000	1.08
883	317	7	1	2	1000	0.902
884	317	8	1	1	1000	0.806
885	317	9	1	2	1000	0.982
886	317	10	1	2	1000	1.034
887	317	11	1	1	1000	
888	317	12	1	1	1000	1.018
889	317	13	1	2	1000	1.031
890	317	14	1	1	1000	1.006
891	325	1	1	1	1000	1.076
892	325	2	2		1000	
893	325	3	1	2	1000	1.156
894	325	4	1	1	1000	1.128
895	325	5	1	2	1000	1.129
896	325	6	1	2	1000	1.082
897	325	7	1	2	1000	1.176
898	325	8	1	1	1000	1.037
899	325	9	1	1	1000	1.187
900	325	10	1	1	1000	1.08
901	325	11	1	1	1000	1.134
902	325	12	1	1	1000	1.068
903	325	13	1	1	1000	1.003
904	325	14	1	1	1000	0.935
905	325	15	1	2	1000	0.985
906	340	1	1	1	1000	1.071
907	340	2	1	1	1000	1.106
908	340	3	1	2	1000	1.078
909	340	4	1	1	1000	1.112
910	340	5	1	2	1000	1.045
911	340	6	1	1	1000	1.017
912	340	7	2		1000	
913	340	8	1	1	1000	1.09
914	340	9	2		1000	

915	340	10	1	2	1000	1.143
916	340	11	1	1	1000	1.138
917	340	12	1	2	1000	1.069
918	340	13	1	1	1000	1.056
919	365	1	1	1	1000	0.846
920	365	2	1	2	1000	0.829
921	365	3	1	2	1000	0.937
922	365	4	1	2	1000	0.688
923	365	5	1	2	1000	0.868
924	365	6	1	2	1000	0.57
925	365	7	1	1	1000	0.839
925 926	365	8	1	2	1000	0.945
920 927	365	9	1	1	1000	0.943
927 928	365	10	1	1	1000	0.902
928 929	365	10	2	1	1000	0.010
929 930	303 374	1	2		1000	
930 931		1 2	2	r	1000	1.022
	374	2 3		2 2		
932	374	3 4	1		1000	1.048
933	374		1	1	1000	1.091
934	374	5	1	1	1000	1.048
935	374	6	1	1	1000	1.15
936	374	7	1	1	1000	1.201
937	374	8	1	1	1000	1.068
938	374	9	1	1	1000	1.092
939	374	10	1	1	1000	1.077
940	377	1	1	2	1000	1.189
941	377	2	1	1	1000	1.129
942	377	3	1	2	1000	1.049
943	377	4	1	2	1000	1.127
944	377	5	1	1	1000	0.985
945	377	6	2		1000	
946	377	7	1	1	1000	1.056
947	377	8	1	1	1000	1.248
948	377	9	1	1	1000	1.02
949	377	10	1	2	1000	1.188
950	377	11	1	2	1000	1.082
951	377	12	1	2	1000	1.025
952	377	13	1	2	1000	1.155
953	389	1	1	1	1000	1.087
954	389	2	1	2	1000	1.025
955	389	3	1	2	1000	1.074
956	389	4	1	2	1000	1.14
957	389	5	2		1000	
958	389	6	1	2	1000	1.015
959	389	7	1	2	1000	0.945
960	389	8	1	1	1000	1.191
		-	-	-		

961	389	9	1	1	1000	1.182
962	389	10	4		1000	
963	389	11	2		1000	
964	400	1	1	2	1000	1.276
965	400	2	1	2	1000	1.375
966	400	3	1	1	1000	1.341
967	400	4	1	1	1000	1.468
968	400	5	1	2	1000	1.349
969	400	6	2		1000	
970	400	7	1	1	1000	1.249
971	400	8	2		1000	
972	400	9	1	1	1000	1.358
973	400	10	1	1	1000	1.368
974	400	11	1	1	1000	1.415
975	400	12	1	2	1000	1.3
976	427	1	1	2	1000	1.119
977	427	2	1	1	1000	1.235
978	427	3	1	1	1000	1.222
979	427	4	1	1	1000	1.153
980	427	5	1	1	1000	1.078
981	427	6	1	2	1000	1.032
982	427	7	1	1	1000	0.975
983	427	8	1	1	1000	1.092
984	427	9	1	1	1000	1.217
985	427	10	1	2	1000	1.121
986	427	11	1	2	1000	1.105
987	427	12	1	1	1000	1.172
988	427	13	2		1000	
989	427	14	1	1	1000	1.188
990	428	1	1	2	1000	1.017
991	428	2	1	1	1000	0.965
992	428	3	1	2	1000	1.044
993	428	4	1	2	1000	0.993
994	428	5	1	2	1000	0.971
995	428	6	1	1	1000	1.011
996	428	7	1	2	1000	0.928
997	428	8	1	2	1000	0.956
998	428	9	2		1000	
999	428	10	1	2	1000	1.069
1000	428	11	1	2	1000	0.935
1001	428	12	1	2	1000	0.982
1002	445	1	1	1	1000	0.889
1003	445	2	4		1000	
1004	445	3	1	2	1000	0.926
1005	445	4	1	1	1000	1.105
1006	445	5	1	1	1000	1.058

1007	445	6	1	2	1000	
1008	445	7	1	2	1000	1.039
1009	445	8	1	1	1000	1.067
1010	445	9	1	2	1000	0.983
1011	445	10	1	1	1000	1.056
1012	445	11	1	2	1000	0.977
1013	445	12	1	2	1000	0.975
1014	445	13	1	2	1000	0.993
1015	445	14	2		1000	
1016	445	15	1	2	1000	1.021
1017	445	16	1	2	1000	0.53
1018	445	17	1	1	1000	1.03

Some 6's were 8's and have been corrected for fetalwt Animal 445 Pup 14 had status 2 and FetalSex=2 in report, it was changed to FetalSex missing

1 2

# 1 Appendix 2. BMC Modeling for Acute ReV

	Polynomial Model. (Version: 2.12; Date: 02/20/2007) Input Data File: C:\BMDS\MATERNAL TOXICITY\EXTRAGESTATIONAL WEIGH
G	AIN FOUR DOSES.(d)
w	Gnuplot Plotting File: C:\BMDS\MATERNAL TOXICITY\EXTRAGESTATIONAL EIGHT GAIN FOUR DOSES.plt
_	Fri Jun 15 12:02:13 2007
B ~~	MDS MODEL RUN
r	The form of the response function is:
•	$Y[dose] = beta_0 + beta_1 * dose + beta_2 * dose^2 +$
]	Dependent variable = MEAN
]	Independent variable = dose
1	rho is set to 0
	Signs of the polynomial coefficients are not restricted
1	A constant variance model is fit
,	Total number of dose groups $= 4$
	Total number of records with missing values $= 0$
	Maximum number of iterations = $250$
	Relative Function Convergence has been set to: 1e-008
	Parameter Convergence has been set to: 1e-008
	-
	Default Initial Parameter Values
	alpha = 2.84578
	rho = 0 Specified
	$beta_0 = 7.46597$ beta_1 = -0.00778544
	beta $2 = 6.23087e-006$
	00m_2 0.250070-000
	Asymptotic Correlation Matrix of Parameter Estimates

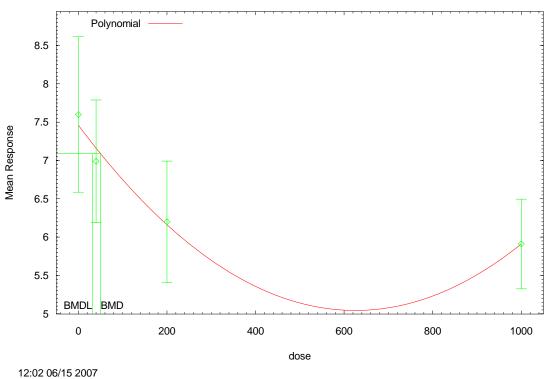
1 2	(*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,
3	and do not appear in the correlation matrix )
4 5	alpha beta 0 beta 1 beta 2
5 6	alpha beta_0 beta_1 beta_2
7	alpha 1 -1.4e-009 -4.1e-010 1.7e-010
8	•
9	beta_0 -1.4e-009 1 -0.7 0.64
10 11	beta 1 -4.1e-010 -0.7 1 -0.99
12	beta_1 -4.10-010 -0.7 1 -0.7
13	beta_2 1.7e-010 0.64 -0.99 1
14	
15 16	
10 17	Parameter Estimates
18	
19	95.0% Wald Confidence Interval
20	Variable Estimate Std. Err. Lower Conf. Limit Upper Conf. Limit
21	alpha 2.71186 0.434246 1.86076 3.56297
22 23	beta_0 7.46083 0.307655 6.85783 8.06382 beta_1 -0.00772578 0.00307077 -0.0137444 -0.00170717
23 24	beta 2 $6.17626e-006$ $2.89958e-006$ $4.93184e-007$ $1.18593e-005$
25	
26	
27	
28 29	Table of Data and Estimated Values of Interest
29 30	Dose N Obs Mean Est Mean Obs Std Dev Est Std Dev Scaled Res.
31	
32	
33	0 18 7.6 7.46 2.04 1.65 0.359
34	40 19 6.99 7.16 1.66 1.65 -0.454
35	200 21 6.2 6.16 1.74 1.65 0.104
36	1000 20 5.91 5.91 1.25 1.65 -0.00354
37 38	
38 39	
40	Model Descriptions for likelihoods calculated
41	r
42	
43	Model A1: $Yij = Mu(i) + e(ij)$
44	$Var\{e(ij)\} = Sigma^2$
45	
46 47	Model A2: $Yij = Mu(i) + e(ij)$ $Var \{e(ij)\} = Sigma(i)^2$
4/	$vai(0) = \delta(0) = \delta(0) = \delta(0) = \delta(0)$

1							
2	Model A3:	Yij = Mu(i) + e(i)	j)				
3	$Var{e(ij)} = Sigma^2$						
4	Model A3 uses any fixed variance parameters that						
5	were specified by the user						
6	1	5					
7	Model R:	Yi = Mu + e(i)					
8		$\} = Sigma^2$					
9		)) ~ 8 ~					
10							
11	I	Likelihoods of Inte	rest				
12							
13	Model	Log(likelihood)	# P	aram's	AIC		
14	Al	-77.734504	5	165.4	69009		
15		-75.503795	8	167.0			
16	A3	-77.734504		165.4	69009		
17	fitted	-77.907806	4	163.8	15611		
18	R	-83.514230					
19	R	05.51 1250	-	171.02	20100		
20	Exi	planation of Tests					
21	12/1						
22	Test 1. Do res	nonses and/or vari	ances	differ	among Dose levels?		
23	(A2 vs. H		uneet	, annor			
24		ariances Homogen	eous?	) (A1 vs	s A2)		
25		riances adequately					
26		he Model for the N					
27					Test 2 will be the same.)		
28			1 1 0 0				
29	Т	ests of Interest					
30	-						
31	Test -2*log	(Likelihood Ratio)	Tes	t df	p-value		
32		(			F		
33	Test 1	16.0209 6	0	0.01364			
34	Test 2	4.46142 3		0.2158			
35				0.2158			
36	Test 4			0.556			
37							
38	The p-value for	Test 1 is less than	n .05.	There	appears to be a		
39					among the dose levels		
40		priate to model the			8		
41	11 1						
42	The p-value for	r Test 2 is greater t	han .1	I. A ho	mogeneous variance		
43	<u> </u>	to be appropriate h			5		
44		11 -F	-				
45	The p-value for	Test 3 is greater t	han .1	1. The	modeled variance appears		
46	to be appropria		,-				
47	TT T						

	1,3-Butadiene PROPOSED DSD Page 104				
1	The p-value for Test 4 is greater than .1. The model chosen seems				
2	to adequately describe the data				
3					
4	Benchmark Dose Computation				
5					
6	Specified effect = $0.05$				
7					
8	Risk Type = Relative risk				
9					
10	$Confidence \ level = 0.95$				
11					
12	BMD = 50.3086				
13					
14	BMDL = 31.2322				
15	DMDL commutation foiled for one on more point on the DMDL sums. The DMDL				
16	BMDL computation failed for one or more point on the BMDL curve. The BMDL curve will not be				

plotted 17

18



Polynomial Model with 0.95 Confidence Level

19 20

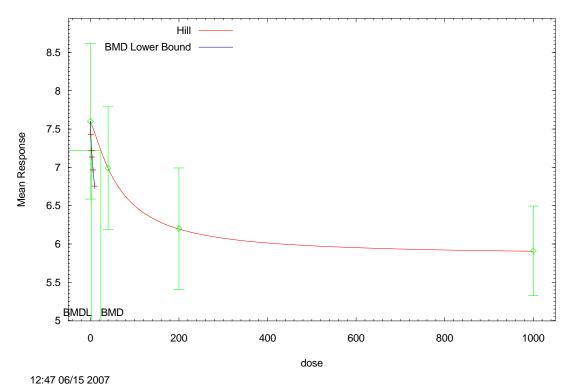
Hill Model. (Version: 2.12; Date: 02/20/2007) Input Data File: C:\BMDS\ MATERNAL TOXICITY\EXTRAGESTATIONAL V GAIN FOUR DOSES.(d) Gnuplot Plotting File: C:\BMDS\ MATERNAL TOXICITY\EXTRAGESTATIO WEIGHT GAIN FOUR DOSES.plt Fri Jun 15 12:47:56 2007			
BMDS	MODEL RUN		
The fo	orm of the response function is:		
Y[dos	$e] = intercept + v*dose^n/(k^n + dose^n)$		
	ident variable = MEAN		
	endent variable = dose		
	set to 0 parameter restricted to be greater than 1		
	stant variance model is fit		
Total	number of dose groups $= 4$		
	number of records with missing values $= 0$		
	num number of iterations = 250 ve Function Convergence has been set to: 1e-008		
	eter Convergence has been set to: 1e-008		
	Default Initial Parameter Values		
	alpha = 2.84578		
	rho = 0 Specified		
	intercept = $7.6$		
	v = -1.69 n = 1.31786		
	k = 87.5949		
	An and the Completion Metric of Decompton E. (1)		
F	Asymptotic Correlation Matrix of Parameter Estimates		
(	*** The model parameter(s) -rho		
(	have been estimated at a boundary point, or have been specified by the user,		
	and do not appear in the correlation matrix )		

1	alp	ha int	ercept	v	n	k	
2 3	alpha	1 8.	.3e-010	1.9e-008	9.6e-0	09	-2.2e-008
4 5	intercept 8.	3e-010	1	-0.59	-0.18		-0.38
6 7	v 1.9e	-008	-0.59	1	0.7	-0.2	34
8 9		-009	-0.18	0.7	1		
10							
11 12	k -2.2e	-008	-0.38	-0.34	-0.4		1
13 14							
15 16		Р	arameter	Estimates			
10				95 0%	wald C	onfic	lence Interval
18	Variable	Est	imate				f. Limit Upper Conf. Limit
19	alpha		984	0.43232		8525	
20	intercept			0.387287		4093	
21	v	-1.745		).695364			
22	n	1.255	83	1.43345	-1.5	5368	4.06534
23	k	65.59	03 6	6.7166	-65.1	692	196.355
25 26 27 28 29 30	Table of Da Dose N					ev Es	st Std Dev Scaled Res.
31 32	0 18	7.6	7.6	2.04	1.64 -:	5 18e	-009
33			6.99		1.64		6e-008
34	200 21			1.74			e-008
35	1000 20			1.25			
36 37 38	Degrees of fre						
39 40							
41 42 43	Model Descri	ptions f	or likelih	oods calcu	lated		
44 45	Model A1: Var{e(i	Yij = ij)} = Si	Mu(i) + gma^2	e(ij)			
46 47	Model A2:	Yij =	Mu(i) +	e(ij)			

	1,3-Butadier Page 107	ne PROPOSED D	SD				
1 2	Var{	e(ij) = Sigma(i) <sup>2</sup>	2				
3	Model A3:	Yij = Mu(i) +	e(ij)				
4		e(ij) = Sigma <sup>2</sup>					
5		3 uses any fixed v	ariance	e parameters that			
6	were spe	cified by the user					
7							
8	Model R: $Yi = Mu + e(i)$						
9	Var	$\{e(i)\} = Sigma^2$					
10							
11		T :11:1 1 C1	·				
12 13		Likelihoods of I	nterest	t			
13 14	Mod	lel Log(likeliho	od) #	Param's AIC			
15		-77.734504		5 165.469009			
16		-75.503795					
17		-77.734504		5 165.469009			
18	fitted	-77.734504	5	5 165.469009			
19	R			2 171.028460			
20							
21							
22		Explanation of Tes	sts				
23							
24		<u>^</u>	varianc	ces differ among Dose levels?			
25	(A2 v			$\mathbf{P}(\mathbf{A} 1, \mathbf{A} 2)$			
26	Test 2: Are Variances Homogeneous? (A1 vs A2) Test 3: Are variances adequately modeled? (A2 vs. A3)						
27 28							
28 29	Test 4: Does the Model for the Mean Fit? (A3 vs. fitted) (Note: When rho=0 the results of Test 3 and Test 2 will be the same.)						
30	(INDIC: WII	en mo-o the resul	13 01 10	est 5 and Test 2 will be the same.)			
31		Tests of Interest					
32							
33	Test -2*	log(Likelihood Ra	tio) T	est df p-value			
34			,				
35	Test 1	16.0209	6	0.01364			
36	Test 2	4.46142	3	0.2158			
37	Test 3	4.46142	3	0.2158			
38	Test 4	5.68434e-014	0	NA			
39	TT1 1	C T (1 1 )	1 0				
40				5. There appears to be a			
41 42	difference between response and/or variances among the dose levels It seems appropriate to model the data						
42	it seems app		une uai	ta			
44	The <b>p-</b> value	for Test 2 is great	er than	n .1. A homogeneous variance			
45		ars to be appropria					
46	Tr r	Tr P					
47							

	1,3-Butadiene PROPOSED DSD Page 108				
1	The p-value for Test 3 is greater than .1. The modeled variance appears				
2	to be appropriate here				
3					
4	NA - Degrees of freedom for Test 4 are less than or equal to 0. The Chi-Square				
5	test for fit is not valid				
6					
7					
8	Benchmark Dose Computation				
9					
10	Specified effect = $0.05$				
11					
12	Risk Type = Relative risk				
13	0 = 6 + 1 = 0.05				
14 15	$Confidence \ level = 0.95$				
15 16	BMD = 23.6911				
10	BIVID = 23.0911				
17	BMDL = 2.8111				
18 19	$D_{W}DL = 2.0111$				
17					

#### Hill Model with 0.95 Confidence Level



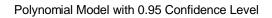
20

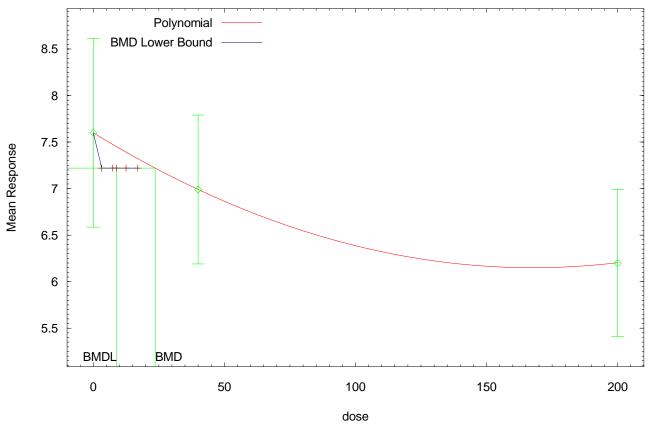
Input D	ata File: C:\B		.12; Date: 02/20/2007) ATERNAL TOXICITY\EXTRAGESTATIONAL WEIGI
GAIN THREE I			
Gnupic WEIGHT GAIN			OS\ MATERNAL TOXICITY\EXTRAGESTATIONAL
		Lo.pit	Fri Jun 15 13:14:35 2007
BMDS MODEI			
The form of th			~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~
Y[dose] = beta	$_0 + beta_1 * deta_1$	ose + beta	$a_2*dose^2 +$
Dependent var		[	
Independent va	riable = dose		
rho is set to 0	le maneial a a aff	Ciaianta an	a mat martinistad
A constant var			e not restricted
A constant var	ance model is	111	
Total number of	of dose groups	= 3	
Total number of			values $= 0$
Maximum nun			
Relative Funct	on Convergen	ce has bee	en set to: 1e-008
Parameter Con	vergence has b	been set to	: 1e-008
	14 Luitin 1 Der	• • • • • • • • • • •	
	lt Initial Paran lpha = 3.28		
	1	) Specifie	ed
h	0 = 7		
	$eta_{1} = -0.01$		
	eta 2 = 5.1562		
	—		
Asymptot	ic Correlation	Matrix of	Parameter Estimates
( *** The	model parame	oter(s) _rh	0
	*		dary point, or have been specified by the user,
	not appear in		
	1		,
alpha	beta 0	beta 1	beta 2

1 2 alpha 1 -4.5e-008 9.1e-009 3.8e-009 3 4 beta 0 -4.5e-008 1 -0.7 0.63 5 6 beta 1 9.1e-009 -0.7 1 -0.99 7 8 beta 2 3.8e-009 0.63 -0.99 1 9 10 11 12 Parameter Estimates 13 14 95.0% Wald Confidence Interval Variable Lower Conf. Limit Upper Conf. Limit 15 Estimate Std. Err. alpha 3.11897 4.25413 16 0.579177 1.9838 17 beta 0 7.6 0.416264 6.78414 8.41586 18 beta 1 -0.0173125 0.0177901 -0.0521805 0.0175555 19 beta 2 5.15625e-005 8.28263e-005 -0.000110774 0.000213899 20 21 22 23 Table of Data and Estimated Values of Interest 24 25 Dose Obs Mean Est Mean Obs Std Dev Est Std Dev Scaled Res. Ν 26 -----27 28 0 18 7.6 7.6 2.04 1.77 2.42e-007 29 40 - 19 6.99 6.99 1.66 1.77 1.56e-007 200 21 30 6.2 6.2 1.74 1.77 -3.67e-007 31 32 Degrees of freedom for Test A3 vs fitted  $\leq 0$ 33 34 35 36 Model Descriptions for likelihoods calculated 37 38 39 Yij = Mu(i) + e(ij)Model A1: 40  $Var{e(ij)} = Sigma^2$ 41 42 Model A2: Yij = Mu(i) + e(ij)43  $Var\{e(ij)\} = Sigma(i)^2$ 44 45 Model A3: Yij = Mu(i) + e(ij) $Var{e(ij)} = Sigma^2$ 46 47 Model A3 uses any fixed variance parameters that

	Page 111	ROPUSED DS.	D				
1 2	were specifi	ed by the user					
3	Model R:	Yi = Mu + e(i)					
4		$\} = Sigma^2$					
5		,, 0					
6							
7	Ι	likelihoods of In	terest	t			
8							
9	Model	Log(likelihoo	d) #	Pa	aram's AIC		
10	A1	-61.987540		4	131.975080		
11	A2	-61.553857	(	6	135.107715		
12	A3	-61.987540	4	4	131.975080		
13	fitted	-61.987540	2	1	131.975080		
14	R	-64.921732	2		133.843464		
15							
16	-						
17	Exp	planation of Test	S				
18	T (1 D	1/					
19 20			arianc	es	differ among Dose levels?		
20 21	(A2 vs. F	· · · · · · · · · · · · · · · · · · ·		. <u>_</u> 9	$(A1 \times A2)$		
21 22		ariances Homogo			(A1 vs A2) eled? (A2 vs. A3)		
22					Fit? (A3 vs. fitted)		
23 24					3 and Test 2 will be the same.)		
25	(1000 When I	ino o the results	01 1	0.51	5 and 16st 2 will be the sume.)		
26	Τe	ests of Interest					
27	1						
28	Test -2*log	(Likelihood Rati	io) T	est	df p-value		
29		(	-)		r t		
30	Test 1	6.73575	4	(	0.1505		
31	Test 2	0.867366	2		0.6481		
32	Test 3	0.867366	2		0.6481		
33	Test 4 5.	.82645e-013	0		NA		
34							
35					5. There may not be a		
36		*			ances among the dose levels		
37	Modelling the o	data with a dose/	respo	ns	e curve may not be appropriate		
38			.1	1			
39	· ·	-			. A homogeneous variance		
40	model appears	to be appropriate	e here	,			
41							
42	The n value for	Tost 2 is greate	r than	. 1	The modeled verience encours		
43 44	The p-value for Test 3 is greater than .1. The modeled variance appears to be appropriate here						
44 45	to be appropria						
46	NA - Degrees o	of freedom for Te	est 4 :	are	e less than or equal to 0. The Chi-Square		
47	test for fit is		-50 10	~ ~	The share of equal to of the one offune		

1	
2	Benchmark Dose Computation
3	
4	Specified effect = $0.05$
5	
6	Risk Type = Relative risk
7	Confidence level = $0.95$
8 9	Confidence level = 0.95
10	BMD = 23.6096
11	
12	BMDL = 8.90895
13	





13:14 06/15 2007

1

# 2 2.2 Body Weight Gain (GD 11-16)

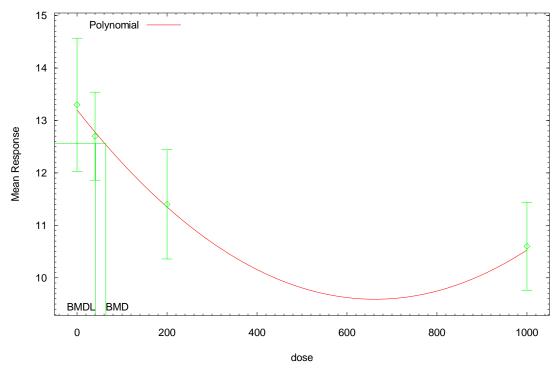
Fri Jun 15 10:27:22 2007         BMDS MODEL RUN         The form of the response function is: $Y[dose] = beta_0 + beta_1 * dose + beta_2 * dose^2 +$ Dependent variable = MEAN         Independent variable = Dose         rho is set to 0         Signs of the polynomial coefficients are not restricted         A constant variance model is fit         Total number of dose groups = 4         Total number of records with missing values = 0         Maximum number of iterations = 250         Relative Function Convergence has been set to: 1e-008         Parameter Convergence has been set to: 1e-008         Default Initial Parameter Values         alpha = 4.47026         rho = 0       Specified         beta_0 = 13.2228         beta_1 = -0.0108878         beta_2 = 8.2658e-006         Asymptotic Correlation Matrix of Parameter Estimates         (*** The model parameter(s) -rho         have been estimated at a boundary point, or have been specified by the user, and do not appear in the correlation matrix )		OSES.(d) Gnuplot Plotting File: C:\BMDS\ MATERNAL TOXICITY\BODYWEIGHT GAIN GI OUR DOSES.plt
The form of the response function is: Y[dose] = beta_0 + beta_1*dose + beta_2*dose^2 + Dependent variable = MEAN Independent variable = Dose tho is set to 0 Signs of the polynomial coefficients are not restricted A constant variance model is fit Total number of dose groups = 4 Total number of iceords with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	=	1
Y[dose] = beta_0 + beta_1*dose + beta_2*dose^2 + Dependent variable = MEAN Independent variable = Dose rho is set to 0 Signs of the polynomial coefficients are not restricted A constant variance model is fit Total number of dose groups = 4 Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	В	MDS MODEL RUN
Y[dose] = beta_0 + beta_1*dose + beta_2*dose^2 + Dependent variable = MEAN Independent variable = Dose rho is set to 0 Signs of the polynomial coefficients are not restricted A constant variance model is fit Total number of dose groups = 4 Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	~~	
Dependent variable = MEAN Independent variable = Dose rho is set to 0 Signs of the polynomial coefficients are not restricted A constant variance model is fit Total number of dose groups = 4 Total number of iccords with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	,	The form of the response function is:
Independent variable = Dose rho is set to 0 Signs of the polynomial coefficients are not restricted A constant variance model is fit Total number of dose groups = 4 Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,		$Y[dose] = beta_0 + beta_1 * dose + beta_2 * dose^2 +$
<pre>rho is set to 0 Signs of the polynomial coefficients are not restricted A constant variance model is fit Total number of dose groups = 4 Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values</pre>	]	Dependent variable = MEAN
Signs of the polynomial coefficients are not restricted A constant variance model is fit Total number of dose groups = 4 Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	]	Independent variable = Dose
A constant variance model is fit Total number of dose groups = 4 Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	1	rho is set to 0
Total number of dose groups = 4 Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	ł	Signs of the polynomial coefficients are not restricted
Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,		A constant variance model is fit
Total number of records with missing values = 0 Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	,	Total number of dose groups $= 4$
Maximum number of iterations = 250 Relative Function Convergence has been set to: 1e-008 Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,		
Parameter Convergence has been set to: 1e-008 Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	]	Maximum number of iterations = $250$
Default Initial Parameter Values alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,	]	Relative Function Convergence has been set to: 1e-008
<pre>alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,</pre>	]	Parameter Convergence has been set to: 1e-008
<pre>alpha = 4.47026 rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,</pre>		
<pre>rho = 0 Specified beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,</pre>		Default Initial Parameter Values
<pre>beta_0 = 13.2228 beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,</pre>		alpha = 4.47026
<pre>beta_1 = -0.0108878 beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,</pre>		rho = 0 Specified
<pre>beta_2 = 8.2658e-006 Asymptotic Correlation Matrix of Parameter Estimates (*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,</pre>		
Asymptotic Correlation Matrix of Parameter Estimates ( *** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,		
(*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,		$beta_2 = 8.2658e-006$
(*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,		
(*** The model parameter(s) -rho have been estimated at a boundary point, or have been specified by the user,		
have been estimated at a boundary point, or have been specified by the user,		Asymptotic Correlation Matrix of Parameter Estimates
have been estimated at a boundary point, or have been specified by the user,		
and do not appear in the correlation matrix )		
		and do not appear in the correlation matrix )
alpha beta 0 beta 1 beta 2		

1 2 alpha 1 -5.8e-010 -6.2e-009 6.3e-009 3 4 beta 0 -5.8e-010 1 -0.7 0.64 5 6 beta 1 -6.2e-009 -0.7 1 -0.99 7 8 beta 2 6.3e-009 0.64 -0.99 1 9 10 Parameter Estimates 11 12 13 95.0% Wald Confidence Interval 14 Variable Estimate Lower Conf. Limit Upper Conf. Limit Std. Err. 4.245 5.57728 15 alpha 0.679745 2.91273 beta 0 0.384919 13.9743 16 13.2198 12.4654 17 beta 1 -0.0108534 0.00384196 -0.0183835 -0.00332333 18 beta 2 8.23434e-006 3.62777e-006 1.12404e-006 1.53446e-005 19 20 21 Table of Data and Estimated Values of Interest 22 23 Dose N Obs Mean Est Mean Obs Std Dev Est Std Dev Scaled Res. 24 ----- ----------\_\_\_\_\_ ---25 26 0 18 13.2 2.55 0.165 13.3 2.06 27 40 19 12.8 12.7 1.74 2.06 -0.209 28 200 21 11.4 11.4 2.29 2.06 0.0478 29 1000 20 10.6 10.6 1.79 2.06 -0.00163 30 31 32 Model Descriptions for likelihoods calculated 33 34 Model A1: Yij = Mu(i) + e(ij)35  $Var{e(ij)} = Sigma^2$ 36 37 Model A2: Yij = Mu(i) + e(ij)38  $Var{e(ij)} = Sigma(i)^2$ 39 40 Model A3: Yij = Mu(i) + e(ij) $Var{e(ij)} = Sigma^2$ 41 Model A3 uses any fixed variance parameters that 42 43 were specified by the user 44 45 Model R: Yi = Mu + e(i) $Var{e(i)} = Sigma^2$ 46 47

1							
2	Likelihoods of Interest						
3							
4	Model	Log(likelihood)	# Pa	aram's AIC			
5	A1	-95.347306	5	200.694612			
6	A2	-93.364104	8	202.728208			
7	A3	-95.347306	5	200.694612			
8	fitted	-95.383968	4	198.767936			
9	R	-104.387527	2	212.775055			
10							
11							
12	Exp	planation of Tests					
13	_						
14	Test 1: Do res	ponses and/or variation	ances	differ among Dose levels?			
15	(A2 vs. H	R)					
16	Test 2: Are V	ariances Homogen	eous?	(A1 vs A2)			
17		riances adequately					
18		he Model for the M					
19	(Note: When	rho=0 the results of	f Test	3 and Test 2 will be the same.)			
20							
21	Те	ests of Interest					
22							
23	Test -2*log	(Likelihood Ratio)	Test	t df p-value			
24							
25	Test 1	22.0468 6		001187			
26	Test 2	3.9664 3		.2651			
27	Test 3	3.9664 3		.2651			
28	Test 4	0.073324 1		0.7866			
29			0.5				
30				There appears to be a			
31				iances among the dose levels			
32	It seems approp	priate to model the	data				
33 34	The <b>p</b> value for	r Tost 2 is graatar t	non 1	A homogonoous variance			
34 35		to be appropriate h		. A homogeneous variance			
35 36	model appears	to be appropriate in	ere				
30 37							
38	The p value for	r Test 2 is greater t	non 1	. The modeled variance appears			
39	to be appropria		1411.1	. The modeled variance appears			
40	to be appropria						
40 41	The n-value for	r Test / is greater t	nan 1	. The model chosen seems			
42	*	lescribe the data	1 <b>a</b> 11 . 1	. The model enosen seems			
43	to adequatery d	eseribe the data					
44							
45	Bench	nark Dose Comput	ation				
46	Denem	inan 2000 compu	acion				
47	Specified effec	t = 0.05					
	r						

```
1
2
3
4
5
6
7
      Risk Type
                         Relative risk
                     =
      Confidence level =
                              0.95
              BMD =
                          64.0102
8
9
             BMDL =
                           40.9875
10
11
12
      BMDL computation failed for one or more point on the BMDL curve.
      The BMDL curve will not be plotted
13
14
15
```

Polynomial Model with 0.95 Confidence Level



10:27 06/15 2007

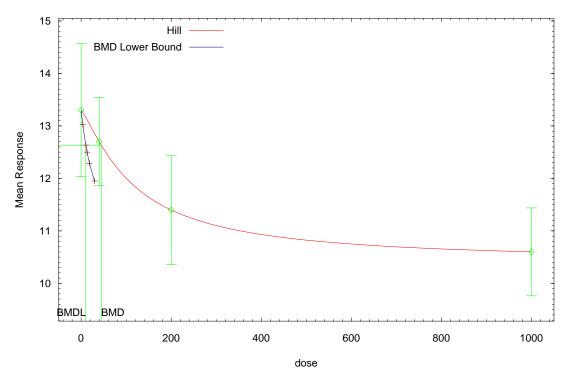
	ill Model. (Version: 2.12; Date: 02/20/2007) put Data File: C:\BMDS\MATERNAL TOXICITY\BODYWEIGHT GAIN GD11-16 FOU					
DOSES.(d) Gnuplot Plotting File: C:\BMDS\ MATERNAL TOXICITY\BODYWEIGHT GAIN GD1						
	OUR DOSES.plt					
	Fri Jun 15 10:38:57 2007					
	DDEL RUN					
The form	of the response function is:					
Y[dose] =	= intercept + $v*dose^n/(k^n + dose^n)$					
	nt variable = MEAN					
	ent variable = Dose					
rho is set						
	rameter restricted to be greater than 1					
A consta	nt variance model is fit					
Total nur	nber of dose groups $= 4$					
	nber of records with missing values = $0$					
	n number of iterations = $250$					
Relative	Function Convergence has been set to: 1e-008					
Paramete	r Convergence has been set to: 1e-008					
	Default Initial Parameter Values					
	alpha = 4.47026					
	rho = 0 Specified					
	intercept = $13.3$					
	v = -2.7					
	n = 1.57036					
	k = 132.308					
Α.	materia Completion Materia of Denometer Detingston					
Asy	mptotic Correlation Matrix of Parameter Estimates					
( **	* The model parameter(s) -rho					
	have been estimated at a boundary point, or have been specified by the user,					
	in a courrent of the work of a courrent of point, or have courrent of the about					

1 2	alpha intercept v n k							
3 4	alpha 1 4.2e-009 2.8e-008 9.6e-009 -3.2e-008							
5 6	intercept 4.2e-009 1 -0.59 -0.38 -0.32							
7 8	v 2.8e-008 -0.59 1 0.77 -0.44							
9 10	n 9.6e-009 -0.38 0.77 1 -0.37							
11 12	k -3.2e-008 -0.32 -0.44 -0.37 1							
13 14								
15 16	Parameter Estimates							
17 18 19 20	95.0% Wald Confidence Interval Variable Estimate Std. Err. Lower Conf. Limit Upper Conf. Limit alpha 4.24102 0.679106 2.90999 5.57204							
20 21 22	$\begin{array}{cccccccccccccccccccccccccccccccccccc$							
23	n 1.2362 1.02244 -0.767751 3.24015							
24 25	k 118.32 98.6055 -74.9435 311.583							
26 27 28	Table of Data and Estimated Values of Interest							
29 30	Dose N Obs Mean Est Mean Obs Std Dev Est Std Dev Scaled Res.							
31 32								
33 34	0 18 13.3 13.3 2.55 2.06 -2.61e-008 40 19 12.7 12.7 1.74 2.06 3.18e-008							
35 36	200         21         11.4         11.4         2.29         2.06         5e-008           1000         20         10.6         10.6         1.79         2.06         -2.63e-007							
37 38	Degrees of freedom for Test A3 vs fitted $\leq 0$							
39 40	Degrees of freedom for rest AS vs fitted <= 0							
41 42 43	Model Descriptions for likelihoods calculated							
44 45 46 47	Model A1: $Yij = Mu(i) + e(ij)$ Var{ $e(ij)$ } = Sigma^2							

	1,3-Butadiene PROPOSED DSD Page 119								
1 2 3	Model A2: $Yij = Mu(i) + e(ij)$ $Var{e(ij)} = Sigma(i)^2$								
4 5	Model A3: $Yij = Mu(i) + e(ij)$ $Var{e(ij)} = Sigma^2$								
6 7 8	Model A3 uses any fixed variance parameters that were specified by the user								
8 9 10	Model R: $Yi = Mu + e(i)$ Var{ $e(i)$ } = Sigma^2								
11 12									
13 14	Likelihoods of Interest								
15	Model Log(likelihood) # Param's AIC								
16	A1 -95.347306 5 200.694612								
17	A2 -93.364104 8 202.728208								
18	A3 -95.347306 5 200.694612								
19	fitted -95.347306 5 200.694612								
20	R -104.387527 2 212.775055								
21									
22									
23	Explanation of Tests								
24									
25	Test 1: Do responses and/or variances differ among Dose levels?								
26	(A2  vs.  R)								
27	Test 2: Are Variances Homogeneous? (A1 vs A2)								
28	Test 3: Are variances adequately modeled? (A2 vs. A3)								
29 20	Test 4: Does the Model for the Mean Fit? (A3 vs. fitted)								
30 31	(Note: When rho=0 the results of Test 3 and Test 2 will be the same.)								
32	Tests of Interest								
33	Tests of Interest								
34	Test -2*log(Likelihood Ratio) Test df p-value								
35	rest 2 log(Elikelinood Rullo) rest dr p vulde								
36	Test 1 22.0468 6 0.001187								
37	Test 2 3.9664 3 0.2651								
38	Test 3 3.9664 3 0.2651								
39	Test 4 2.84217e-013 0 NA								
40									
41	The p-value for Test 1 is less than .05. There appears to be a								
42	difference between response and/or variances among the dose levels								
43	It seems appropriate to model the data								
44									
45	The p-value for Test 2 is greater than .1. A homogeneous variance								
46	model appears to be appropriate here								
47	• • •								

1							
2	The p-value for Test 3 is greater than .1. The modeled variance appears						
3	to be appropriate here						
4							
5	NA - Degrees of freedom for Test 4 are less than or equal to 0. The Chi-Square						
6	test for fit is not valid						
7							
8							
9	Benchmark Dose Computation						
10							
11	Specified effect = $0.05$						
12							
13	Risk Type = Relative risk						
14							
15	Confidence level = $0.95$						
16							
17	BMD = 44.4937						
18							
19	BMDL = 10.3097						
20							

#### Hill Model with 0.95 Confidence Level



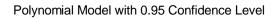
10:38 06/15 2007

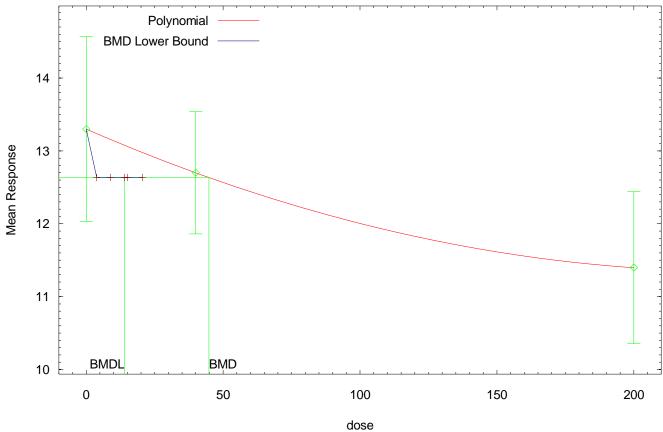
In	lynomial Model. (Version: 2.12; Date: 02/20/2007) put Data File: C:\BMDS\ MATERNAL TOXICITY\BODYWEIGHT GAIN GD11-16					
THREE DOSES.(d) Gnuplot Plotting File: C:\BMDS\ MATERNAL TOXICITY\BODYWEIGHT GAIN GD11						
THREE DC	DSES.plt Fri Jun 15 13:34:32 2007					
BMDS MC	DDEL RUN					
The form	of the response function is:					
Y[dose] =	$beta_0 + beta_1 * dose + beta_2 * dose^2 + \dots$					
Dependen	t variable = MEAN					
	ent variable = Dose					
rho is set						
	he polynomial coefficients are not restricted					
A constan	t variance model is fit					
Total num	where of dose groups $= 3$					
	where of the second se					
	n number of iterations = $250$					
	Function Convergence has been set to: 1e-008					
	Convergence has been set to: 1e-008					
1	Default Initial Denomentary Values					
J	Default Initial Parameter Values alpha = 4.90766					
	rho = 0 Specified					
	beta $0 = 13.3$					
	beta $1 = -0.016375$					
	beta $2 = 3.4375e-005$					
	_					
Asyr	nptotic Correlation Matrix of Parameter Estimates					
	The model parameter(s) -rho					
	ave been estimated at a boundary point, or have been specified by the user,					
a	nd do not appear in the correlation matrix )					

1	alpha beta_0 beta_1 beta_2
2 3	alpha 1 -4e-010 9.1e-011 1.8e-011
4 5	beta 0 -4e-010 1 -0.7 0.63
6 7	beta 1 9.1e-011 -0.7 1 -0.99
8	_
9 10	beta_2 1.8e-011 0.63 -0.99 1
11 12	
13	Parameter Estimates
14 15	95.0% Wald Confidence Interval
16	Variable Estimate Std. Err. Lower Conf. Limit Upper Conf. Limit
17 18	alpha4.653820.8641922.960036.3476beta013.30.50847412.303414.2966
19	beta 1 $-0.016375$ $0.0217309$ $-0.0589669$ $0.0262169$
20	beta_2 3.4375e-005 0.000101174 -0.000163922 0.000232672
21	
22 23	
23 24	Table of Data and Estimated Values of Interest
25	
26	Dose N Obs Mean Est Mean Obs Std Dev Est Std Dev Scaled Res.
27	
28 29	0 18 13.3 13.3 2.55 2.16 2.16e-009
30	40 19 12.7 12.7 1.74 2.16 1.33e-009
31	200 21 11.4 11.4 2.29 2.16 -3.03e-009
32	
33 34	Degrees of freedom for Test A3 vs fitted $\leq 0$
34 35	
36	
37	Model Descriptions for likelihoods calculated
38	
39 40	Model A1: $Yij = Mu(i) + e(ij)$
41	$Var{e(ij)} = Sigma^2$
42	
43	Model A2: $Yij = Mu(i) + e(ij)$
44 45	$Var\{e(ij)\} = Sigma(i)^2$
43 46	Model A3: $Yij = Mu(i) + e(ij)$
47	$Var{e(ij)} = Sigma^2$

	Page 123						
1 2 3	Model A3 u were specifi	•		nce p	arameters that		
4 5 6	Model R: Var{e(i	Yi = Mu + )} = Sigma					
7 8 9	Ι	Likelihoods	of Inter	est			
10	Model	Log(like	lihood)	# P	ıram's AIC		
11	Al	-73.5929			155.185871		
12	A2	-72.2327	25	6	156.465449		
13		-73.5929			155.185871		
14					155.185871		
15	R	-77.34437			158.688744		
16		,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	-	-	100.000711		
17							
18	Ext	planation of	Tests				
19	2.1		10505				
20	Test 1. Do res	ponses and	/or varia	ances	differ among Dose levels	?	
21	(A2 vs. F					-	
22	Test 2: Are V	/	mogene	eous?	(A1 vs A2)		
23					eled? (A2 vs. A3)		
24					Fit? (A3 vs. fitted)		
25					3 and Test 2 will be the s	ame.)	
26	(						
27	Те	ests of Inter	est				
28							
29	Test -2*log	(Likelihood	l Ratio)	Tes	df p-value		
30	C	× ·	,		1		
31	Test 1	10.2233	4	0	.03683		
32	Test 2	2.72042	2	(	0.2566		
33	Test 3	2.72042	2	(	0.2566		
34	Test 4	0	0	Ν	A		
35							
36	The p-value for	Test 1 is le	ess than	.05.	There appears to be a		
37	difference betw	veen respon	se and/o	or vai	iances among the dose lev	vels	
38	It seems approp	priate to mo	del the	data	C		
39							
40	The p-value for	Test 2 is g	reater th	nan .1	. A homogeneous varian	ce	
41	model appears	to be appro	priate he	ere	-		
42			-				
43							
44	The p-value for	Test 3 is g	reater th	nan .1	. The modeled variance a	appears	
45	to be appropriate here						
46							
47	NA - Degrees of	of freedom	for Test	4 are	less than or equal to 0. T	The Chi-Square	

1	test for fit is not valid
2 3	
4	Benchmark Dose Computation
5 6	Specified effect = $0.05$
7	
8 9	Risk Type = Relative risk
10	Confidence level = $0.95$
11 12	BMD = 44.8295
13	DMDI 12.0502
14 15	BMDL = 13.9502





13:34 06/15 2007

2	Appendix 3. Statistical Analyses of Reproductive Endpoints
3	
4 5	Robert L. Sielken Jr., Ph.D., and Ciriaco Valdez Flores, Ph.D., P.E.
	Sielken & Associates Consulting Inc.
6 7	3833 Texas Avenue, Suite 230, Bryan, TX 77802
7	Tel: 979-846-5175; Fax: 979-846-2671;
8 9	Email: SielkenAssoc@aol.com
10	August 6, 2007
11	
12	<b>TCEQ Contract 582-7-81521</b>
13	
14	
15	
16	EPA's 2002 final risk assessment for BD (USEPA. 2002. Health Assessment of 1,3-Butadiene.
17	EPA/600/P-98/001F) derived a reference concentration using the ovarian atrophy in female mice exposed
18	to butadiene via inhalation. This animal study was conducted by the NTP in 1993 (NTP. 1993.
19	Toxicology and carcinogenesis studies of 1,3-butadiene (CAS No. 106-99-0) in B6C3F1 mice (inhalation
20	studies). Research Triangle Park, NC: National Toxicology Program, U.S. Public Health Service, U.S.
21	Department of Health and Human Services. TR 434). EPA used a Weibull time-to-tumor dose-response
22	model to fit the time-to-ovarian atrophy data and excluded the highest dose group because of excessive
23 24	early mortality. The ECs and LECs for ovarian atrophy were calculated at an equivalent human age of 50 years "to reflect only the time before average age at menopause when follicles are no longer present and
24	available for ovulation, because in the mouse studies of ovarian atrophy, the atrophy occurs as a result of
26	follicular failure."
20	Tomeutar failure.
28	In the NTP 1993 critical study, female mice were exposed to 0, 6.25, 20, 62.5, 200, or 625 ppm BD for 6
29	hours/day, 5 days/week for two years (i.e., equivalent to 0, 1.12, 3.57, 11.2, 35.7, and 111.6 ppm BD of
30	continuous exposure – for example, $6.25 \times (5/7) \times (6/24) = 1.12$ ). The air concentration 6.25 ppm was
31	identified as a LOAEL for ovarian atrophy. The final 2002 EPA's risk assessment for BD reports several
32	analyses of these data, including application of a log-logistic model, a quantal Weibull model, and a
33	Weibull time-to-response model.
34	
35	The final Weibull time-to-response model that EPA used is linear in dose with time raised to a power.
36	EPA used TOX RISK version 3.5 (Crump et al., ICF Kaiser International, Ruston, LA) for the model
37	fitting and the estimation of the ECs and LECs. In February 2006, the Olefins Panel of the American
38	Chemistry Council asked the Sapphire Group, Inc. to recalculate EPA's ECs and LECs for ovarian
39	atrophy (Kirman, C. R. and M. L. Gargas. 2006. Benchmark Dose Analyses for Reproductive and
40	Developmental Endpoints for 1,3-Butadiene, Submitted to Olefins Panel, American Chemistry Council,
41	Arlington, VA, February 2006). The Sapphire Group, Inc.'s report included the time-to-response data for
42	ovarian atrophy of the NTP 1993 study, and those data are reproduced here in Attachment A.
43	

1 Sielken & Associates Consulting, Inc. reanalyzed the ovarian atrophy data using the Weibull time-to-2 response model and the data presented in Attachment A. The linear Weibull time-to-response model had 3 the following form: 4 5 Probability of a response (ovarian atrophy) by week T at dose d = $1 - exp \{ \text{ - [} Q_0 + Q_1 \times d \text{ ]} \times T^Z \}.$ 6 7 8 Tables 1 and 2 list the results of the analyses when the highest exposure group is not included in the 9 estimation of the model and when all exposure groups are included, respectively. The results labeled SA# 10 were calculated using Sielken & Associates, Inc.'s GEN.T software package - however, Sielken & 11 Associates verified that the parameter estimates are identical to those estimated with TOX RISK version 12 3.5. The LEC<sub>10</sub> values for the SA# analyses in the table were estimated using 99 simulated bootstrap data 13 sets. The two analyses in addition to EPA's analyses included in Tables 1 and 2 are: 14 15 1) Analysis SA1 parallels the analysis performed by EPA. The small discrepancies between the SA1 16 and EPA analyses may be due to assumptions that EPA may have made and did not describe in 17 their report. 18 19 2) Analysis SA2 uses a modified data set in which all animals that lived beyond age 521 days (74.3) 20 weeks – which is equivalent to 50 years in a 70-year human lifetime --  $(50/70) \times 104$  weeks) were 21 excluded from the parameter estimation. 22 23 In Tables 1 and 2, the range of EC<sub>10</sub> values derived by EPA, SA1, and SA2 analyses is 1.05 to 1.25 ppm 24 whereas the range of the LEC<sub>10</sub> values derived by EPA, SA1, and SA2 analyses is 0.768 to 0.958 ppm. 25 26 Table 1 and 2 also show the results for concentrations corresponding to an extra risk of 0.05. Because the 27 Weibull time-to-tumor model in these analyses is linear in dose, the  $EC_{05}$  and  $LEC_{05}$  values are 28 approximately half the corresponding  $EC_{10}$  and  $LEC_{10}$  values. 29 30 Table 1. Parameters (Q<sub>0</sub>, Q<sub>1</sub>, and Z) for Weibull time-to-response model for ovarian atrophy and corresponding human benchmark 1,3-butadiene exposure concentrations for extra risks of 0.1 and 0.05 at 31 32 50 years of age using different methods of calculation – excluding the highest dose group 33

Analysis	$Q_0$	Q1	Z	$EC_{10}$	LEC <sub>10</sub>	EC <sub>05</sub>	LEC <sub>05</sub>
EPA	4.86×10 <sup>-6</sup>	7.06×10 <sup>-6</sup>	2.21	1.05	0.878	n/a	n/a
SA1	6.96×10 <sup>-6</sup>	8.62×10 <sup>-6</sup>	2.15	1.15	0.881	0.560	0.429
SA2	6.76×10 <sup>-23</sup>	6.90×10 <sup>-5</sup>	1.66	1.18	0.768	0.573	0.374

- **Table 2.** Parameters for Weibull time-to-response model for ovarian atrophy and corresponding human
- benchmark 1,3-butadiene exposure concentrations for extra risks of 0.1 and 0.05 at 50 years of age using
- 37 different methods of calculation **including** the highest dose group
- 38

Analysis	$Q_0$	Q1	Ζ	$EC_{10}$	LEC <sub>10</sub>	$EC_{05}$	LEC <sub>05</sub>
EPA	9.01×10 <sup>-6</sup>	1.32×10 <sup>-6</sup>	2.58	1.13	0.958	n/a	n/a
SA1	1.68×10 <sup>-6</sup>	2.04×10 <sup>-6</sup>	2.47	1.25	0.949	0.607	0.462
SA2	3.61×10 <sup>-25</sup>	1.95×10 <sup>-6</sup>	2.49	1.17	0.812	0.569	0.396

1

2 The estimated values of  $EC_{10}$  and  $LEC_{10}$  are close to the lowest experimental dose (1.12 ppm) while the 3 values of EC<sub>05</sub> and LEC<sub>05</sub> are approximately half way between the lowest experimental dose and zero. 4 The values of  $EC_{05}$  and  $LEC_{05}$  can be used if the dose-response relationship below the lowest 5 experimental dose is believed to be the linear Weibull time-to-response model fit to the data. The 6 assumption of linearity below the lowest experimental dose is usually conservative and, therefore, health 7 protective. However, the motivation behind the benchmark dose methodology is to identify the point of 8 departure (EC or LEC) to be within the range of the experimental data (the range of the non-zero doses in 9 the experimental data) and to be a dose whose risk can be reasonably reliably estimated without undue 10 sensitivity to the dose-response model selected or the model estimation. Here, the EC<sub>05</sub> and LEC<sub>05</sub> in the SA1 and SA2 analyses are below the range of the experimental data and, hence, introduce an additional 11 12 element of uncertainty into the point of departure. 13

14 The EPA and SA1 analyses include ovarian atrophy responses beyond the equivalent of age 50 years in 15 humans. These older-age responses in mice may not be relevant to humans and may inappropriately 16 impact the fitted dose-response model used to estimate the risk at age 50. SA2 eliminates all animals that 17 lived beyond the equivalent of age 50. However, it is known that some of these animals did not have an 18 observed response (ovarian atrophy) and this information is ignored/lost and not incorporated into the 19 dose-response modeling as it should be. The fitted models for all the mice (analyses SA1) are very 20 similar to the fitted models for only mice that died on or before week 74.3 (analyses SA2). This suggests 21 that the older-age animals in the SA1 analyses are not distorting those analyses. Therefore, the results for 22 analyses SA1 are preferable to the SA2 analyses because the SA1 analyses include more data (i.e., mice 23 that lived past 74.3 weeks) and the inclusion of mice older than 74.3 weeks does not distort the fit of the 24 model. In other words, the models fit to either all the mice (analyses SA1) or only to mice that died on or 25 before week 74.3 are (analyses SA2) very similar but the confidence limits for analyses SA1 are more

- 26 reliable because they are based on more animals.
- 27

28 The ovarian atrophy data were analyzed excluding the highest dose group (Table 1) and also including all

the data (Table 2). The analyses that exclude the high dose were performed to parallel those analyses
used by EPA. Traditionally, EPA drops the highest dose group when the model does not fit the data well

31 due to some biological phenomenon or when quantal data are fit with a quantal model and there is high

32 mortality in the highest dose group. The ovarian atrophy data, however, were modeled with a time-to-

response model (i.e., a model that accounts for the time of death) as opposed to a quantal model which do not account for time of death. Furthermore, the model fit to the data that excluded the highest dose group

was not better than the model fit to the data that included the highest dose group. Figure 1 shows the fit of

analysis SA1 to the lower four dose groups and the control group while Figure 2 shows the fit of analysis
 SA1 to all dose groups and the control group.

38

In summary, the SA1 analysis in Table 2 that includes all the exposure groups and all animals in each exposure group is the most statistically sound analysis of the ovarian atrophy study because: 1) the model

fit using all animals is similar to the model fit using only animals that died on or before 74.3 weeks of

41 In using an annuals is similar to the model in using only annuals that died on of before 74.5 weeks of 42 age, 2) the model fit using all dose groups is similar to the model fit to only the four lowest dose groups,

and 3) using all the data results in more reliable maximum likelihood estimates and corresponding

44 confidence limits.

1 Figure 1. Observed versus multistage-Weibull model predicted proportions of mice with ovarian atrophy

- 2 when only the four lowest dose groups and the control group are used to fit the model

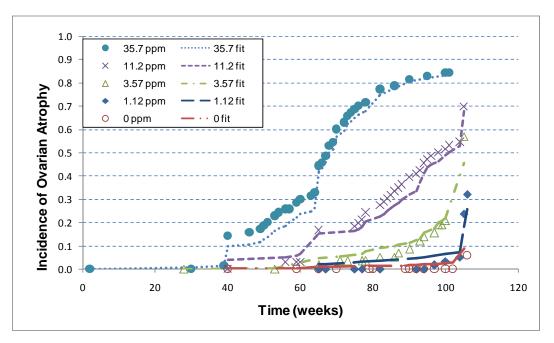
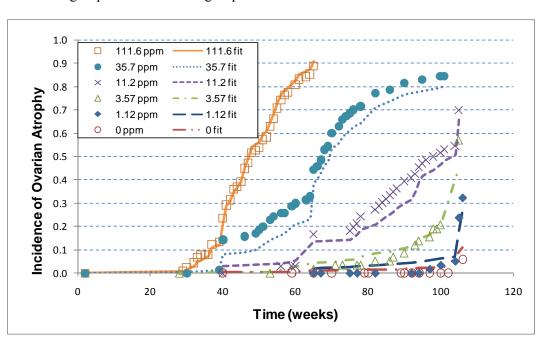


Figure 2. Observed versus multistage-Weibull model predicted proportions of mice with ovarian atrophy when all five dose groups and the control group are used to fit the model.



### Attachment A

Time-to-response for ovarian atrophy as reported by the Sapphire Group, Inc. of the NTP 1993 study (NTP. 1993.

Toxicology and carcinogenesis studies of 1,3-butadiene (CAS No. 106-99-0) in B6C3F1 mice (inhalation studies). Research Triangle Park, NC: National Toxicology Program, U.S. Public Health Service, U.S. Department of Health and Human Services. TR 434).

Concentration (ppm)	Responders	Non-Responders	n	Day	Week
0	0	10	10	280	40
0	0	1	1	413	59
0	0	10	10	455	65
0	0	1	1	490	70
0	0	1	1	553	79
0	0	1	1	560	80
0	0	1	1	623	89
0	0	1	1	630	90
0	0	1	1	644	92
0	0	1	1	658	94
0	0	1	1	679	97
0	0	1	1	700	100
0	0	3	3	714	102
0	4	32	36	742	106
6.25	0	10	10	455	65
6.25	0	1	1	469	67
6.25	0	2	2	525	75
6.25	0	1	1	539	77
6.25	0	1	1	574	82
6.25	0	3	3	644	92
6.25	0	1	1	658	94
6.25	1	0	1	679	97
6.25	1	1	2	700	100
6.25	1	0	1	728	104
6.25	11	10	21	735	105
6.25	5	10	15	742	106
20	0	1	1	196	28
20	0	1	1	371	53
20	1	9	10	455	65
20	1	0	1	497	71
20	0	1	1	511	73
20	0	1	1	539	77

20	0	2	2	546	78	
20	1	1	2	574	82	
20	0	1	1	602	86	
20	1	0	1	609	87	
20	1	0	1	630	90	
20	2	0	2	651	93	
20	1	2	3	658	94	
20	1	1	2	679	97	
20	2	1	3	686	98	
20	0	1	1	693	99	
20	1	0	1	700	100	
20	21	3	24	735	105	
62.5	0	10	10	280	40	
62.5	2	0	2	392	56	
62.5	0	1	1	413	59	
62.5	0	1	1	420	60	
62.5	9	1	10	455	65	
62.5	1	0	1	525	75	
62.5	1	0	1	532	76	
62.5	1	0	1	539	77	
62.5	2	0	2	546	78	
62.5	2	0	2	574	82	
62.5	1	0	1	581	83	
62.5	1	0	1	588	84	
62.5	1	0	1	595	85	
62.5	1	0	1	602	86	
62.5	1	0	1	609	87	
62.5	1	0	1	616	88	
62.5	2	0	2	630	90	
62.5	1	0	1	644	92	
62.5	1	2	3	651	93	
62.5	2	1	3	658	94	
62.5	1	1	2	665	95	
62.5	1	0	1	672	96	
62.5	1	0	1	686	98	
62.5	1	1	2	700	100	
62.5	1	0	1	707	101	
62.5	1	1	2	728	104	
62.5	10	1	11	735	105	

200	0	1	1	14	2
200	0	1	1	210	30
200	1	0	1	2733*	390.4286
200	9	1	10	280	40
200	1	0	1	322	46
200	1	0	1	343	49
200	1	0	1	350	50
200	1	0	1	357	51
200	2	0	2	371	53
200	1	0	1	378	54
200	1	0	1	392	56
200	0	1	1	399	57
200	2	0	2	413	59
200	1	0	1	420	60
200	1	0	1	441	63
200	1	0	1	448	64
200	8	4	12	455	65
200	1	0	1	462	66
200	2	0	2	469	67
200	3	0	3	476	68
200	1	0	1	483	69
200	4	0	4	490	70
200	2	0	2	504	72
200	2	0	2	511	73
200	1	0	1	518	74
200	1	0	1	525	75
200	1	0	1	532	76
200	1	0	1	546	78
200	4	1	5	574	82
200	1	1	2	602	86
200	2	0	2	630	90
200	1	0	1	665	95
200	1	0	1	700	100
200	0	1	1	707	101
625	0	1	1	14	2
625 625	1	0	1	203	29
625 625	1	0	1	210	30
625	2	0	2	210	32
	-	v	4		~ -

625	1	0	1	238	34
625	0	1	1	245	35
625	0	1	1	252	36
625	4	0	4	259	37
625	1	0	1	273	39
625	9	1	10	280	40
625	5	2	7	287	41
625	2	0	2	294	42
625	4	0	4	301	43
625	1	1	2	308	44
625	2	0	2	315	45
625	5	1	6	322	46
625	2	2	4	329	47
625	4	0	4	336	48
625	1	0	1	343	49
625	1	0	1	350	50
625	3	0	3	357	51
625	1	0	1	364	52
625	4	0	4	371	53
625	3	0	3	378	54
625	4	0	4	385	55
625	3	0	3	392	56
625	2	0	2	399	57
625	1	0	1	406	58
625	3	0	3	420	60
625	2	0	2	427	61
625	1	0	1	441	63
625	1	0	1	448	64
625	3	0	3	455	65

## Appendix 4. Cox Proportional Hazards Models Not Included in Cheng et al. (2007)

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16 Cheng et al. presented several analyses with the objective of showing different alternatives they thought 17 could be relevant. For example, they restricted the analyses to include only cumulative ppm-years, 18 average intensity or lagged cumulative ppm-years as the relevant doses. There is no evidence that any of 19 these measures of dose is the relevant dose. They also fit models that adjusted for race, year of birth, race, 20 years since hire, plant and number of high intensity tasks (HITs) and exposures to DMDTC. Cheng et al. 21 did not give any biological reasons to include or exclude from the model. Ideally, the final model should 22 adjust for effects that are biologically relevant to the outcome of study. However, there is not enough 23 scientific knowledge to indicate what, if any, covariate effects should be included in a model of leukemia 24 mortality with cumulative exposure to butadiene. The research closest to shedding some light on which

covariates to include in the model is that published by Albertini et al. (2007), which seems to indicate that leukemia does not occur at low exposure to butadiene.

26 leu 27

Although the decision of whether or not to adjust for a confounder should ideally be based on biological knowledge, Sielken et al. (2007) adjustment for confounders was determined using a statistically-based approach. The use of statistical methodology instead of biological arguments serves for the purpose of

31 corroborating new biological evidence about possible confounders – specifically the role of the number of

high intensity tasks in leukemia rate ratios. That is, the inclusion of the number of HITs as a covariate,

although based on statistical arguments, was consistent with the biological findings of Albertini et al.

34 (2007). In other words, not only was the number of HITs a plausible explanation of the increase in the

number of leukemia deaths from a biological and mechanistic standpoint but also the statistical analysis of the data reached the same conclusion. Other attributes to see in model selection are issues like:

36 of the data reached the same conclusion. Other attributes to see in model selection are issues like: 37 consistency with biological expectations (i.e., the model should make biological sense), model parsimony

(i.e., include as few variables as necessary to explain the relationship when there is no sufficient

biological knowledge to justify the inclusion or exclusion of a variable), etc.

- 41 Cheng et al. (2007) presented a model that adjusts for age and the number of HITs (BD peaks). That is,  $\beta$
- 42 =  $2.5 \times 10-4$ , p = 0.03 presented in Section 3.5 of the Cheng et al. (2007) paper. This results in a S.E. of
- 43  $1.2 \times 10-4$ . This model is close to the Poisson regression model in the Sielken et al. (2007) paper with the

1 exceptions that: 1) Sielken et al. adjusted for the number of HITs using a nonparametric relation based on

2 quintiles whereas Cheng et al. adjusted for the number of HITs using a parametric linear relationship, 2)

3 Cheng et al. models assume an exponential relationship between rate ratios and cumulative BD ppm-years

4 whereas Sielken et al. uses a linear relationship, 3) Cheng et al. use Cox proportional hazards model and

5 Sielken et al. use Poisson regression model, and 4) Cheng et al. use continuous cumulative BD ppm-years

6 and Sielken et al. uses BD ppm-years mean-scored deciles.

7

Model	Covariates	Parameter	<sup>•</sup> Estimate	URF <sup>a</sup> (ppm	-1)
				Air Concen an excess ri 100,000 (pp	sk of 1 in
		β (S.E.)	95% UCL	URF (MLE)	URF(95% UCL); 95% LCL on Conc.
Cox regression Cheng et al. (2007)	Age	2.5E-04 (1.2E-04)	4.474E-04	1.284E-04	2.298E-04
ppm-years continuous <sup>b</sup> , # of HITS continuous <sup>c</sup>	number of HITs > 100 ppm			77.88	43.52
Cox regression ppm-years continuous <sup>b</sup> ,	Age	2.0E-04 (1.3E-04)	4.138E-04	1.027E-04	2.125E-04
# of HITS categorical <sup>d</sup>	number of HITs > 100 ppm			97.35	47.05
Cox regression ppm-years mean-scored deciles <sup>e</sup> ,	Age	2.8E-04 (2.4E-04)	6.748E-04	1.438E-04	3.466E-04
# of HITS categorical <sup>d</sup>	number of HITs > 100 ppm			69.53	28.85
Poisson regression (Sielken et al. (2007)	Age	1.89E-04 (3.6E-04)	7.812E-04	8.083E-05	3.314E-04
ppm-years mean-scored deciles <sup>e</sup> , # of HITS categorical <sup>d</sup>	number of HITs > 100 ppm			123.7	29.93

8 a URF(MLE) = 0.001 / EC<sub>001</sub> and URF(95% UCL) = 0.001 / LEC<sub>001</sub>

9 <sup>b</sup>ppm-years is included as a continuous variable (untransformed) in a parametric model of the effect of

10 ppm-years

11 <sup>c</sup> number of HITS > 100 ppm is included as a continuous variable (untransformed) in a parametric model

12 of the effect of the number of HITS > 100 ppm

<sup>d</sup> number of HITS > 100 ppm is included as a categorical variable (based on quintiles) in a nonparametric model of the effect of the number of HITS > 100 ppm

<sup>15</sup> <sup>e</sup> ppm-years is included as a categorical variable (based on mean-scored deciles, untransformed) in a

16 parametric model of the effect of ppm-years

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1 Despite all these differences, the models are close and converge to very similar results if some of the discrepancies are resolved. For example, if the Cox proportional hazards exponential model presented by 2 3 Cheng et al. were non-parametrically adjusted for BD peaks, then the estimate of the coefficient for 4 cumulative BD ppm-years would be  $\beta = 0.00020$  (S.E.=0.00013), which is close to the parameter 5 estimates reported in Sielken et al. (i.e.,  $\beta = 0.000189$ , S.E.=0.00036) for the Poisson linear model. If, in addition to adjusting for the number of HITs nonparametrically, the Cox proportional hazards exponential 6 7 model used BD ppm-years mean-scored deciles instead of continuous exposures, then the coefficient for 8 cumulative BD ppm-years would be  $\beta = 0.00028$  (S.E.=0.00024). This last model differs from Sielken et 9 al, model only in that Sielken et al. used a Poisson regression model and a linear relationship as opposed 10 to the Cox proportional hazards model and a log-linear relationship. The following table summarizes the results of the Cox proportional hazards model and the Sielken et al. Poisson regression model when 11 12 adjusting for the number of HITs.

13

14 In the above discussion, a parametric model is a model that assumes a specified functional form (e.g.,

15 linear or log-linear), and a nonparametric model is a model that does not assume a specified functional

16 form. This is analogous to the difference between regression which assumes a specified functional form

17 (e.g., linear or polynomial) and hence is parametric and analysis of variance (ANOVA or AOV) which is

18 nonparametric. Continuing with the analogy, if a treatment can be characterized by a number (e.g., 19 concentration or amount), then in a regression analysis (say, a linear regression) the magnitudes of the

20 different treatment values are important and a treatment with twice the magnitude has twice the effect. On

21 the other hand, in an analysis of variance the different treatments are dealt with nonparametrically (say, as

22 treatments A, B, C, etc.) and the magnitudes (numerical values) are ignored. Therefore, in an analysis of variance there is no functional relationship specified between the effects of the different treatments.

23 24

25 If a variable is said to be treated continuously, then each individual value of that variable is used – the values are not grouped and no representative values for the groups are used. On the other hand, if a 26 27 variable is treated categorically, then the individual values of that variable are grouped and representative 28 values for the groups replace the individual values in the analysis. Cumulative butadiene ppm-years and 29 cumulative number of HITS > 100 ppm can both be treated either as continuous or categorical variables.

30 Since the categorical (group) values for these variables are numerical, a categorical variable could be

31

included in both parametric and nonparametric models. 32

33 In the table above, both the Cox and Poisson regressions assume a parametric model for the effect of 34 cumulative butadiene ppm-years. The model for the effect of ppm-years is log-linear in Cox regression 35 and is linear in Poisson regression. In Cox regression, ppm-years is treated as a continuous variable in the 36 first two models and treated as a categorical variable in the third model. In the Poisson regression, ppm-37 years is treated as a categorical variable.

38

39 In the first model in the table above, the cumulative number of HITS > 100 ppm is treated as a continuous 40 variable and treated parametrically. In the other three models, the cumulative number of HITS > 100 ppm

- 41 is treated as a categorical variable and treated nonparametrically.
- 42 43

44 Albertini, R., Sram, R. J., Vacek, P. M., Lynch, J., Rossner, P., Nicklas, J. A., McDonald, J. D., Boysen,

45 G., Georgieva, N., and Swenberg, J. A. (2007). Molecular epidemiological studies in 1,3-butadiene

exposed Czech workers: Female-male comparisons. Chemico-Biological Interactions, Volume 166, Issues 46

47 1-3, 20 March 2007, Pages 63-77.

# 1 Appendix 5. Leukemia Mortality Rates and Survival Rates

US Total Poj 2000-20		Texas Statewide 1999-2003		
Total Leukemia Mortality Rates per 100,000		Total Leukemia Mortality Rates per 100,000 <sup>2</sup>		
	Rate		Rate	
00 years	0.7	00 years	0.9	
01-04 years	0.9	01-04 years	0.9	
05-09 years	0.7	05-09 years	0.6	
10-14 years	0.8	10-14 years	0.9	
15-19 years	1.1	15-19 years	1.3	
20-24 years	1.2	20-24 years	1.5	
25-29 years	1.1	25-29 years	1.1	
30-34 years	1.3	30-34 years	1.4	
35-39 years	1.6	35-39 years	1.5	
40-44 years	2.0	40-44 years	1.8	
45-49 years	2.9	45-49 years	3.4	
50-54 years	4.4	50-54 years	4.2	
55-59 years	7.5	55-59 years	8.4	
60-64 years	12.9	60-64 years	13.2	
65-69 years	20.8	65-69 years	21.3	
70-74 years	33.0	70-74 years	31.8	
75-79 years	47.0	75-79 years	43.4	
80-84 years	63.2	80-84 years	65.5	
85+ years	81.5	85+ years	81.3	

 Table XIII-8, Seer Cancer Statistics Review 2000-2003Surveillance, Epidemiology, and End Results database (SEER 2006))

<sup>2</sup> Texas-specific mortality rates for 1999-2003 for all leukemia and Texas-specific survival rates for 2003 were kindly provided by the Texas Department of State Health Services, Cancer Epidemiology and Surveillance Branch, Texas Cancer Registry.

	2000 US All <sup>1</sup>	Total Texas P	Copulation 2003 <sup>2</sup>
Age	Survival	Life Tables	
0	1	0	1
1	0.99307	1	0.99342
5	0.99177	5	0.99191
10	0.99095	10	0.99105
15	0.98992	15	0.99005
20	0.98654	20	0.98659
25	0.98181	25	0.9818
30	0.97696	30	0.9772
35	0.97132	35	0.97192
40	0.96349	40	0.9641
45	0.9521	45	0.95248
50	0.93522	50	0.93546
55	0.91113	55	0.91092
60	0.87498	60	0.87584
65	0.82131	65	0.82385
70	0.74561	70	0.75079
75	0.64244	75+	0.65073
80	0.51037		
85	0.34959		

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US survival rates for 2000 (Arias 2002) Texas-specific survival rates for 2003 were kindly provided by the Texas Department of State Health Services, Cancer Epidemiology and Surveillance Branch, Texas Cancer Registry.

1 2	Appendix 6. Calculating Excess Risk with Age-Dependent Adjustment Factors
3 4 5 6 7 8	Robert L. Sielken Jr., Ph.D., and Ciriaco Valdez Flores, Ph.D., P.E. Sielken & Associates Consulting Inc. 3833 Texas Avenue, Suite 230, Bryan, TX 77802 Tel: 979-846-5175; Fax: 979-846-2671; Email: SielkenAssoc@aol.com
9 10	March 12, 2007
11 12 13	<b>TCEQ Contract 582-7-81521</b>
14 15 16	1. Background:
17 18 19 20 21	When calculating an excess risk, a general guiding principle is that the dose-response model, model parameter, dose metric, response, and population used in the excess risk calculation using the BEIR IV approach (NRC 1988) should be the same as the dose-response model, model parameter, dose metric, response, and population used in the dose-response modeling of the epidemiological study data.
21 22 23 24 25 26 27	If the population in the dose-response modeling has specific characteristics (e.g., gender, race, and geographic region), then the inference (the calculated excess risk) applies directly to that specified population. It only applies to a more general population under the assumption that the estimated model parameter and dose-response model apply to that population – this is an assumption, not a guarantee, and not something that is necessarily proven or implied by the study data.
28 29 20	2. Age-Dependent Adjustment Factor (ADAF): General
30 31 32 33 34 35 36	An ADAF is intended to be used when the epidemiological study data do not include exposures at an early age (generally before age 16). According to U.S. EPA (Barton, H., et al. Supplemental Guidance for Assessing Susceptibility from Early-Life Exposure to Carcinogens, EPA/360/R-03/003F, March 2005, Washington, D.C.), an ADAF is intended to address the "potential for increased susceptibility to cancer from early-life exposure, relative to comparable exposure later in life".
37 38	ADAFs are age-specific adjustments to the susceptibility (slope) in the dose-response model and are not adjustments to the dose metric itself.
<ol> <li>39</li> <li>40</li> <li>41</li> <li>42</li> <li>43</li> <li>44</li> <li>45</li> </ol>	If the epidemiological study data do not include exposures at an early age (e.g., before age 16) such as would generally be the case for occupational epidemiological studies, then ADAFs that are only different than 1 before age 16 do not impact the dose-response modeling and cannot reflect different susceptibilities at early ages relative to later ages. Thus, the fitted model parameter cannot directly reflect different susceptibilities at early ages relative to later ages. Therefore, it is reasonable to do an excess risk calculation using the dose metric and model parameter estimated from the epidemiological data – without

explicitly recomputing the model parameter based on the susceptibilities implied by the ADAFs – since
 the model parameter won't change anyway.

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4 Although it is somewhat of an aside, it is important to note the treatment of "background doses" needs to 5 be the same in the dose-response modeling and the excess risk calculation. Specifically, if for example, 6 there is a general background exposure of X ppm per year to the chemical of interest, then the excess risk 7 calculation should treat that X ppm per year in the same way as the dose-response model fitting. If the 8 dose-response modeling was done with the dose metric including that X ppm per year, then the excess 9 risk calculation should be done with the dose metric including that X ppm per year. If the dose-response 10 modeling was done with the dose metric excluding that X ppm per year, then the excess risk calculation should be done with the dose metric excluding that X ppm per year. It would be invalid for the dose-11 12 response modeling to be done with the dose metric excluding that X ppm per year, and the excess risk 13 calculation to be done with the dose metric including that X ppm per year – or vice versa.

# 15 3. Age-Dependent Adjustment Factor (ADAF): EPA Guidelines 16

EPA guidelines call for the default use of ADAFs be considered only when the chemicals mode of action
is mutagenic.

EPA guidelines (pages 32-34) also include the following text:

The adjustments described below reflect the potential for early-life exposure to make a greater contribution to cancers appearing later in life. The 10-fold adjustment represents an approximation of the weighted geometric mean tumor incidence ratio from juvenile or adult exposures in the repeated dosing studies (see Table 8). This adjustment is applied for the first 2 years of life, when toxicokinetic and toxicodynamic differences between children and adults are greatest (Ginsberg et al., 2002; Renwick, 1998). Toxicokinetic differences from adults, which are greatest at birth, resolve by approximately 6 months to 1 year, while higher growth rates extend for longer periods. The 3-fold adjustment represents an intermediate level of adjustment that is applied after 2 years of age through <16 years of age. This upper age limit represents middle adolescence following the period of rapid developmental changes in puberty and the conclusion of growth in body height in NHANES data (Hattis et al., 2005). Efforts to map the approximate start of mouse and rat bioassays (i.e., 60 days) to equivalent ages in humans ranged from 10.6 to 15.1 years (Hattis et al., 2005). Data are not available to calculate a specific dose response adjustment factor for the 2 to <16-year age range, so EPA selected the 3-fold adjustment because it reflects a midpoint, i.e., approximately half the difference between 1 and 10 on a logarithmic scale  $(10^{1/2})$ , between the 10-fold adjustment for the first two years of life and no adjustment (i.e., *1-fold) for adult exposure.* ...

...the Supplemental Guidance emphasizes that chemical-specific data should be used in preference to these default adjustment factors whenever such data are available.

The following adjustments represent a practical approach that reflects the results of the preceding analysis, which concluded that cancer risks generally are higher from early-life exposure than from similar exposure durations later in life:

3 4 5 6 7 8	<ul> <li>For exposures between 2 and &lt;16 years of age (i.e., spanning a 14-year time interval from a child's second birthday up until their sixteenth birthday), a 3-fold adjustment</li> <li>For exposures after turning 16 years of age, no adjustment.</li> <li>This Supplemental Guidance focuses on carcinogens with a mutagenic mode of action.</li> <li>When data, including well established mode of action data, are available that allow specific evaluation of lifestage differences in toxicokinetics or toxicodynamics that would lead to lesser or greater susceptibility from early-life exposures to carcinogens, then those data should be used,</li> </ul>
6 7 8	<ul> <li>For exposures after turning 16 years of age, no adjustment.</li> <li>This Supplemental Guidance focuses on carcinogens with a mutagenic mode of action.</li> <li>When data, including well established mode of action data, are available that allow specific evaluation of lifestage differences in toxicokinetics or toxicodynamics that would lead to lesser or</li> </ul>
7 8	This Supplemental Guidance focuses on carcinogens with a mutagenic mode of action. When data, including well established mode of action data, are available that allow specific evaluation of lifestage differences in toxicokinetics or toxicodynamics that would lead to lesser or
	When data, including well established mode of action data, are available that allow specific evaluation of lifestage differences in toxicokinetics or toxicodynamics that would lead to lesser or
9	evaluation of lifestage differences in toxicokinetics or toxicodynamics that would lead to lesser or
10	evaluation of lifestage differences in toxicokinetics or toxicodynamics that would lead to lesser or
11 12	
12	
14	as generally discussed in EPA's cancer guidelines (U.S. EPA, 2005), in preference to the default
15	procedures described in this Supplemental Guidance.
16	
17	The 10-fold and 3-fold adjustments in slope factor are to be combined with age-specific
18	exposure estimates when estimating cancer risks from early life exposure to carcinogens that act
19	through a mutagenic mode of action. It is important to emphasize that these adjustments are
20	combined with corresponding age-specific estimates of exposure to assess cancer risk. For
21 22	example, for a 70-year lifetime, where there are data showing negligible exposure to children, the estimated cancer risk from childhood exposure would be also negligible and the lifetime cancer
22	risk would be reduced to that resulting from the relevant number of years of adult exposure (in
24	the absence of specific information, 55 years). Where there are data (measured or modeled) for
25	childhood exposures, the age-group specific exposure values are used along with the
26	corresponding adjustments to the slope factor. Where there are no relevant data or models for
27	childhood exposures and only lifetime average exposure data are available, the lifetime exposure
28	data are used with the <b>adjustments to the slope factor for each age segment</b> .
	(emphasis added)
30	
	There are several important points/clarifications in this last paragraph. The first is that the ADAF
	s an adjustment to the slope factor (as opposed to an adjustment to the dose metric). The second is that the ADAF is to be applied on an age-specific basis. That is, the ADAFs are applied to each year
	in a life and summed to get the lifetime risk, as opposed to calculating a lifetime excess risk without
	ADAFs and then multiplying this calculated value by a constant ADAF.
36	
	This second point is reinforced in the examples provided by EPA (Sections 6.1 and 6.2, pages 36 to 41).
38 .	Although EPA's examples do not explicitly refer to cumulative doses, they do refer to age-dependent
	loses and cumulative doses are age-dependent doses.
40	
42 1	4. Age-Dependent Adjustment Factor (ADAF): Recent Implementations by EPA and Others when the Dose Metric is Cumulative Exposure are Inconsistent with EPA Guidelines
43	In resent risk assessments (a.g. for athylans suide) when the days matrice is sumulating any part
	In recent risk assessments (e.g., for ethylene oxide) when the dose metric is cumulative exposure, EPA has implemented the ADAF by calculating the excess risk by first calculating the excess risk without any
46	ADAFs and then multiplying this excess risk by a weighted average of the age-specific ADAFs over the 'lifetime'' (i.e., the period over which the excess risk is calculated, 70, 78, or 85 years). Similarly, the

1 EPA has implemented the ADAF by calculating the point of departure (POD) by first calculating the 2 POD without an ADAF and then dividing this POD by a weighted average of the age-specific ADAFs 3 over the "lifetime". 4 5 In these recent risk assessments (e.g., for ethylene oxide) when the dose metric is cumulative exposure, 6 EPA's method of incorporating an ADAF has been inconsistent with EPA's Guidelines, has not properly 7 incorporated age-dependence, and is mathematically incorrect. There is no good scientific/mathematical 8 reason for incorporating an ADAF in the manner in which EPA has attempted to do it. Others (including 9 the first draft assessment of butadiene by NC SAB) have made the same mistake. 10 11 For inhalation exposure of a chemical with a mutagenic mode of action, EPA guidelines [Barton, H., et al. 12 Supplemental Guidance for Assessing Susceptibility from Early-Life Exposure to Carcinogens. 13 EPA/630/R-03/003F, March 2005] suggest that the increased risk caused by early-life exposure be 14 determined through the use of three Age-Dependent Adjustment Factors (ADAFs): 15 16 (1) ADAF(age) = 10 for exposure before 2 years of age 17 (2) ADAF(age) = 3 for exposure between ages 2 and < 16 years of age 18 (3) ADAF(age) = 1 for exposure after turning 16 years of age. 19 20 Furthermore, assuming that exposure to a mutagenic chemical via inhalation is constant over a 70-21 year lifetime, EPA's proposed overall adjustment factor (ADAF) for early-life exposure is: 22 23  $ADAF = \sum_{i} (ADAF(i) \times Age Interval) / 70 \text{ years}$ 24 25  $= [(10 \times 2 \text{ years}) + (3 \times 13 \text{ years}) + (1 \times 55 \text{ years})]/70 \text{ years} = 1.63.$ 26 27 For a 78-year lifespan, the corresponding ADAF would be 1.56 because 28 29  $[(10 \times 2 \text{ years}) + (3 \times 13 \text{ years}) + (1 \times 63 \text{ years})]/78 \text{ years} = 1.56.$ 30 31 For an 85-year lifespan, the corresponding ADAF would be 1.52 because 32 33  $[(10 \times 2 \text{ years}) + (3 \times 13 \text{ years}) + (1 \times 70 \text{ years})]/85 \text{ years} = 1.52.$ 34 35 Then, the point of departure (POD) is "adjusted for early-life exposure" by dividing the unadjusted POD 36 by ADAF, according to the EPA proposed method of adjustment. 37 38 This adjustment is consistent with EPA Guidelines provided that exposure to a mutagenic chemical via 39 inhalation is constant over the lifetime and the dose metric is the exposure concentration (as 40 opposed to cumulative exposure). Here, "lifetime" should be interpreted as the period over which the excess risk is calculated (e.g., 70, 78, or 85 years). 41 42 43 Specifically, in Example 2 (part a) in Section 6.1 of EPA Guidelines, the calculation of excess risk for 70 44 years exposure to a constant dose (0.0001 mg/kg-d) when the dose metric is exposure concentration (as 45 opposed to cumulative exposure) is as follows:

1 2	a. To calculate lifetime risk for a population with average life expectancy of 70 years, sum the risk associated with each of the three relevant time periods:
3	• Risk during the first 2 years of life (where the ADAF = 10);
4	• Risk for ages 2 through $< 16$ (ADAF = 3); and
5	• Risk for ages 16 until 70 years $(ADAF = 1)$ .
6 7 8	Thus, risk equals the sum of:
9	• Risk for birth through $< 2$ yr =
10 11	$(2 \text{ per } mg/kg-d) \ge 10 (ADAF) \ge (0.0001  mg/kg-d) \ge 2yr/70yr = 0.6 \ge 10^{-4}$
12	• Risk for ages 2 through $< 16 =$
13 14	$(2 \text{ per } mg/kg-d) \ge 3 (ADAF) \ge (0.0001  mg/kg-d) \ge (13yr/70yr) = 1.1 \ge 10^{-4}$
15	• Risk for ages 16 until 70 =
16	$(2 \text{ per } mg/kg-d) \ge 1 (ADAF) \ge (0.0001  mg/kg-d) \ge (55yr/70yr) = 1.6 \ge 10-4$
17 18 19	$Risk = 0.6 \times 10^{-4} + 1.1 \times 10^{-4} + 1.6 \times 10^{-4} = 3.3 \times 10^{-4}$
20 21 22 23	Here, where the exposure is to a constant dose (0.0001 mg/kg-d) and the dose metric is exposure concentration (as opposed to cumulative exposure), the risk could be calculated as "risk without ADAFs" times a weighted average adjustment factor here,
23 24 25	$ADAF = \sum_{i} (ADAF(i) \times Age Interval) / 70 \text{ years}$
26 27	= $[(10 \times 2 \text{ years}) + (3 \times 13 \text{ years}) + (1 \times 55 \text{ years})] / 70 \text{ years} = 1.63.$
28 29 30	This works only when the age-specific risk per year (before ADAF) is a constant for all ages. Here, the lifetime risk (before ADAF) is
31 32	(2 per mg/kg-d) x (0.0001 mg/kg-d).
33 34	Here,
35 36	(2 per mg/kg-d) x (0.0001 mg/kg-d)
37 38 39	is a common term in each of the risks being summed, so it can be factored out and the calculation represented as
40	(2 per mg/kg-d) x (0.0001 mg/kg-d)
41 42	x [ $(10 \times 2 \text{ years}) + (3 \times 13 \text{ years}) + (1 \times 55 \text{ years})$ ] / 70 years
43 44	$= (2 \text{ per mg/kg-d}) \times (0.0001 \text{ mg/kg-d}) \times 1.63.$

 $= 0.000326 = 3.3 \times 10^{-4}$ .

1

2 3 However, the ADAFs do not factor out when the dose is not constant for each age in the age-specific 4 calculation. For example, in Example 2 (part b) in Section 6.1 of EPA Guidelines, the calculation is as 5 follows: 6 7 b. If exposure varies with age, then such differences are also included. Now suppose the same 8 example as immediately above, except that exposure for ages 1 through <12 was twice as high as 9 exposure for all other ages. In this case, sum the risk associated with each of the five relevant 10 time periods in which exposure rates and/or potencies (slope factors) vary: 11 12 *Risk equals the sum of:* 13 14 • Risk for birth through < 1 yr (1yr) =(2 per mg/kg-d) x 10 (ADAF) x 0.0001 mg/kg-d x  $1yr/70yr = 0.3 \times 10^{-4}$ 15 16 17 • Risk for ages 1 through < 2 (1yr) = (2 per mg/kg-d) x 10 (ADAF) x 0.0002 mg/kg-d x  $1yr/70 yr = 0.6 x 10^{-4}$ 18 19 20 • Risk for ages 2 through < 12 (10yr) = $(2 \text{ per mg/kg-d}) \times 3 (ADAF) \times 0.0002 \text{ mg/kg-d} \times 10 \text{ yr}/70 \text{ yr} = 1.7 \times 10^{-4}$ 21 22 23 • Risk for ages 12 through < 16 (4yr) = $(2 \text{ per } mg/kg-d) \ge 3 (ADAF) \ge 0.0001 mg/kg-d \ge 4yr/70yr = 0.3 \ge 10^{-4}$ 24 25 26 • Risk for ages 16 until 70 years (55yr) = 27  $(2 \text{ per mg/kg-d}) \times 1 (ADAF) \times 0.0001 \text{ mg/kg-d} \times 55 \text{ yr}/70 \text{ yr} = 1.6 \times 10^{-4}$ 28  $Risk = 0.3 \times 10^{-4} + 0.6 \times 10^{-4} + 1.7 \times 10^{-4} + 0.3 \times 10^{-4} + 1.6 \times 10^{-4} = 4.5 \times 10^{-4}$ 29 30 31 Here, the dose x slope does not factor out of the above calculation – even though the slope is constant for 32 all ages (namely, 2 per mg/kg-d) -- since the dose is 0.0002 mg/kg-d for ages between 1 and 12 and 33 0.0001 mg/kg-d for ages <1 and ages 12 and above. If one calculated the risk without ADAFs, namely, 34 35 36 Risk equals the sum of: 37 38 • Risk for birth through < 1 yr (1yr) = 39  $(2 \text{ per mg/kg-d}) \ge 0.0001 \text{ mg/kg-d} \ge 1 \text{ yr}/70 \text{ yr} = 2.9 \ge 10^{-6}$ 40 • Risk for ages 1 through < 2 (1yr) =41  $(2 \text{ per mg/kg-d}) \ge 0.0002 \text{ mg/kg-d} \ge 1 \text{ yr}/70 \text{ yr} = 5.7 \ge 10^{-6}$ 42 43

1	• Risk for ages 2 through $< 12 (10 \text{yr}) =$
2	$(2 \text{ per mg/kg-d}) \ge 0.0002 \text{ mg/kg-d} \ge 10 \text{ yr}/70 \text{ yr} = 5.7 \ge 10^{-5}$
3	$(2 \text{ per mg/kg-u}) \times 0.0002 \text{ mg/kg-u} \times 1001700 \text{ mg/kg-u} = 5.7 \times 100000000000000000000000000000000000$
4	• Risk for ages 12 through < 16 (4yr) =
5	$(2 \text{ per mg/kg-d}) \ge 0.0001 \text{ mg/kg-d} \ge 4 \text{ yr}/70 \text{ yr} = 1.1 \ge 10^{-5}$
6	
7	• Risk for ages 16 until 70 years (55yr) =
8	$(2 \text{ per mg/kg-d}) \ge 0.0001 \text{ mg/kg-d} \ge 55 \text{yr}/70 \text{yr} = 1.6 \ge 10^{-4}$
9	
10	$Risk = 2.9 \times 10^{-6} + 5.7 \times 10^{-6} + 5.7 \times 10^{-5} + 1.1 \times 10^{-5} + 1.6 \times 10^{-4}$
11	$= 2.4 \times 10^{-4}$
12	
13	and then multiplied this sum by a weighted average adjustment factor here,
14	$ADAF = [(10 \times 2 \text{ years}) + (3 \times 13 \text{ years}) + (1 \times 55 \text{ years})]/70 \text{ years} = 1.63 - 1.63$
15	the result would be
16	
17	$(2.4 \times 10^{-4}) \times 1.63 = 3.9 \times 10^{-4}$
18	
19	and not $4.5 \ge 10^{-4}$ .
20	
21	Except for the trivial case in which the exposure concentration is only non-zero in the first year,
22	cumulative exposure changes from year to year and is not constant throughout the period included in the
23	excess risk calculation. Hence, when dose is cumulative exposure, the ADAFs do not factor out of the
24	excess risk calculation and the risk can NOT be calculated as "risk without ADAFs" times a weighted
25	average adjustment factor.
26	
27	
28	5. Age-Dependent Adjustment Factor (ADAF): An Implementation When the Dose Metric is
29	Cumulative Exposure That Is Consistent with EPA Guidelines
30	An implementation that is consistent with EDA suidelines when the days matric is successful time
31 32	An implementation that is consistent with EPA guidelines when the dose metric is cumulative exposure
	would be to calculate the excess risk as in Example 2 (part b) in Section 6.1 of EPA Guidelines. That is,
33 34	calculate the excess risk in each year using the age-specific dose (cumulative dose) for that year and multiplying the slope by the age specific ADAE for that year (age). This would be consistent with EPA's
34 35	multiplying the slope by the age-specific ADAF for that year (age). This would be consistent with EPA's Guidelines from the point of view of both the excess risk calculation being done using age-specific
55	Guidelines from the point of view of both the excess fisk calculation being done using age-specific

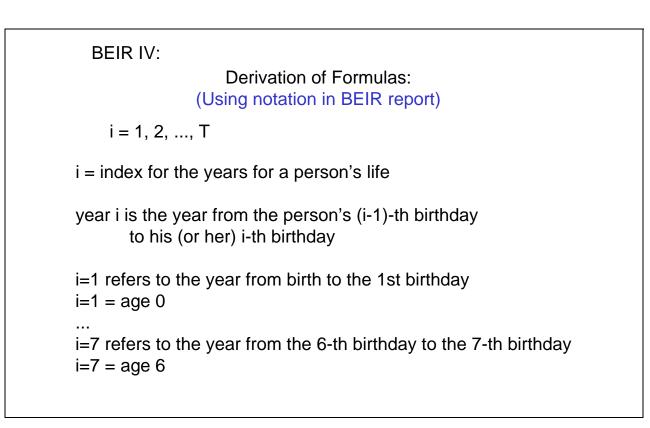
using age-specific 35 exposures and also the ADAFs being age-specific modifiers of the slope (potency). This implementation 36

37 of the ADAF is NOT equivalent to computing the excess risk by first calculating the excess risk without

any ADAFs and then multiplying this excess risk by a weighted average of the age-specific ADAFs over 38

- 39 the lifetime.
- 40

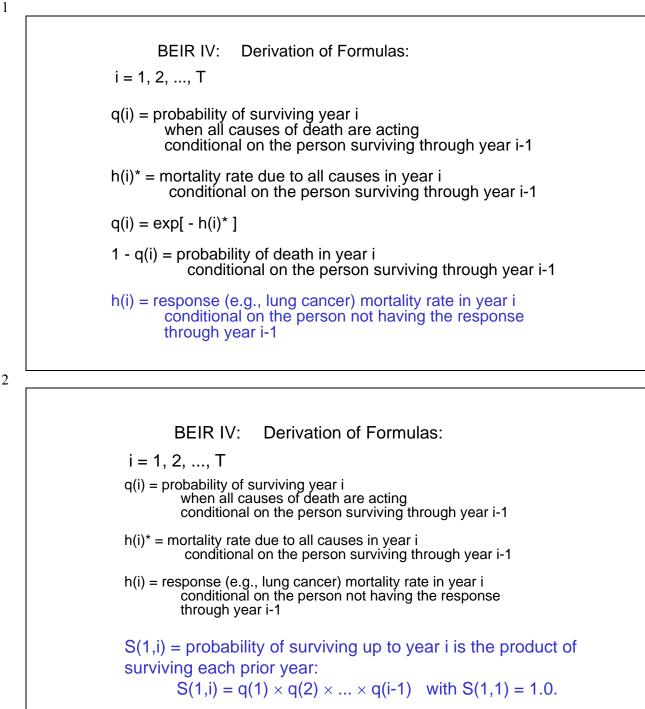
1 **APPENDIX 7. CALCULATING EXCESS RISK WHEN** 2 SPECIFIED RESPONSE IS MORTALITY VERSUS INCIDENCE 3 4 5 **Issues in Quantitative Epidemiology** 6 **Calculating Excess Risk When Specified Response is Mortality** 7 Vs When the Specified Response is Incidence 8 9 Robert L. Sielken Jr., Ph.D. 10 Ciriaco Valdez-Flores, Ph.D., P.E. Sielken & Associates Consulting, Inc. 11 3833 Texas Avenue, Suite, 230, Bryan, TX 77802 12 Tel: 979-846-5175; Fax: 979-846-2671; Email: SielkenAssoc@aol.com 13 14 15 January 17, 2007 16 17 **TCEQ Contract 582-7-81521** 18 19 The BEIR IV methodology for calculating excess risk is mathematically correct when the specified 20 response is mortality; however, the BEIR IV methodology is mathematically incorrect when the specified 21 response is incidence (not death). 22 23 The following slides are divided into two presentations. The first presentation provides a step-by-step 24 derivation of the BEIR IV methodology when the specified response is mortality. This presentation 25 directly parallels the same derivation in BEIR IV. The second presentation provides a step-by-step 26 derivation that is "parallel" to that in the first presentation except that in the second presentation the 27 specified response is incidence (not death). However, the steps and result are fundamentally different 28 when the specified response is incidence (not death) than when the response is death. 29 30 The fact that the "result" (i.e., the mathematical formula for calculating excess risk) is different when the response is mortality than it is when the response is incidence, means that when the response is incidence 31 32 (not death) the excess risk cannot be validly calculated using the formula (BEIR IV methodology) for 33 death. 34 35 The First Presentation: Issues in Quantitative Epidemiology: Calculating **Excess Risk: When Specified Response is Mortality** 36 37 38 Calculating Excess Risk using Actuarial Method or Life Table Method. This way of calculating excess 39 risks from a RR function is the implementation of the methodology described in "BEIR IV. Health Risks 40 of Radon and Other Internally Deposited Alpha-Emitters. Committee on the Biological Effects of 41 Ionizing Radiations. Board on Radiation Effects Research Commission of Life Sciences. National 42 Research Council. National Academy Press, Washington, DC, 1988." 43 44 45



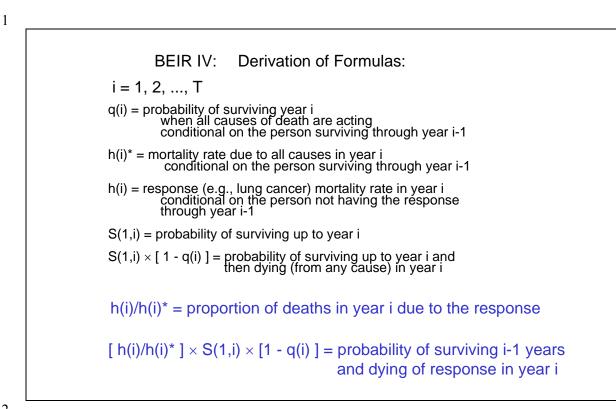
## 2

1

BEIR IV:	Derivation of Formulas:
i = 1, 2,, T	
	of surviving year i auses of death are acting I on the person surviving through year i -1
	of reaching a person 's 7-th birthday t he reached his 6 -th birthday
q(7) = P( Death $\ge$	7   Death $\geq$ 6 )
	te due to all causes in year i al on the person surviving through year i -1
q(i) = exp[ - h(i)* ]	
	ty of death in year i onal on the person surviving through year i -1



 $S(1,i) \times [1 - q(i)] =$  probability of surviving up to year i and then dying (from any cause) in year i



2

**BEIR IV:** Derivation of Formulas: i = 1, 2, ..., T q(i) = probability of surviving year i when all causes of death are acting conditional on the person surviving through year i-1 h(i)\* = mortality rate due to all causes in year i conditional on the person surviving through year i-1 h(i) = response (e.g., lung cancer) mortality rate in year i conditional on the person not having the response through year i-1 S(1,i) = probability of surviving up to year i = q(1) × q(2) × ... × q(i - 1)  $S(1,i) \times [1 - q(i)] =$  probability of surviving up to year i and then dying (from any cause) in year i  $h(i)/h(i)^* =$  proportion of deaths in year i due to the response  $[h(i)/h(i)^*] \times S(1,i) \times [1 - q(i)] =$  probability of surviving i-1 years and dying of response in year i  $R_0 = \sum_{i=1,...,T} [h(i)/h(i)^*] \times S(1,i) \times [1 - q(i)]$ = probability of a response mortality in the first T years of life (i.e., up to the T-th birthday, age T) at dose 0 (no exposure in addition to background exposure)

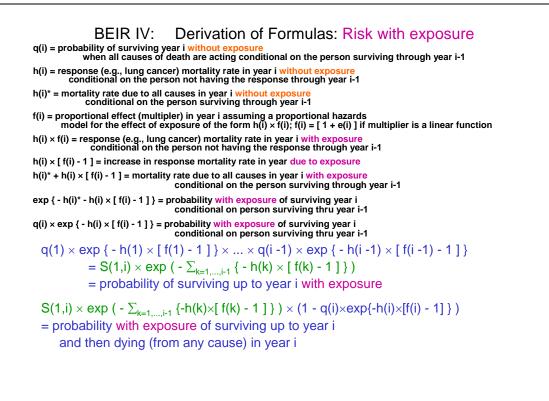
1

BEIR IV: Derivation of Formulas: Risk with exposure i=1, 2,, T
q(i) = probability of surviving year i <mark>without exposure</mark> when all causes of death are acting conditional on the person surviving through year i-1
h(i) = response (e.g., lung cancer) mortality rate in year i without exposure conditional on the person not having the response through year i-1
h(i)* = mortality rate due to all causes in year i without exposure conditional on the person surviving through year i-1
$\begin{array}{l} f(i) = \text{proportional effect (multipler) in year i assuming a proportional hazards} \\ \text{model for the effect of exposure of the form } h(i) \times f(i) \\ f(i) = [1 + e(i)] \text{ if the multiplier is a linear function} \end{array}$
h(i) × f(i) = response (e.g., lung cancer) mortality rate in year i with exposure conditional on the person not having the response through year i-1
$h(i) \times [f(i) - 1] = increase in response mortality rate in year due to exposure$

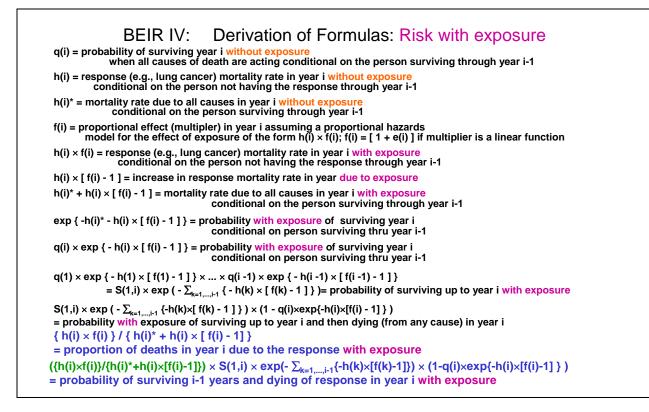
BEIR IV: Derivation of Formulas: Risk with exposure i=1, 2, ..., T q(i) = probability of surviving year i without exposure when all causes of death are acting conditional on the person surviving through year i-1 h(i) = response (e.g., lung cancer) mortality rate in year i without exposure conditional on the person not having the response through year i-1 h(i)\* = mortality rate due to all causes in year i without exposure conditional on the person surviving through year i-1 f(i) = proportional effect (multipler) in year i assuming a proportional hazards model for the effect of exposure of the form  $h(i) \times f(i)$ f(i) = [1 + e(i)] if multiplier is a linear function  $h(i) \times f(i)$  = response (e.g., lung cancer) mortality rate in year i with exposure conditional on the person not having the response through year i-1  $h(i) \times [f(i) - 1] =$  increase in response mortality rate in year due to exposure  $h(i)^* + h(i) \times [f(i) - 1] = mortality rate due to all causes in year i with exposure$ conditional on the person surviving through year i-1

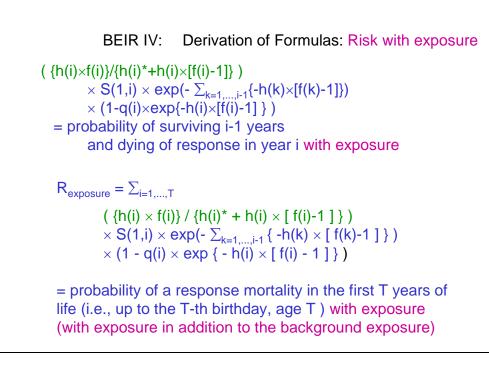
BEIR IV: Derivation of Formulas: Risk with exposure i=1, 2, ..., T q(i) = probability of surviving year i without exposure when all causes of death are acting conditional on the person surviving through year i-1 h(i) = response (e.g., lung cancer) mortality rate in year i without exposure conditional on the person not having the response through year i-1 h(i)\* = mortality rate due to all causes in year i without exposure conditional on the person surviving through year i-1 f(i) = proportional effect (multipler) in year i assuming a proportional hazards model for the effect of exposure of the form  $h(i) \times f(i)$ f(i) = [1 + e(i)] if multiplier is a linear function h(i) × f(i) = response (e.g., lung cancer) mortality rate in year i with exposure conditional on the person not having the response through year i-1  $h(i) \times [f(i) - 1] = increase in response mortality rate in year i due to exposure$ h(i)\* + h(i) × [f(i) - 1] = mortality rate due to all causes in year i with exposure conditional on the person surviving through year i-1 exp { -  $h(i)^*$  -  $h(i) \times [f(i) - 1]$  } = probability with exposure of surviving year i conditional on person surviving thru year i-1  $q(i) \times exp \{ -h(i) \times [f(i) - 1] \} = probability with exposure of surviving year i$ conditional on person surviving thru year i-1

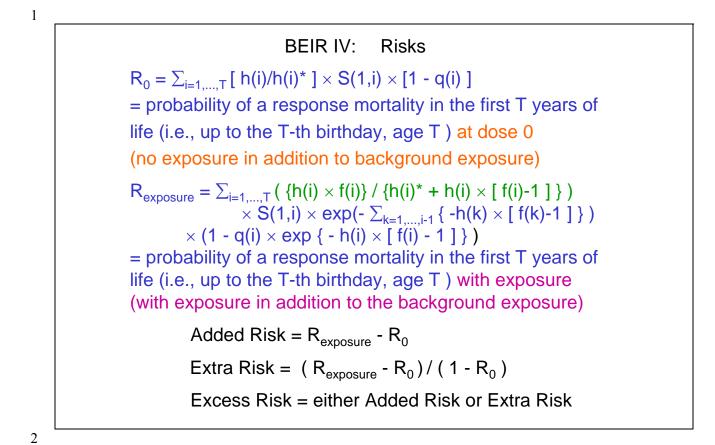
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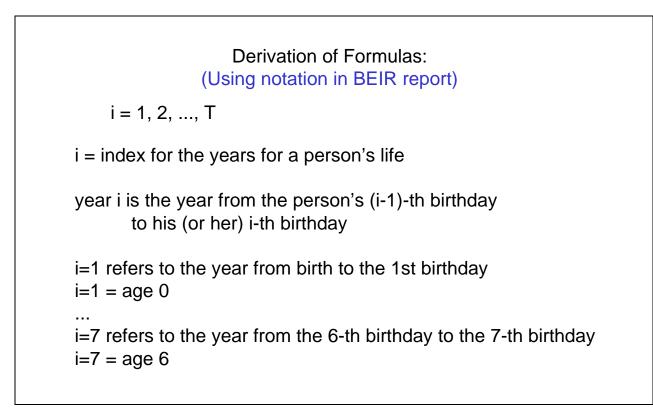




# The Second Presentation: 3.1 Issues in Quantitative Epidemiology: Calculating Excess Risk: When Specified Response is Incidence

Calculating Excess Risk using Actuarial Method or Life Table Method. The following derivation for the
situation in which the specified response is incidence (not death) "parallels" the derivation in BEIR IV;
however, the derivation and result are necessarily different for incidence than for mortality.

"BEIR IV. Health Risks of Radon and Other Internally Deposited Alpha-Emitters. Committee on the
Biological Effects of Ionizing Radiations. Board on Radiation Effects Research Commission of Life
Sciences. National Research Council. National Academy Press, Washington, DC, 1988."



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Z
_

i = 1, 2,, T
q(i) = probability of surviving year i when all causes of death are acting conditional on the person surviving through year i-1
q(7) = probability of reaching a person's 7-th birthday given that he reached his 6-th birthday
$q(7) = P(Death \ge 7   Death \ge 6)$
h(i)* = mortality rate due to all causes in year i conditional on the person surviving through year i-1
q(i) = exp[ - h(i)* ] definition of hazard rate
1 - q(i) = probability of death in year i conditional on the person surviving through year i1

Derivation of Formulas:
i = 1, 2, ..., T
q(i) = probability of surviving year i when all causes of death are acting conditional on the person surviving through year i-1
h(i)\* = mortality rate due to all causes in year i conditional on the person surviving through year i-1
q(i) = exp[ - h(i)\* ]
1 - q(i) = probability of death in year i conditional on the person surviving through year i-1
h(i) = response (e.g., lung cancer) incidence rate in year i conditional on the person not having the response through year i-1
Note that h(i) is NOT part of h(i)\*, because h(i) refers to incidence and h(i)\* refers to death.

Derivation of Formulas:
i = 1, 2, ..., T
q(i) = probability of surviving year i when all causes of death are acting conditional on the person surviving through year i-1
h(i)\* = mortality rate due to all causes in year i conditional on the person surviving through year i-1
q(i) = exp[ - h(i)\* ]
1 - q(i) = probability of death in year i conditional on the person surviving through year i-1
h(i) = response (e.g., lung cancer) incidence rate in year i conditional on the person hot having the response through year i-1
qr(i) = exp[ - h(i) ] = probability of no response in year i conditional on the person not responding through year i-1
1 -qr(i) = probability of response (incidence) in year i conditional on the person hot responding through year i-1

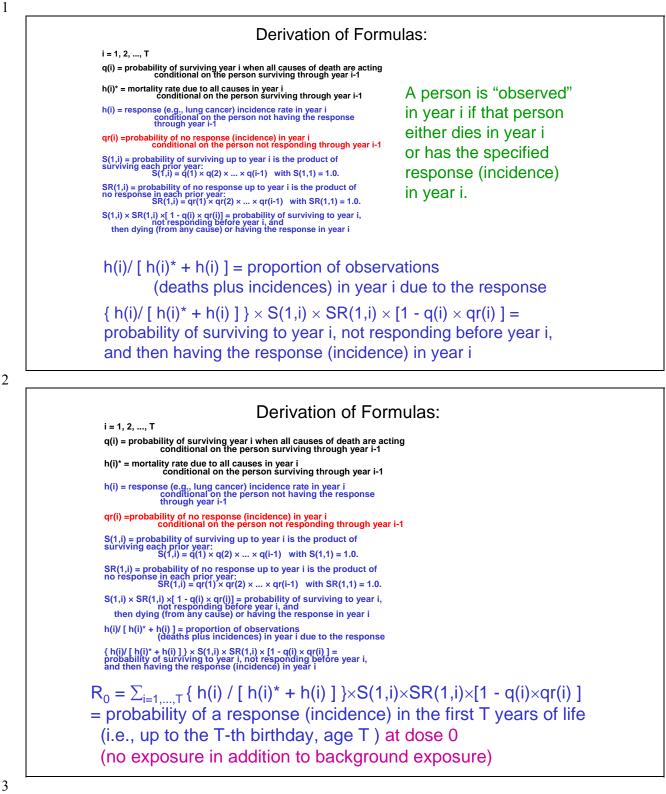
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	Derivation of Formulas:
i	i = 1, 2,, T
	q(i) = probability of surviving year i when all causes of death are acting conditional on the person surviving through year i-1
	h(i)* = mortality rate due to all causes in year i conditional on the person surviving through year i-1
	h(i) = response (e.g., lung cancer) incidence rate in year i conditional on the person not having the response through year i-1
	qr(i) =probability of no response (incidence) in year i conditional on the person not responding through year i-1
	S(1,i) = probability of surviving up to year i is the product of surviving each prior year: $S(1,i) = q(1) \times q(2) \times \times q(i-1)$ with $S(1,1) = 1.0$ .
	$SR(1,i) = probability of no response up to year i is the product ofno response in each prior year:SR(1,i) = qr(1) \times qr(2) \times \times qr(i-1) with SR(1,1) = 1.0.$

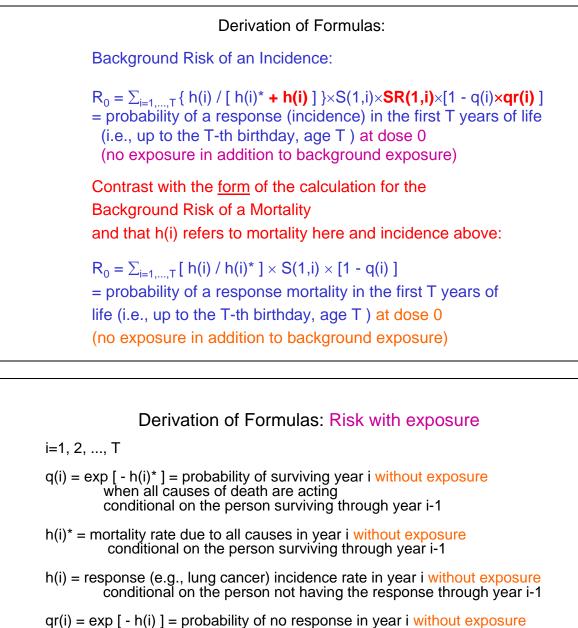
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# Derivation of Formulas:

i = 1, 2, ..., T q(i) = probability of surviving year i when all causes of death are acting conditional on the person surviving through year i-1 h(i)\* = mortality rate due to all causes in year i conditional on the person surviving through year i-1 h(i) = response (e.g., lung cancer) incidence rate in year i conditional on the person not having the response through year i-1 qr(i) = probability of no response (incidence) in year i conditional on the person not responding through year i-1 S(1,i) = probability of surviving up to year i is the product of surviving each prior year: S(1,i) = q(1) × q(2) × ... × q(i-1) with S(1,1) = 1.0. SR(1,i) = probability of no response up to year i is the product of no response in each prior year: SR(1,i) = qr(1) × qr(2) × ... × qr(i-1) with SR(1,1) = 1.0. S(1,i) × SR(1,i) × [1 - q(i) × qr(i)] = probability of surviving to year i, not responding before year i, and then dying (from any cause) or having the response in year i



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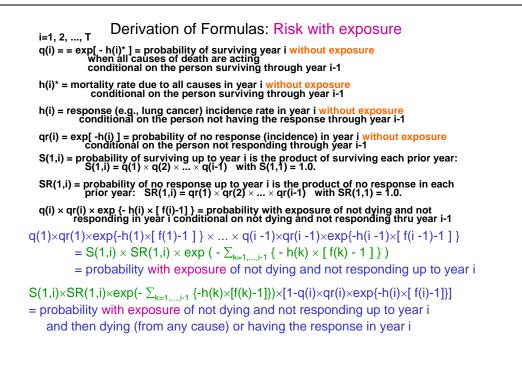
conditional on the person not responding through year i-1

 $\begin{array}{l} f(i) = \text{proportional effect (multipler) in year i assuming a proportional hazards} \\ \text{model for the effect of exposure of the form } h(i) \times f(i) \\ f(i) = [ \ 1 + e(i) \ ] \ \text{if the multiplier is a linear function} \end{array}$ 

 $h(i) \times f(i)$  = response (e.g., lung cancer) incidence rate in year i with exposure conditional on the person not having the response through year i-1

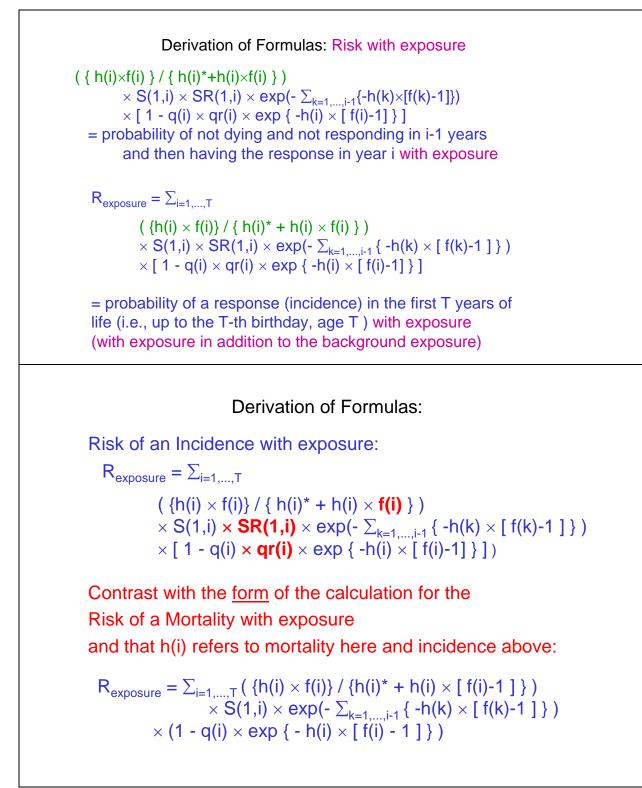
Derivation of Formulas: Risk with exposure	
i=1, 2,, T	
q(i) = exp [ - h(i)* ] = probability of surviving year i <mark>without exposure</mark> when all causes of death are acting conditional on the person surviving through year i-1	
h(i)* = mortality rate due to all causes in year i without exposure conditional on the person surviving through year i-1	
h(i) = response (e.g., lung cancer) incidence rate in year i without exposure conditional on the person not having the response through year i-1	
qr(i) = exp [ - h(i) ] = probability of no response (incidence) in year i without exposure conditional on the person not responding through year i-1	
f(i) = proportional effect (multipler) in year i assuming a proportional hazards model for the effect of exposure of the form h(i) × f(i) f(i) = [ 1 + e(i) ] if the multiplier is a linear function	
h(i) × f(i) = response (e.g., lung cancer) incidence rate in year i with exposure conditional on the person not having the response through year i-1	
A person is "observed" in year i if that person either dies in year i or has the specified response (incidence) in year i.	
h(i)* + h(i) × f(i) = observation rate due to all causes in year i with exposure conditional on the person not dying or having the response through year	
Derivation of Formulas: Risk with exposure	
i=1, 2,, T q(i) = = exp[ - h(i)* ] = probability of surviving year i without exposure when all causes of death are acting conditional on the person surviving through year i-1	
h(i)* = mortality rate due to all causes in year i without exposure conditional on the person surviving through year i-1	
h(i) = response (e.g., lung cancer) incidence rate in year i without exposure conditional on the person not having the response through year i-1	
qr(i) = exp[ -h(i) ] = probability of no response (incidence) in year i without exposure conditional on the person not responding through year i-1	
f(i) = proportional effect (multipler) in year i assuming a proportional hazards model for the effect of exposure of the form h(i) × f(i) f(i) = [ 1 + e(i) ] if the multiplier is a linear function	
h(i) × f(i) = response (e.g., lung cancer) incidence rate in year i with exposure conditional on the person not having the response through year i-1	
h(i)* + h(i) × f(i) = observation rate due to all causes in year i with exposure conditional on the person not dying or having the response through year i-1	
$\begin{array}{l} \exp \left\{ \begin{array}{l} -h(i)^{*} -h(i) \times f(i) \right\} =q(i) \times \exp \left\{ \begin{array}{l} -h(i) \times f(i) \right\} \\ =q(i) \times \exp \left\{ \begin{array}{l} -h(i) -h(i) \times [f(i) -1] \right\} =q(i) \times qr(i) \times exp \left\{ \begin{array}{l} -h(i) \times [f(i) -1] \end{array} \right\} \\ \text{probability with exposure of not dying and not} \end{array}$	}

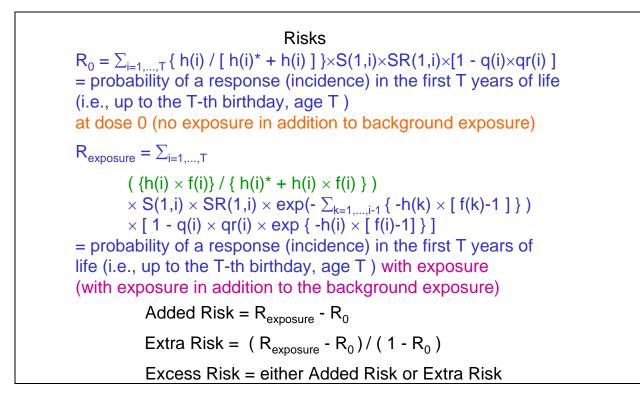
probability with exposure of not dying and not responding in year i conditional on not dying and not responding thru year i-1



### 1

Derivation of Formulas: Risk with exposure i=1, 2, ..., T q(i) = = exp[ - h(i)\* ] = probability of surviving year i without exposure when all causes of death are acting conditional on the person surviving through year i-1 h(i)\* = mortality rate due to all causes in year i without exposure conditional on the person surviving through year i-1 ure h(i) = response (e.g., lung cancer) incidence rate in year i without exposure conditional on the person not having the response through year i-1 qr(i) = exp[ -h(i) ] = probability of no response (incidence) in year i without exposure conditional on the person not responding through year i-1 S(1,i) = probability of surviving up to year i is the product of surviving each prior year:S(1,i) = q(1) × q(2) × ... × q(i-1) with S(1,1) = 1.0. $\begin{array}{l} {\sf SR}(1,i) = \mbox{priobability of no response up to year $i$ is the product of no response in each prior year:} $$ {\sf SR}(1,i) = qr(1) \times qr(2) \times ... \times qr(i-1) $$ with ${\sf SR}(1,1) = 1.0. $$ \end{array}$ q(i) × qr(i) × exp {- h(i) × [ f(i)-1] } = probability with exposure of not dying and not responding in year i conditional on not dying and not responding thru year i-1 S(1,i) × SR(1,i) × exp(-∑ k=1,...,i-1 {-h(k) ×[ f(k) - 1 ] }) × [1 - q(i) × qr(i) × exp{-h(i)×[ f(i)-1]} ] = probability with exposure of not dying and not responding up to year i and then dying (from any cause) or having the response in year i { h(i) × f(i) } / { h(i)\* + h(i) × f(i) } = proportion of observations (deaths plus incidences) in year i due to the response with exposure  $({h(i) \times f(i)}/{h(i)^{+}+h(i) \times f(i)}) \times S(1,i) \times SR(1,i) \times exp(-\sum_{k=1,...,i-1}{-h(k) \times [f(k)-1]})$  $\times$  [1-q(i)×qr(i)×exp{-h(i)×[f(i)-1]}] = probability with exposure of not dying and not responding up to year i and then having the response in year i





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